



IN VITRO STUDY ON GLUCOSE UPTAKE AND
INSULIN STIMULATING PROPERTIES OF
PLUCHEA INDICA (L.) LESS.

BY

WASTUTI HIDAYATI SURIYAH

A thesis submitted in fulfilment of the requirement for
the degree of Master in Pharmaceutical Sciences
(Pharmacology)

Kulliyyah of Pharmacy
International Islamic University
Malaysia

NOVEMBER 2012

ABSTRACT

Insulin resistance and pancreatic β -cells defect are central features of diabetes disorder that may progress to several serious complications. Some of medicinal plants are potential sources for antidiabetic agents. *Pluchea indica* (*beluntas*) is widely distributed in Malaysia and it is believed to have antidiabetic properties. A hypoglycemic effect of *P. indica* in normal rats was reported in the previous study. This research was aimed to study the effects of *P. indica* in glucose and insulin regulation through cell-based *in vitro* model by using 3T3-L1 adipocytes and RIN-5F pancreatic β -cells. *P. indica* was extracted using soxhlet apparatus with *n*-hexane, dichloromethane, ethyl acetate and methanol consecutively. The plant also was macerated using water to yield water extract. In cell viability test, the concentration of 0.2 mg/mL was found to be the maximum concentration of *P. indica* extracts in the absence of cytotoxicity. The preadipocytes were induced to differentiate into mature adipocytes prior to assay. The methanol extract at concentration of 0.05 mg/mL increased glucose uptake in adipocytes ($p < 0.05$), as indicated by up regulation of adipogenesis-regulator *Ppar γ* and insulin-sensitive glucose transporter 4 (*Glut4*) mRNAs. The *n*-hexane and water extracts at concentration of 0.05 mg/mL and 0.1 mg/mL respectively stimulated insulin release in β -cells ($p < 0.05$). Moreover, these extracts elevated the transcription level of insulin receptor substrate 2 (*Irs2*) and glucose-transporter *Glut2* in β -cells. Taken together, this *in vitro* study was useful for a screening model of *P. indica* extracts to demonstrate the glucose uptake in adipocytes and insulin secretion activity in β -cells. These findings also suggest that *P. indica* extract deserves further investigation as a potential agent for diabetes management.

ملخص البحث

المقاومة للإنسولين والخلل في الخلايا بيتا البنكرياسية هي مميزات جوهرية في عيوب الداء السكري التي قد تتطور إلى تعقيدات خطيرة. تعتبر بعض النباتات الطبية مصادر محتملة للعوامل المضادة للسكري. نبات *Pluchea indica* (*beluntas*) واسع الانتشار في ماليزيا. ذكر التأثير الخافض للسكر لـ *P. indica* عند الجرذان في الدراسات السابقة. تهدف هذه الدراسة لدراسة تأثير الـ *P. indica* على الغلوكوز وتنظيم الغلوكوز من خلال نموذج خلوي في الزجاج باستخدام خلايا 3T3-L1 adipocytes و RIN-5F pancreatic β -cells. تم إجراء الاستخلاص المتعاقب للـ *P. indica* للحصول على أربع خلاصات خام اعتماداً على القطبية وهي خلاصات الهكسان، الديكلورومتان، الإيثيل أسيتات، والميتانول. تم تضمين الخلاصة المائية أيضاً في هذه الدراسة. اعتماداً على اختبار حيوية الخلايا، وُجد أن 0.2 mg/mL هو التركيز الأعظمي من خلاصة الـ *P. indica* في غياب السمية الخلوية. تم تحريض طلائع الخلايا الدهنية preadipocytes للتمايز و عملية الصبغ تمت للخلايا البالغة. أظهرت خلاصة المتانول بتركيز 0.05 mg/mL زيادة ملحوظة في قبض الغلوكوز في الخلايا الدهنية ($p < 0.05$)، كما هو ملاحظ من خلال زيادة التنظيم للـ mRNAs لـ adipogenesis-regulator و ناقل الغلوكوز 4 الحساس للإنسولين (*Glut4*). خلاصة الهكسان والخلاصة المائية بتركيز 0.05 mg/mL و 0.1 mg/mL على الترتيب أظهرت تحرير تحريضي للإنسولين في خلايا بيتا ($p < 0.05$). إضافة إلى ذلك، هذه الخلاصات رفعت مستوى النسخ لـ *Insulin receptor substrate 2 (Irs2)* و ناقل الغلوكوز (*Glut2*) في خلايا بيتا البنكرياسية. بالنظر إلى النتائج معاً، هذه الدراسة ضمن الزجاج مفيدة كنموذج اختباري لخلاصات الـ *P. indica* لتوضيح قبض الغلوكوز في الخلايا الدهنية وفعالية إفراز الإنسولين في خلايا بيتا. هذه النتائج أيضاً تقترح أن خلاصة الـ *P. indica* تستحق المزيد من التحري كعامل محتمل للوقاية والعلاج لداء السكري.

