ANTIHYPERGLYCAEMIC ACTIVITIES OF XANTHONE RICH EXTRACT OF MANGOSTEEN (GARCINIA MANGOSTANA)

BY

TENGKU MUHAMAD FARIS SYAFIQ BIN TENGKU ZAKARIA

A thesis submitted in fulfilment of the requirement for the degree of Master in Pharmaceutical Science (Pharmaceutical Technology)

Kulliyyah of Pharmacy International Islamic University Malaysia

FEBRUARY 2015

ABSTRACT

Mangosteen (Garcinia mangostana Linn.) fruit pericarp has been used for centuries as a folk medicine. The study was conducted to evaluate the in vitro and in vivo antihyperglycaemic potential of G. mangostana extract (GME). The α-mangostin content in the extract was measured using HPLC and investigated for total phenolic and flavonoid contents. Antioxidant activities were measured by DPPH radical scavenging and reducing power assays whereas in vitro antidiabetic activities were evaluated by inhibition of α -glucosidase and α -amylase enzymes. Effects of GME on adipocyte cells were assessed through MTT assay, adipogenesis and glucose uptake measurements. In animal study, oral administration of GME1 (50 mg/kg), GME2 (100 mg/kg) and GME3 (200 mg/kg) to STZ-induced diabetic rats in single-dose (acute) and multiple-dose study (sub-acute) were examined. Serum biochemical parameters and histopathological alterations were evaluated and compared to standard hypoglycaemic drug, glibenclamide. The results showed that total phenolic and total flavonoid contents were 122.2±1.04 mg GAE/g and 72.8±1.75 mg QE/g of dry extract, respectively. DPPH radical scavenging activity and reducing power capacity reported with EC₅₀ of 48.2 µg/ml and IC₅₀ of 98.2 µg/ml, respectively. The enzymatic inhibition of α -glucosidase and α -amylase revealed higher percentage of inhibition comparable to acarbose with IC₅₀ of 0.41 and 0.24 mg/ml, respectively. In cellular study, cytotoxicity assay reported that dose of less than 12.5 µg/ml does not affect cell viability. The differentiations of adipocytes were increased with higher GME concentration at 2.5 µg/ml, 5.0 µg/ml and 10.0 µg/ml. Glucose uptake measurements revealed a higher uptake of 2-deoxyglucose in GME2-treated cell (2243.3±232.3 cpm) as compared to GME1 (1864.0±146.3 cpm) and GME3 (1246.0±155.8 cpm). Oral administration of GME on diabetic rats indicated safe usage with absence of behavioural alterations, autonomic, neurological and toxic effects up to 2000 mg/kg. The results showed a significant reduction of glucose level in GME2 and GME3 (p<0.001) as compared to GME1. Total cholesterol, serum triglyceride, urea and creatinine were reduced in the treatment group while total protein contents were increased. Histological assessment of livers and kidneys revealed reduced lesions whereas mild regenerative activity of β -cell was observed in pancreas of diabetic rats. In conclusion, the findings demonstrated that GME could be a potential source in diabetes management owing to its antioxidant content, delayed carbohydrate digestion, induction of adipocyte differentiation, improvement in glucose uptake and antihyperglycaemic effect in diabetic rats.

ملخص البحث

قشرة فاكهة المانغوستين (غارسينيا مانغوستانا لين.) تمّ استخدامها لقرون في الطبّ الشعبي. الدراسة تمّ إجراءها لتقييم إمكانية مستخلص ج مانغوستانا كما تمّ التحقيق في كمية الفينولات HPLC محتوى ألفا-مانغوستين في المستخلص تمّ قياسه باستخدام تقنية لرصد الجذر الحرّ DPPH والفلافونويد الإجمالية. الخصائص المضادة للأكسدة تمّ قياسها باستخدام تقنية وكذا فحص قوة الإرجاع بينما تمّ تقييم الأنشطة ضد السكري عن طريق كبت إنزيميّ ألفا-غلوكوزيداز. Adipogenesis ,MTT على الخلايا الدهنية تمّ تقييمه بواسطة تقنيات GME وألفا-أميلاز. تأثير مستخلص بتركيز 50 مغ/كغ GME1 وكذا قدرة امتصاص الغلوكوز. في الدراسة على الحيوان، تقديم مستخلص المستحثة بداء STZ بتركيز 200 مغ/كغ إلى فئران GME3 بتركيز 100 مغ/كغ ومستخلص GME2 السكري من خلال جرعة واحدة (حادة) وجرعات متعدّدة (شبه حادة) تمّت دراستها. معلمات المصل البيوكيميائية والتعديلات المرضية تمّ تقييمها ومقارنتها مع معيار الدواء المضاد للسكري غليبنكلامايد. / غ GAE النتائج أظهرت بأنّ كمية الفينولات والفلافونويدز الإجمالية كانت تعادل 1.04±122.2 مغ لرصد الجذر الحرّ وكذاDPPH/غرام IC50. من المستخلص الجاف على التوالي. تقنية QE و 472.8 ± 1.75 مغ بمقدار 98.2 مكغ/مل على التوالي بمقدار 48.2 مكغ/مل و EC_{50} قدرة قوة الإرجاع مع IC_{50} عملية كبت إنزيميّ ألفا–غلوكوزيداز وألفا–أميلاز أظهرت نسبة كبت عالية بالمقارنة مع أكاربوز ببمقدار 0.41 و 0.24 مغ/مل على التوالي. في الدراسة الخلوية، الفحص السمّى أظهر بأنّ جرعة أقل من 12.5 مكغ/مل لا تؤثر على قابلية حياة الخلايا. تفاضل الخلايا الدهنية كان في ارتفاع مع زيادة تركيز ال عند 2.5 مكغ/مل ، 5.0 مكغ/مل و10.0مكغ/مل. قياسات امتصاص الغلوكوز GME مستخلص GME2 في الخلايا التي تمت معالجتها بمستخلص 2-deoxyglucose أظهرت امتصاصا عاليا ل(GME1 ومستخلص 1864.0±146.3 cpm) ومستخلص GME1) ومستخلص (2243.3±232.3cpm) بالمقارنة مع مستخلص (إلى الفئران المصابة بالسكري أظهر GME تقديم مستخلص) GME3 1246.0±155.8 cpm استخداما آمنا مع غياب التعديلات السلوكية، اللاإرادية، العصبية والآثار السامة إلى غاية تركيز قدره GME3 و 2000GME2 مغ/كغ. أظهرت النتائج انخفاضا في معدلات الغلوكوز في مستخلصات. معدل الكوليسترول الكلّي، ثلاثيات الغليسريد في المصل GME1 بالمقارنة مع مستخلص (p<0.001) اليوريا وكذا الكرياتينين تمّ تخفيضها في المجموعة الضابطة بينما ارتفع معدّل البروتينات الكليّة. التقييم النسيجي للكبد والكلية أظهر انخفاضا في الأضرار حيث نشاط الإصلاح المعتدل لخلايا بيتا تمّت ملاحظتها في البنكرياس الخاص بالفئران المصابة بالسكري. قد يكون مصدرا محتملا في التعامل مع مرض السكري GME كخلاصة، النتائج أظهرت بأنّ مستخلص نتيجة احتواءه على مضادات الأكسدة، القدرة المؤجلة لهضم الكربوهيدرات، استقراء تفاضل الخلايا الدهنية، تحسين قدرة امتصاص الغلوكوز وكذا القدرة على تخفيض مستوى السكر عند الفئران المصابة بداء السكري.

ABSTRAK

Kulit manggis (Garcinia mangostana Linn.) telah terbukti sejak dahulu lagi digunakan dalam perubatan tradisional. Kajian ini bermatlamat mengkaji potensi ekstrak G. mangostana (GME) terhadap penurunan glukosa dalam darah secara tabung uji dan keatas haiwan. Kandungan α-mangostin dalam ekstrak diukur melalui HPLC and diselidiki jumlah kandungan fenol dan flavonoidnya. Aktiviti antioksida diukur melalui cerakin hapus sisa radikal DPPH dan cerakin upaya penurunan kuasa manakala aktiviti antidiabetik secara tabung uji dinilai berdasarkan perencatan enzim α-glukosida and α-amilas. Kesan GME terhadap sel adiposit dinilai melalui cerakin MTT, induksi adipogenesis dan pengukuran pengambilan glukosa. Dalam ujian keatas haiwan, dos GME1 (50 mg/kg), GME2 (100 mg/kg) dan GME3 (200 mg/kg) secara oral kepada tikus diabetes STZ teraruh melalui kajian secara dos tunggal (akut) dan dos pelbagai (sub-akut) dinilai. Parameter serum biokimia dan perubahan histopatologi diselidik dan dibanding dengan dengan ubat standard diabetes, glibenclamide. Keputusan menunjukkan jumlah fenol dan flavonoid masing-masing adalah 122.2±1.04 mg GAE/g dan 72.8±1.75 mg QE/g daripada ekstrak kering. Aktiviti hapus sisa radikal DPPH dan cerakin upaya penurunan kuasa masing-masing melaporkan nilai EC₅₀ 48.2 µg/ml dan IC₅₀ 98.2 µg/ml. Melalui cerakin antidiabetik secara tabung uji, enzim α-glukosida dan α-amilas menunjukkan peratus perencatan yang tinggi berbanding akarbos dengan nilai IC₅₀ masing-masing 0.41 dan 0.24 mg/ml. Dalam kajian sel, cerakin toksik sel menunjukkan dos kurang daripada 12.5 ug/ml tidak mempengaruhi hayat sel. Perubahan sel preadiposit kepada adiposit matang didapati bertambah dengan peningkatan konsentrasi GME pada 2.5 µg/ml, 5.0 μg/ml dan 10.0 μg/ml. Pengukuran pengambilan glukosa menunjukkan pengambilan 2-deoksiglukosa yang tinggi pada sel yang dirawat dengan GME2 (2243.3±232.3 cpm) berbanding GME1 (1864.0±146.3 cpm) dan GME3 (1246.0±155.8 cpm). Pemberian GME kepada tikus secara oral menunjukkan penggunaan ekstrak yang selamat dengan ketiadaan perubahan tingkahlaku, autonomik, neurologi dan kesan toksik sehingga dos 2000 mg/kg. Keputusan menunjukkan kadar penurunan glukosa yang signifikan dalam GME2 dan GME3 (p<0.001) berbanding GME1. Jumlah kolesterol, serum trigliserid, urea dan kreatinin didapati menurun dalam kumpulan yang dirawat sementara jumlah kandungan protin didapati bertambah. Penilaian histologi menunjukkan aktiviti regenerasi yang ringan pada sel β dalam tikus diabetes. Kesimpulannya, penemuan ini menunjukkan bahawa GME berupaya menjadi satu sumber berpotensi dalam mengatur diabetes merujuk kepada kandungan berupaya melambatkan proses antioksidanya, penghadaman karbohidrat, menggalakkan proses adipogenesis, pemulihan pengambilan glukosa serta kesan anti peningkatan glukosa dalam darah terhadap tikus diabetes.