APPROVAL PAGE

I certify that I have supervised and read this study and that in my opinion, it conforms to acceptable standards of scholarly presentation and is fully adequate, in scope and quality, as a thesis for the degree of Master in Pharmaceutical Sciences (Pharmacology).

.....
Abdul Razak bin Kasmuri
Supervisor

.....
Muhammad Taher
Co-Supervisor

I certify that I have read this study and that in my opinion, it conforms to acceptable standards of scholarly presentation and is fully adequate, in scope and quality, as a thesis for the degree of Master in Pharmaceutical Sciences (Pharmacology).

.....
Wan Mohd. Azizi bin Mohd.
Sulaiman
Internal Examiner

.....
Mohd. Zaini bin Asmawi
External Examiner

This thesis was submitted to the Department of Basic Medical Sciences and is acceptable as a fulfillment of the requirement for the degree of Master in Pharmaceutical Sciences (Pharmacology).

.....
Wan Mohd. Azizi bin Mohd.
Sulaiman
Head, Department of Basic Medical
Sciences

This thesis was submitted to the Kulliyah of Pharmacy and is acceptable as a fulfillment of the requirement for the degree of Master in Pharmaceutical Sciences (Pharmacology).

.....
Mohamed bin Awang
Dean, Kulliyah of Pharmacy

DECLARATION

I hereby declare that this thesis is the result of my own investigation, except where otherwise stated. I also declare that it has not been previously or concurrently submitted as a whole for any other degree at IIUM or other institutions.

Wastuti Hidayati Suriyah

Signature

Date

INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

**DECLARATION OF COPYRIGHT AND AFFIRMATION
OF FAIR USE OF UNPUBLISHED RESEARCH**

Copyright © 2012 by International Islamic University Malaysia. All right reserved.

***IN VITRO* STUDY ON GLUCOSE UPTAKE AND INSULIN STIMULATING
PROPERTIES OF *PLUCHEA INDICA* (L.) LESS.**

I hereby affirm that The International Islamic University Malaysia (IIUM) holds all rights in the copyright of this Work and henceforth any reproduction or use in any form or by means whatsoever is prohibited without the written consent of IIUM. No part of this unpublished research may be reproduced, store in a retrieval system, or transmitted, in any form or by means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder.

Affirmed by Wastuti Hidayati Suriyah

.....
Signature

.....
Date

*This work is dedicated to my beloved mother and the memory of my father,
who courageously fought diabetes throughout his life until the very end.
His hard work and patience always inspire to those who know him.*

ACKNOWLEDGEMENTS

All praises to Allah (SWT) the Most Beneficent and the Most Merciful for giving me strength and courage to complete this study. This work would have never been possible without the support, understanding and care of many people to whom I owe my sincere thanks and appreciation. May Allah the Almighty bless and reward them all abundantly.

My deepest appreciation and gratitude to my supervisor, Associate Professor Dr. Abdul Razak bin Kasmuri for the enlightening guidance, constructive comment, advice and knowledge in the establishment of this study.

I am deeply indebted to my co-supervisor, Associate Professor Dr. Muhammad Taher for the encouragement, valuable guidance and support throughout the study period.

My sincere thanks and appreciation go to my former Dean's of Kulliyah of Pharmacy, Professor Dato' Dr. Tariq bin Abdul Razak. I am also grateful to Dr. Deny Susanti, Dr. Norazian bt. Mohd Hassan and Sr. Nurulwahida Saad for their kind help and guidance.

I wish to thank to all the helpful laboratory staff in Kulliyah of Pharmacy, Kulliyah of Science and Kulliyah of Allied Health Sciences, especially to Sr. Hazan Haryanti bt. Abdul Halim and Sr. Sriviowarti bt. Noerdin, for their cooperation in laboratory technical assistance.