APPROVAL PAGE

I certify that I have supervised and read this study to acceptable standards of scholarly presentation a quality, as a thesis for the degree of Ma (Pharmaceutical Technology)	and is fully adequate, in scope and
	Muhammad Taher Supervisor
I certify that I have read this study and that in my standards of scholarly presentation and is fully ad thesis for the degree of Master of Pharmac Technology)	lequate, in scope and quality, as a
	Abdul Razak Kasmuri Examiner
I certify that I have read this study and that in my standards of scholarly presentation and is fully ad thesis for the degree of Master of Pharmac Technology)	lequate, in scope and quality, as a
	Zainul Amiruddin Zakaria External Examiner
This thesis was submitted to the Department of laccepted as a fulfilment of the requirements Pharmaceutical Science (Pharmaceutical Technolog	for the degree of Master of
	Juliana Md Jaffri Head, Department of Pharmaceutical Technology
This thesis was submitted to the Department of laccepted as a fulfilment of the requirements Pharmaceutical Science (Pharmaceutical Technolog	for the degree of Master of
	Siti Hadijah Shamsudin Dean, Kulliyyah of Pharmacy

DECLARATION

I hereby declare that this thesis is the result of my ow	on investigation, except where		
otherwise stated. I also declare that it has not bee	n previously or concurrently		
submitted as a whole for any other degrees at IIUM or o	ther institutions.		
Tengku Muhamad Faris Syafiq bin Tengku Zakaria			
Signature	Date		

INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

DECLARATION OF COPYRIGHT AND AFFIRMATION OF FAIR USE OF UNPUBLISHED RESEARCH

Copyright ©2015 by Tengku Muhamad Faris Syafiq bin Tengku Zakaria. All rights reserved.

ANTIHYPERGLYCAEMIC ACTIVITIES OF XANTHONE RICH EXTRACT OF MANGOSTEEN (GARCINIA MANGOSTANA)

No part of this unpublished research may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder except as provided below.

- 1. Any material contained in or derived from this unpublished research may be used by others in their writing with due acknowledgement.
- 2. IIUM or its library will have the right to make and transmit copies (print or electronic) for institutional and academic purposes.
- 3. The IIUM library will have the right to make, store in a retrieval system and supply copies of this unpublished research if requested by other universities and research libraries.

Affirmed by Tengku Munamad Faris Syafiq bin Tengku Zakaria		
Date		

ACKNOWLEDGEMENTS

I wish to express my gratitude to Allah s.w.t for granting me this priceless experience throughout my life seeking for knowledge. My deepest appreciation and gratitude to those who directly or indirectly helped me along this blessed journey. It was highly pleasurable to express my thanks to my supervisor, Assoc. Prof. Muhammad Taher, who taught me much lessons and knowledge always had the time to share and listen to my ideas and indirectly taught me importance of time, research and academia. Your advice and comments will be never forgotten. To my beloved mother, Khasiaah bt Yaacob, my deepest love for her prayers, advice, moral support, and the one who kept me motivated during my hard times. My deepest gratitude to my siblings and family members for their understanding and constant encouragement. To Assoc. Prof. Deny Susanti for giving me the chance to pursue my master degree albeit great obstacles. Also, I wish to express my sincere and warm and appreciation to my labmates, Dedi Noviendri, Anugerah Adina, Putri Nur Hidayah, Huwaida and Afnan for your kind advice, emotional support and continuous suggestion to improve my project progression. Attached undergraduates, Hanisuhana, Amnani, Ichah, Haifaa, Farah, Syamil, Azizah, Shak, Fatim, with mind-blowing questions to keep me aware of theoretical importance before conducting lab works. I am perpetually grateful to fellow colleagues who constantly encouraging me, Fahmi, Monem, Izzati, Anas, Hafizah, Fathin, Azad, Husam, Khairul, Shariff, Azmir, Fahim, Mahmood and all postgraduates for unceasing support, cooperation and friendship that will be treasured. I feel opportunistic to acknowledge administrative staffs, Sis Haslina for keeping me updated of latest news regarding postgraduate study; Department of Pharmaceutical Technology staffs, Sis Yanti, Br Dzadil, Sis Mimi for immediate action regarding instrument handling and lab apparatus request; Science officer, Sis Zaililah, Sis Rusianti and Sis Wahida, for great feedback of any problems; Sis Seri, Sis Ayu, Sis Salmi, Br Faris, Br Hanif, Br Zack, Br Adi for inter-departmental lab access and instrumentation. Not to forget, En Azhar for his time taking me to conferences and sampling purposes; Br Razif for voucher specimen preparation; Hj Karim for sample processing; En Kufli for his help on animal handling and supply; En Zul from Forestry Department Kuantan for license of sampling activity; Dr Abd Kadir, En Kamaruzaman and Sis Hayani at Malaysian Nuclear Agency for great assistance and all who are directly or indirectly helped me through my study. Lastly, I would like to thank International Islamic University Malaysia, Ministry of Education and Civil Service Department for providing me the scholarship. The research was supported by eScience MOSTI Research Grant: 02-01-08-SF 0110.

TABLE OF CONTENTS

Abstract	ii
Abstract in Arabic	iii
Abstract in Bahasa Malaysia	iv
Approval Page	iv
Declaration Page	vi
Copyright Page	vii
Acknowledgements	. viii
List of Table	.xiii
List of Figures	. xiv
List of Equations	xvii
List of Abbreviations	xviii
CHAPTER 1: INTRODUCTION	
1.1 General Overview	
1.2 Significance of the Study	
1.3 General Objectives of the Study	
1.4 Research Problem	4
1.5 Hypothesis	5
1.6 Experimental Design	6
CHAPTER 2: LITERATURE REVIEW	7
2.1.1 Drug Discovery and Natural Products	
2.1.2 Medicinal Plants Research in Malaysia	
2.2 Diabetes mellitus	
2.2.1 Prevalence of Diabetes in Malaysia	
2.2.2 Classification of Diabetes	
2.2.3 Mechanisms Associating Obesity to Insulin Resistance and Type Diabetes	
2.2.4 Hyperglycaemia-Induced Oxidative Stress and Role of Antioxida	
	15
2.2.5 Oral Hypoglycaemic Agents	17
2.2.6 Carbohydrate Digestion	18
2.2.7 Alpha-glucosidase Inhibition: Mode of Action	19
2.2.8 Herbal Medicines for Diabetes Management	
2.2.9 Insulin: Mechanism of Action	
2.2.10 GLUT4 Translocation	23
2.3 Garcinia mangostana	
2.3.1 Guttiferae Family	24
2.3.2 Garcinia Species	
2.5.2 Garcina Species	24
	24
2.3.3 Botanical Description G. mangostana Linn	24 24 25
	24 24 25 28

2.4 Cellular and Molecular Regulation of Adipogenesis	31
2.4.1 3T3-L1 Preadipocytes	31
2.4.2 Adipocyte Differentiation	33
2.4.3 Transcriptional Regulation of Adipogenesis	34
2.4.4 PPARγ: Mechanism of Action and Their Role	
2.5 Animal Model for Study of Antihyperglycaemic Effect	
2.5.1 Animal Model of Hyperglycaemia	
2.5.2 Alloxan and Streptozotocin-induced Diabetes	
2.5.3 Bioavailability of Polyphenols	
2.5.4 Pharmacokinetics of α-, γ- mangostin and GME	
2.5.5 Toxicity of G. mangostana	
2.5.6 Ethical Concern in Animal Handling	42
CHAPTER 3: PHYTOCHEMICAL ANALYSIS, ANTIOXIDANT	AND
ENZYMATIC INHIBITION STUDIES	
3.1 Specific Objectives	
3.2 Materials and Methods	
3.2.1 Chemicals	
3.2.2 Plant Material	
3.2.3 Extraction	
3.2.4 Phytochemical Screening	
3.2.4.1 Test for Phenols	
3.2.4.2 Test for Flavonoids	
3.2.4.3 Test for Tannins	
3.2.4.4 Test for Terpenoids	
3.2.4.5 Test for Anthocyanin	
3.2.4.6 Test for Glycosides	
3.2.4.7 Test for Combined Anthraquinones	
3.2.4.8 Test for Alkaloids	
3.2.4.9 Test for Saponins	
3.2.4.10 Test for Reducing sugars	
3.2.5 Phytochemical Analysis	
3.2.5.1 Thin Layer Chromatography (TLC)	
3.2.5.2 UV Spectrophotometry	
3.2.5.3 High Performance Liquid Chromatography (HPLC)	
3.2.6 Determination of Phenolic and Flavonoid Contents	
3.2.6.1 Total Phenolic Content (TPC)	
3.2.6.2 Total Flavonoids Content (TFC)	
3.2.7 Determination of Antioxidant Activities	
3.2.7.1 2,2-diphenyl-1-picrylhydrazyl (DPPH) Radical Scaveng	
Activity	
3.2.7.2 Reducing Power Capacity	
3.2.8 Enzymatic Inhibition Study	
3.2.8.1 α-Glucosidase Inhibition Assay	
3.2.8.2 α-Amylase Inhibition Assay	
3.2.9 Statistical Analysis	
3.3 Results and Discussion	56
3.3.1 Phytochemical Analysis	56