Sincere thanks go to my special teammate, Sr. Nuraniza bt. Azahari and Br. Mohd. Zaffar bin Mohd. Amiroudine for their warm support during our research journey. Not to forget to all my friends in the postgraduate studies and all those who contributed in this research, I feel very much grateful for your assistance.

Special thanks go to my beloved husband, Solachuddin J. A. Ichwan and my sweet children, Ayako Aziza, Ar Razi Ryoshi and Shafa Syahida for their prayers, patience, unconditional love and encouragement throughout this study. Finally, big thanks to IIUM for providing me the research facilities and grants.

TABLE OF CONTENTS

Abstract.....	ii
Abstract in Arabic.....	iii
Approval Page	iv
Declaration Page.....	v
Copyright Page	vi
Dedication.....	vii
Acknowledgements	viii
List of Tables	xiii
List of Figures.....	xiv
List of Abbreviation.....	xvi
List of Symbols.....	xix
CHAPTER ONE: INTRODUCTION AND LITERATURE REVIEW.....	1
1.1 The Growing Health Problem of Diabetes Mellitus	1
1.1.1 Epidemiology, Definition and Classification of Diabetes Mellitus.....	1
1.1.2 Complications of Diabetes Mellitus and Metabolic Aspect Related to Insulin Resistance.....	3
1.1.3 Interactions between Insulin Resistance and Insulin Secretion in the Development of Glucose Intolerance	5
1.1.4 Molecular Mechanism of Insulin Resistance.....	6
1.2 Pharmacotherapeutic Aspects of Diabetes Mellitus	8
1.2.1 Weight Loss and Exercise	8
1.2.2 Reduce Carbohydrate Absorption.....	9
1.2.3 Delay Gastric Emptying	9
1.2.4 Increase Pancreatic Insulin Secretion	9
1.2.5 Reduce Hepatic Glucose Output (HGO)	10
1.2.6 Increase Insulin Sensitivity.....	11
1.2.7 Insulin Replacement and Insulinmimetic Agents	11
1.3 The Ethnomedicine Approaches in Type 2 Diabetes Mellitus Treatment.....	12
1.4 <i>Pluchea indica</i> L. Less.....	13
1.4.1 Chemical Constituents of <i>P. indica</i>	14
1.4.2 Traditional Uses of <i>P. indica</i>	15
1.4.3 Pharmacological Activities of <i>P. indica</i>	16
1.4.4 <i>P. indica</i> in the Antidiabetic Screening Models	18
1.5 <i>In Vitro</i> Models in Diabetes Mellitus Research.....	18
1.5.1 Adipocytes as A Model Study on Glucose and Lipid Metabolism	19
1.5.2 Pancreatic β Cell Line as A Model Study on Insulin Secretion.....	21
1.6 Objectives of The Study	22
1.7 Flowchart of The Study	23

CHAPTER TWO: EXTRACTION AND PHYTOCHEMICAL SCREENING OF <i>PLUCHEA INDICA</i>	24
2.1 Introduction.....	24
2.2 Objectives	25
2.3 Materials and Methods.....	26
2.3.1 Collection of Plant Materials	26
2.3.2 <i>P. indica</i> Sequential Extraction Procedures by Using Organic Solvents.....	26
2.3.3 <i>P. indica</i> Water Extraction (Aqueous Extract) Procedures	27
2.3.4 Qualitative Phytochemical Screening Procedures	27
2.3.4.1 Detection of Alkaloids.....	27
2.3.4.2 Detection of Carbohydrate and Glycosides	27
2.3.4.2.1 Fehling’s Test.....	28
2.3.4.2.2 Benedict’s Test.....	28
2.3.4.3 Detection of Saponins.....	28
2.3.4.4 Detection of Protein and Amino Acids.....	28
2.3.4.5 Detection of Phytosterols.....	28
2.3.4.5.1 Libermann Burchard’s Test.....	28
2.3.4.6 Detection of Phenolic and Flavonoids.....	29
2.3.4.6.1 Ferric Chloride Test for Phenolic.....	29
2.3.4.6.2 Magnesium and Hydrochloric Acid Reduction Test for Flavonoids	29
2.4 Results.....	29
2.5 Discussion and Conclusion.....	30
CHAPTER THREE: THE EFFECTS OF <i>PLUCHEA INDICA</i> EXTRACTS ON 3T3-L1 ADIPOCYTES	33
3.1 Introduction.....	33
3.2 Objectives	35
3.3 Materials and Methods.....	35
3.3.1 Materials, Equipment, Chemicals.....	35
3.3.1.1 Materials	35
3.3.1.2 Equipment.....	36
3.3.1.3 Chemicals	36
3.3.1.4 Reagents for Assay Systems.....	37
3.3.2 General Procedures for Cell Culture.....	37
3.3.2.1 Cell Thawing	37
3.3.2.2 Cell Subculturing and Maintaining.....	38
3.3.2.3 Cell Freezing.....	38
3.3.2.4 Cell Counting.....	38
3.3.3 <i>P. indica</i> Extracts Preparation	39
3.3.4 Induction of 3T3-L1 Preadipocytes	39
3.3.5 The Cell Viability Screening on Differentiated 3T3-L1 Adipocytes induced by <i>P. indica</i>	39
3.3.6 Lipid Droplet Formation Analysis by Oil Red <i>O</i> staining.....	40
3.3.7 Optical Density Measurement	40
3.3.8 Adipolysis Assay	41
3.3.9 Glucose Uptake Assay	41