3.3.1.1 Phytochemical Screening	56
3.3.1.2 Thin Layer Chromatography	58
3.3.1.3 Ultraviolet Spectrophotometry	
3.3.1.4 HPLC Analysis	
3.3.2 Total Phenolic and Flavonoid Contents (TPC and TFC)	
3.3.3 Antioxidant Activities	
3.3.3.1 DPPH Radical Scavenging Activity	
3.3.3.2 Reducing Power Capacity	
3.3.4 Enzymatic Inhibition Study	
3.3.4.1 α-Glucosidase and α-Amylase Inhibitory Activites 3.4 Summary of Results	
CHAPTER 4: EFFECTS OF GME ON 3T3-L1 ADIPOCYTES	73
4.1 Specific Objectives	73
4.2 Materials and Methods	73
4.2.1 General	73
4.2.2 Cell Culture	
4.2.2.1 3T3-L1 Preadipocytes	
4.2.2.2 Cell Thawing	
4.2.2.3 Cell Counting	
4.2.2.4 Subculture/ Cell Splitting	
4.2.2.5 Trypsinization	
4.2.2.6 Cell Storage	
4.2.2.8 Adipocyte Differentiation (Adipogenesis)	
4.2.2.9 Staining with Oil-Red-O (ORO)	
4.2.2.10 ORO Quantification	
4.2.2.11 2-Deoxy-[³ H]-D-Glucose Uptake Assay	
4.2.3 Statistical Analysis	
4.3 Results and Discussion	
4.3.1 3T3-L1 Preadipocytes Growth Pattern	82
4.3.2 Effect of GME on Cell Viability (Cytotoxic Assay)	
4.3.3 Effect of GME on Lipid Accumulation (Adipogenesis)	
4.3.4 Effect of GME on Glucose Uptake Regulation	93
CHAPTER 5: EFFECTS OF GME ON STREPTOZOTOCIN-IN DIABETIC RATS	
5.1 Specific Objectives	
5.2 Materials and Methods	
5.2.1 Chemicals	
5.2.2 Animal Study	
5.2.2.1 Experimental Animals	
5.2.2.2 Experimental Design	
5.2.2.3 Single-dose Study	100
5.2.2.4 Multiple-dose Study	
5.2.2.5 Toxicity Study	
5.2.3 Methods for Blood Preparation	
5.2.3.1 Collection of Blood	
7 / 3 / Serum Biochemical Analysis	[117]

5.2.4 Methods for Histopathological Assessment	103
5.2.4.1 Tissue Processing	103
5.2.4.2 Haematoxylin & Eosin Staining	104
5.2.5 Statistical Analysis	104
5.3 Results and Discussion	105
5.3.1 Single-dose Study	105
5.3.1.1 Effect of GME on Normoglycaemic Rats	105
5.3.1.2 Effect of GME on STZ-induced Diabetic Rats	
5.3.2 Multiple-dose Study	
5.2.3.1 Effect of GME on Blood Glucose and Body Weight I	n STZ-
induced Diabetic Rats	106
5.3.3 Toxicity Study	
5.3.4 Effect of GME's on Serum Biochemical Index	
5.3.5 Effect of GME's on Histopathological Morphology	118
5.3.5.1 Liver Histology	
5.3.5.2 Kidney Histology	
5.3.5.3 Pancreas Histology	
5.4 Summary of Results	
CHAPTER 6: CONCLUSION	
6.1 General	
6.2 Limitations	
6.3 Recommendations for Future Works	131
REFERENCES	132
ADDENIDIV I. Evangimental in vitue Asserva	150
APPENDIX I: Experimental <i>in vitro</i> Assays	
APPENDIX II: Sprague-Dawley Rats After 28-Days of Treatment	
APPENDIX IV: UV Spectrophotometry Report of α-mangostin	
APPENDIX V: UV Spectrophotometry Report of GME	
APPENDIX VI: HPLC Analysis Report of GME	
APPENDIX VIII: Animal Ethics Approval	139

LIST OF TABLES

Table No.		Page No
Table 2.1	Oral antidiabetic agents with their mode of actions	17
Table 2.2	Plants used as herbal medicine in diabetic management	21
Table 2.3	Traditional medical use of G. mangostana	27
Table 2.4	Pharmacological effects of G. mangostana in vitro	27
Table 2.5	Chemical constituents of mangosteen pericarp	31
Table 2.6	Comparison of adipogenic cell models in proliferation and differentiation capacity	33
Table 2.7	Differences between widely used cytotoxic glucose analogues	39
Table 2.8	Several effects of polyphenolics	40
Table 3.1	Indications of GME's phytochemical screening results	57
Table 3.2	Total phenolic and flavonoids of <i>G. mangostana</i> pericarp extracts	63
Table 4.1	The seeding densities of ATCC® CL-173 $^{\text{TM}}$ (3T3-L1) cell line	76
Table 4.2	Overview of differentiation of 3T3-L1 adipocytes	79
Table 4.3	Inducer cocktail of adipocyte differentiation	80

LIST OF FIGURES

Figure No.		Page No
Figure 1	Flow chart of the study	6
Figure 2.1	Number of drugs approved in the United States from 1981 to 2007	8
Figure 2.2	Mechanisms of β -cell adaptation to insulin resistance	14
Figure 2.3	Impaired insulin release in linking obesity with insulin resistance and type 2 diabetes	14
Figure 2.4	Mechanism of hyperglycemia-induced cellular damage mediated by oxidative stress	15
Figure 2.5	Hyperglycemia-induced pathways of oxidative stress in obesity, including the polyol pathway, the AGE pathway and glucose auto-oxidation	16
Figure 2.6	Actions of insulin	22
Figure 2.7	Uptake of glucose to intracellular compartments following insulin binding initiating signaling cascade	23
Figure 2.8	Some useful G. mangostana plant parts	26
Figure 2.9	Some of xanthones isolated from G. mangostana	30
Figure 2.10	Adipocytes development from MSCs through two stages of maturation	32
Figure 2.11	Adipocytes secrete proteins with multiple effects on glucose homeostasis	32
Figure 2.12	Molecular pathways of transcriptional events mediating adipocyte differentiation	35
Figure 3.1	A) Fresh mangosteen fruit; B) Dried mangosteen pericarps and (C) pulverized form before maceration	46
Figure 3.2	Standard curve for TPC calculation	51
Figure 3.3	Standard curve for TFC calculation	52
Figure 3.4	TLC chromatogram of GME and α-mangostin	59

Figure 3.5	Comparisons between physical appearances of GME and α -mangostin	59
Figure 3.6	Combined UV spectra of GME (thick line) and α -mangostin (thin line)	60
Figure 3.7	Standard calibration curve of α -mangostin	61
Figure 3.8	HPLC chromatogram of α -mangostin (standard)	62
Figure 3.9	HPLC chromatogram of GME (sample)	62
Figure 3.10	Percentage inhibition (I%) of GME as measured at 517 nm	65
Figure 3.11	Reducing power activities of GME as measured at 700 nm	67
Figure 3.12	Percent inhibition (%) of enzymes at GME concentration of 0.5 mg/ml	68
Figure 3.13	Summary of phytochemical screening, antioxidant activities and enzymatic inhibition of GME	72
Figure 4.1	3T3-L1 preadipocytes growth curves	83
Figure 4.2	Morphology of 3T3-L1 preadipocytes	84
Figure 4.3	Percentage viability of 3T3-L1 preadipocytes	85
Figure 4.4	MTT assay	85
Figure 4.5	Effect of GME on adipocyte differentiation	87
Figure 4.6	Effect of GME on adipocyte differentiation after 4 days following treatment with MDI cocktails and insulin	89
Figure 4.7	Effect of GME on adipocyte differentiation after 8 days following treatment with MDI cocktails and insulin	90
Figure 4.8	Lipid content was stained with Oil-Red-O after 8 days of treatment with GME	91
Figure 4.9	Effect of GME on glucose uptake of 3T3-L1 adipocyte	94
Figure 5.1	Effects of GME on glucose level of normoglycaemic rats	105
Figure 5.2	Effects of GME on glucose level of STZ-induced diabetic rats	106
Figure 5.3	Effects of GME on serum glucose level in diabetic rats for multiple-dose study	107

Figure 5.4	Effects of GME on body weight in diabetic rats for 28 days of treatment	108
Figure 5.5	Comparison between serum glucose level in first and the fourth week	108
Figure 5.6	Comparison between body weight in first and the fourth week	109
Figure 5.7	Concentration of serum lipid profiles after 28-days of GME treatment	115
Figure 5.8	Concentration of serum liver profiles after 28-days of GME treatment	116
Figure 5.9	Concentration of serum urea profiles after 28-days of GME treatment	116
Figure 5.10	Concentration of serum creatinine levels after 28-days of GME treatment	117
Figure 5.11	Concentration of total protein levels after 28-days of GME treatment	117
Figure 5.12	Photograph of histological section of livers	120
Figure 5.13	Photograph of histological section of kidneys	122
Figure 5.14	Photograph of histological section of pancreatic islets	124
Figure 5.15	Summary of GME effects on cell culture and animal study	125