3.3.10 Glucose Oxidase Assay	42
3.4 Statistical Analysis.....	42
3.5 Results.....	43
3.5.1 The 3T3-L1 Preadipocytes Growth	43
3.5.2 The Effects of <i>P. indica</i> on 3T3-L1 Adipocytes Viability	44
3.5.3 Induction of 3T3-L1 Preadipocytes	45
3.5.4 Lipid Droplet Formation of 3T3-L1 Adipocytes	46
3.5.5 Quantification of Lipid Accumulation in 3T3-L1 Adipocytes ..	48
3.5.6 Adipolysis Level of 3T3-L1 Adipocytes	49
3.5.7 Glucose Uptake Assay	50
3.5.8 Glucose Oxidase Assay	51
3.6 Discussion and Conclusion.....	51

**CHAPTER FOUR: INSULIN STIMULATING ACTIVITY OF
PLUCHEA INDICA EXTRACTS ON PANCREATIC β -CELLS 54**

4.1 Introduction.....	54
4.2 Objectives	55
4.3 Materials and Methods.....	55
4.3.1 Materials, Equipment and Chemicals	55
4.3.1.1 Reagents for Assay Systems.....	55
4.3.2 <i>P. indica</i> Extracts Preparation.....	56
4.3.3 Insulin-Producing RIN-5F β -Cells and Culture Conditions	56
4.3.4 The Cell Viability Screening on RIN-5F β -Cells.....	57
4.3.5 Insulin Secretion Measurement.....	57
4.4 Statistical Analysis.....	58
4.5 Results.....	59
4.5.1 The Effects of <i>P. indica</i> Extract on RIN-5F Cell Viability	59
4.5.2 The Effects of <i>P. indica</i> Extract on Insulin Secretion of RIN-5F β -Cells	60
4.6 Discussion and Conclusion.....	60

**CHAPTER FIVE: STUDY ON THE EXPRESSION OF THE GENES
IN 3T3-L1 ADIPOCYTES AND RIN-5F β -CELLS INDUCED BY
P. INDICA EXTRACTS..... 63**

5.1 Introduction.....	65
5.2 Objectives	66
5.3 Materials and Methods.....	66
5.3.1 Materials, Equipment and Chemicals	66
5.3.1.1 Reagents for Assay Systems.....	66
5.3.2 <i>P. indica</i> Extracts Preparation.....	66
5.3.3 Cell Culture and Sample Preparation.....	67
5.3.4 Total RNA Extraction and cDNA Analysis	67
5.3.4.1 Phase Separation.....	67
5.3.4.2 RNA Precipitation	68
5.3.4.3 RNA Wash and Re-dissolve	68
5.3.4.4 Spectrophotometric Analysis.....	68
5.3.5 Quantitative Real-Time RT-PCR Analysis	69
5.3.5.1 Reverse-Transcription PCR.....	69
5.3.5.2 Quantitative Real-Time RT-PCR of The Genes.....	69