LIST OF EQUATIONS

Equations No.		Page No.
Equation 3.1	$R_f = \underline{\text{Distance travelled by compound}}$ Distance travelled by solvent	49
Equation 3.2	Scavenging activity (%) = $[(A_{control} - A_{sample}) / A_{control}] \times 100$	53
Equation 3.3	OD = $(0.0011x [percent]) + 0.0815, R^2 = 0.9983$	54
Equation 3.4	Inhibition (%) = $[(Control_{405} - Extract_{405})/ Control_{405}] \times 100$	55
Equation 3.5	Inhibition (%) = $[(Control_{540} - Extract_{540})/ Control_{540}] \times 100$	56
Equation 4.1	Average cell count = no. of cells x 10^4 /ml	75
Equation 4.2	The total no. of cells/ml = (average count per square) x (the dilution factor) x 10^4	75
Equation 4.3	Viability (%) = $[(A_{Sample} - A_{Blank}) / (A_{Untreated} - A_{Blank})] \times 100$	77

LIST OF ABBREVIATIONS

ABTS 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid)

ADD-1 Adipocyte determination and differentiation-dependent factor 1

ATP Adenosine triphosphate

cAMP Cyclic adenosine monophosphate C/EΒPβ CCAAT/enhancer-binding protein β

CHCl₃ Chloroform cpm Count per minute DEX Dexamethasone

DMEM Dulbecco's modified Eagle's medium

DMSO Dimethylsulfoxide

DPPH 2,2-diphenyl-1-picrylhydrazyl

FBS Fetal bovine serum
GLUT2 Glucose transporter-2
GLUT4 Glucose transporter-4
GME Gracinia managastana avt

GME Garcinia mangostana extract GSK-3 Glycogen synthase kinase-3β

H₂SO₄ Sulphuric acid

IBMX 3-isobutyl-1-methylxanthine

IR Insulin receptor

IS Insulin

KRPH Krebs-Ringer HEPES

MAPK Mitogen-activated protein kinase

MCE Mitotic clonal expansion

MTT 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide

NAD⁺ Nicotinamide adenine dinucleotide

OD Optical density ORO Oil-Red-O

PBS Phosphate buffer saline

PI3K/IRS-1 Phosphatidylinositol 3-kinase/Insulin receptor substrate-1

PIP2 Phosphatidylinositol (4,5)-bisphosphate PIP3 Phosphatidylinositol (3,4,5)-trisphosphate

PKB Protein kinase B

PPARy Peroxisome proliferator-activated receptor gamma

PPH Postprandial hyperglycaemia PS Penicillin-Streptomycin

qRT-PCR Quantitative real-time polymerase chain reaction

 R_f Retention factor

ROS Reactive oxygen species

Rt Retention time

RXRα Retinoid X receptor α

SREBP Sterol regulatory element binding protein

STZ Streptozotocin
TZD Thiazolidinediones
T2D Type 2 diabetes
v/v Volume per volume

CHAPTER ONE

INTRODUCTION

1.1 GENERAL OVERVIEW

Research focused on phytochemicals purified from plant-derived natural products has recently increased all over the world. It is known that various plants may synthesize toxic chemicals to defend against hostile milieu as well as predators. Most of the bioactive compounds have broad range of properties and effects, from being acutely fatal to human to being curative in disease treatment. These secondary metabolites produced by the plants may exert various biochemical and pharmacological functions in animals and men (Acamovic & Brooker, 2005).

Special insight on medicinal plants in tropical countries has shifted researchers to find the lead compounds and pharmacologically viable derivatives for drug design and therapeutic purposes. It is worth noting that the application of scientific approach in modern medicine today has provided skeletons for constructing molecules from plant-derived compounds. On the other hand, crude plant extract also has paid a great attention owing to its properties for treating various ranges of ailments like cancer, heart diseases, diabetes mellitus and obesity (Eyre, Kahn, Robertson, Clark, Doyle & Gansler, 2004)

The recent applications of phytoconstituents were further expanded in areas such as nutraceuticals, agrochemicals and traditional medicines with additional focus as not only curative but delayed onset of complications as well as maintaining health. This has driven meticulous search to unravel underlying mechanisms and biological importance.

The most challenging part while conducting research involving natural products would be the unknown effects and complexity of plant extracts and presence of minute bioactive components in bioassays. It is known that the nature rich with unlimited sources of vital secondary metabolites, which might be of high pharmacological significance. According to World Health Organization, over 21,000 plant species were vastly utilized around the world mainly for medicinal purposes. Tropical countries enormously retained their unexplored medicinal plants and natural products which might contain novel biological activities (Trivedi, 2006).

Malaysia, among the 12 countries rich in biodiversity in the world is estimated to have 1,200 plants species in peninsular alone and 2,000 species in Sabah and Sarawak. Most of the plants are being collected for medicinal purposes or used in herbal preparations. The reserved rainforest of Malaysia offered great chances for research activity due to wide range of available species. Among 12,000 species of flowering plants reported, only 100 were recorded to exhibit medicinal value (Perry & Metzger, 1980). Hence, further investigation should be conducted to elucidate the potential bioactive compounds.

Certain secondary metabolites may hold vital functions in the living plants. For example, flavonoids able to eliminate free radicals produced during photosynthesis. Terpenoids may engage pollinators, as seed dispersers, or inhibit competing plants. Alkaloids usually protect from herbivore animals or insect attacks (phytoalexins). Other secondary metabolites function as cellular signaling molecules or responsible for some other functions in the plants (Mayer, 2004). The prevalence and severity of obesity, type 2-diabetes, and the resultant metabolic syndrome are rapidly increasing. As successful preventive and therapeutic strategies for these life-threatening health ailments often come with adverse side effects, nutritional elements are widely used in

many countries as preventive therapies to prevent or manage metabolic syndrome. Fruits are important dietary components, and contain various bioactive constituents. Many of these constituents have been proven to be useful to manage and treat various chronic diseases such as diabetes, obesity, cancer and cardiovascular diseases. Although exotic fruits are understudied throughout the world due to their limited regional presence, many studies reveal their potent ability to ameliorate metabolic derangements and the resultant conditions i.e. diabetes and obesity. The aim of this article is to review the role of exotic fruits and their constituents in the regulation of which beneficially metabolic functions, can alter diabetes and obesity pathophysiology.

1.2 SIGNIFICANCE OF THE STUDY

Reports concerning *Garcinia mangostana* Linn. are diversely documented for its health-promoting benefits and thus been classified as 'queen of fruit' (Pedraza-Chaverri, Cárdenas-Rodríguez, Orozco-Ibarra & Pérez-Rojas, 2008). All parts of the plant such as leaves, heartwood, ripe fruits, stem barks and fruit hull (pericarp or rind) were reported to elicit significant biological properties. Despite numerous *in vitro* studies shown that the major compound xanthones, a family of tricyclic isoprenylated polyphenols which were extracted from pericarps possess anti-oxidant, anti-proliferative, pro-apoptotic, anti-inflammatory and anti-carcinogenic activities, there were few scientific reports on antihyperglycaemic effect of the plant. Thus, the present study was conducted primarily to ascertain that the crude extracted from pericarp could manage diabetic conditions via cell culture models (*in vitro*) and animal studies (*in vivo*).

1.3 GENERAL OBJECTIVES OF THE STUDY

- 1. To assess the phytochemical profile of *G. mangostana* pericarp extract (GME).
- 2. To determine the α -mangostin content in GME.
- 3. To evaluate the cellular effect of GME on 3T3-L1 adipocyte.
- 4. To examine the antihyperglycaemic effect of GME on streptozotocininduced diabetic rats.
- 5. To perform toxicity study using GME.

1.4 RESEARCH PROBLEM

Diabetes is one of the major metabolic disorders that continue to present as a significant health problem worldwide and mostly associated with chronic hyperglycemic condition and disturbances in protein, carbohydrate and lipid metabolism (Rahimi, Nikfar, Larijani & Abdollahi, 2005). In 2013, a total of 381.8 million adults worldwide were affected with diabetes and estimated to reach 591.9 million in 2035 (Guariguata, Whiting, Hambleton, Beagley, Linnenkamp & Shaw, 2014). Hence, the search for alternative medicinal products is crucial to ameliorate this condition. The study highlighted the development of *G. mangostana* into a widely used nutraceutical. A comprehensive assessment of the biological activities of the extracts was performed using pharmaceutical approach such as phytochemical profiling, antioxidant capacities, enzymatic inhibitions, differentiation capabilities in adipocytes and reduction of blood glucose levels in diabetic rats. The study will provide preclinical evidence and elucidate the extent of pharmacological activities of this extracts as potentially relevant therapeutic drug in diabetes management. The experimental procedure was described in Figure 1.

1.5 HYPOTHESIS

High xanthones content of *G. mangostana* extract may potentiate a number of biological activities including antioxidant properties, inhibition of starch digestion, cellular response and antihyperglycaemic effect in experimental diabetic rats.

1.6 EXPERIMENTAL DESIGN

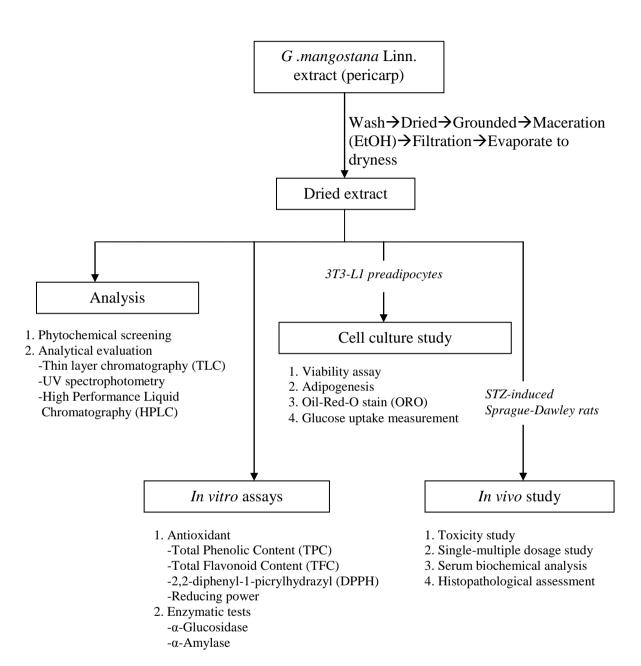


Figure 1. Flow chart of the study.