5.4 Statistical Analysis.....	71
5.5 Results.....	72
5.5.1 The Effects of <i>P. indica</i> Extract on <i>Pparγ</i> and <i>Glut4</i> Gene Expression	72
5.5.2 The Effects of <i>P. indica</i> Extract on <i>Irs2</i> and <i>Glut2</i> Gene Expression	73
5.6 Discussion and Conclusion.....	75
CHAPTER SIX: GENERAL DISCUSSION AND CONCLUSION.....	77
BIBLIOGRAPHY.....	83
LIST OF CONFERENCES ATTENDED	98
APPENDIX A: STANDARD CURVE	99
APPENDIX B: VALIDATION OF <i>β actin</i> RT-qPCR in 3T3-L1 ADIPOCYTES	100
APPENDIX C: VALIDATION OF <i>Glut4</i> RT-qPCR in 3T3-L1 ADIPOCYTES ..	101
APPENDIX D: VALIDATION OF <i>Pparγ</i> RT-qPCR in 3T3-L1 ADIPOCYTES..	102
APPENDIX E: VALIDATION OF <i>β actin</i> RT-qPCR in RIN-5F β -CELLS.....	103
APPENDIX F: VALIDATION OF <i>Irs2</i> RT-qPCR in RIN-5F β -CELL.....	104
APPENDIX G: VALIDATION OF <i>Glut2</i> RT-qPCR in RIN-5F β -CELLS	105

LIST OF TABLES

<u>Table No.</u>		<u>Page No.</u>
1.1	Studies on pharmacological activities of <i>P. indica</i> .	16
2.1	The extraction of <i>P. indica</i> leaves.	29
2.2	Qualitative analysis of the phytochemicals of <i>P. indica</i> extracts.	30
5.1	Components for the reverse transcription reaction.	69
5.2	Primers list for PCR.	70
5.3	Components for PCR.	70
5.4	qRT-PCR reaction under optimal amplification conditions.	71

LIST OF FIGURES

<u>Figure No.</u>		<u>Page No.</u>
1.1	Clinical syndromes associated with insulin resistance.	5
1.2	Insulin signaling pathways.	7
1.3	(A) <i>P. indica</i> (L.) Less. (B) <i>P. indica</i> illustration.	14
1.4	Adipocytes differentiation.	21
1.5	Glucose-stimulated insulin secretion process.	22
1.6	Flowchart of study design.	23
3.1	Growth curve of 3T3-L1 preadipocytes established by cell counting.	43
3.2	The cell viability measured by MTT assay.	44
3.3	Morphology of the 3T3-L1 cells. (A) 3T3-L1 preconfluent preadipocytes (B) 3T3-L1 preadipocytes at two days post confluent. (C) The cells induced by adipogenesis cocktails at day 3. (D) Differentiated cells performed the lipid droplets at day 8.	45
3.4	Lipid droplets formations of 3T3-L1 adipocytes stained with Oil Red O. (A) Undifferentiated cells. (B) Differentiated cells. (C) <i>n</i> -hexane extract-treated cells. (D) Dichloromethane extract-treated cells. (E) Ethyl acetate extract-treated cells. (F) Methanol extract-treated cells. (G) Water extract-treated cells.	47
3.5	(A) The 3T3-L1 cells in differentiated stage stained by Oil Red O after 48 h of sample treatment. (B) Quantification of lipid accumulation after sample treatment.	48
3.6	The adipolysis assay in concentration-dependent manner of the extracts.	49
3.7	Glucose uptake by cells.	50
3.8	Glucose consumption of 3T3-L1 adipocytes measured with glucose oxidase assay.	51

4.1	RIN-5F- β cell line visualized under the inverted microscope.	56
4.2	The RIN-5F β -cell viability rates in several concentrations measured by MTT assay.	59
4.3	<i>P. indica</i> extracts examined to its effect in insulin release activity.	60
5.1	The expression of <i>Pparγ</i> mRNA on 3T3-L1 adipocytes.	72
5.2	The expression of <i>Glut4</i> mRNA on 3T3-L1 adipocytes.	73
5.3	The expression of <i>Irs2</i> mRNA on RIN-5F β -cells.	74
5.4	The expression of <i>Glut2</i> mRNA on RIN-5F β -cells.	74
6.1	Proposed diagram of <i>P. indica</i> action in adipocytes and β -cells.	82

LIST OF ABBREVIATIONS

2hPG	2 hour Plasma Glucose
A	Adenine
ADA	American Diabetes Association
AHPA	American Herbal Products Association
AIM	Adipogenesis-inducing medium
Akt	Murine thymoma viral oncogene homolog
AMM	Adipogenesis-maintaining medium
AMPK	Adenosine Monophosphate
ANOVA	Analysis of Variance
BLAST	Basic Local Alignment Search Tool
BSA	Bovine Serum Albumin
C	Cytosine
C/EBP	CCAAT/enhancer binding protein
Ca ₂₊	Calcium (II)
CaCl ₂	Calcium Chloride
CCK	Cholecystokinin
CDC	Centers for Disease Control
cDNA	complementary DNA
CHD	Coronary Heart Disease
CODE-2	The Cost of Diabetes in Europe– Type II
cm	centimetre
CVD	Cardiovascular Disease
DCM	Dichloromethane
DEPC	Diethylpyrocarbonate
DEX	Dexamethasone
dH ₂ O	double Distilled Water
DKA	Diabetic Ketoacidosis
DM	Diabetes Mellitus
DMEM	Dulbecco's Modified Eagle Medium
DMSO	Dimethyl Sulfoxide
DNA	Deoxyribonucleic Acid
dNTP	deoxynucleoside Triphosphate
DPP	Dipeptidyl Peptidase 4
Eds./ed.	Editions/edition
e.g.	(<i>exempli gratia</i>); for example
ELISA	Enzyme-Linked Immunosorbent Assay
ERK	Extracellular signal
et al	(<i>et alia</i>); and others
EtOH	Ethanol
FBS	Foetal Bovine Serum
FFA	Free Fatty Acid
Fig	Figure
FPG	Fasting Plasma Glucose

g	gram
g	gravity
G	Guanine
GACP	Good Agricultural and Collection Practices
GDM	Gestational Diabetes Mellitus
GIP	Gastric Inhibitory Peptide
GLP-1	Glucagon-Like Peptide 1
GLUT2	Glucose transporter type 2
GLUT4	Glucose transporter type 4
GLUTs	Glucose transporters
Grb	Growth factor receptor-binding protein
GSIS	Glucose-stimulated insulin secretion
h	hour/s
H ₂ O	Water
HCl	Hydrogen Chloride
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
Hex	Hexane
HGO	Hepatic Glucose Output
HGP	Hepatic Glucose Production
IAs	Insulin Analogues
IDDM	Insulin-Dependent Diabetes Mellitus
IFG	Impaired Fasting Glucose
IGT	Impaired Glucose Tolerance
IRS	Insulin Receptor Substrate
IRS2	Insulin Receptor Substrate2
K _{ATP}	ATP-sensitive potassium channel
KCl	Potassium Chloride
KH ₂ PO ₄	Potassium di-hydrogen phosphate
KRB	Krebs Ringer Bicarbonate
KRPH	Krebs Ringer HEPES
MAP	Mitogen-activated protein
MBq	Megabecquerel
MeOH	Methanol
MetS	Metabolic Syndrome
MgCl ₂	Magnesium Chloride
MgSO ₄	Magnesium Sulfate
min	minute
MIX	Methylisobutylxanthine
μL	microlitre
mL	millilitre
mM	millimolar
MMP	Matrix Metalloproteinase
mRNA	messenger Ribonucleic Acid
MTT	3-(4,5-dimethylthiazol-2-y)2,5-diphenyltetrazolium bromide
Na ₃ VO ₄	Natrium Orthovanadate
NaCl	Natrium Chloride
NAFLD	Non Alcoholic Fatty Liver Disease
NaHCO ₃	Sodium Bicarbonate

NaOH	Sodium Hydroxide
NHMS III	National Health Morbidity Survey III
NIDDM	Non-Insulin-Dependent Diabetes Mellitus
NKHS	Non-Ketotic Hyperosmolar State
nm	nanometer
No.	Number/Numbers
N.T	Not Tested
obj	Objective
OD	Optical Density
ODM	Oral Diabetes Medications
OGTT	Oral Glucose Tolerance Test
PBS	Phosphate Buffer Saline
PCOS	Polycystic Ovary Syndrome
PCR	Polymerase Chain Reaction
PDK	PI-3K–dependent kinase
pH	ATP
PI-3K	Phosphatidylinositol 3-kinase
PIP3	Phosphatidylinositol 3,4,5-phosphate
PKC	Protein Kinase C
PPAR	Peroxisome Proliferator Activated Receptor
PPAR γ	Peroxisome Proliferator Activated Receptor γ
RNA	Ribonucleid Acid
RPMI	Roswell Park Memorial Institute medium
RT PCR	Reverse Transcription Polymerase Chain Reaction
RT-qPCR	Real Time Reverse Transcription quantitative PCR
sec	second
SEM	Standard Error of Means
Src	Sarcoma (Schmidt-Ruppin A-2) viral oncogene homolog (avian)
SREBP	Sterol Regulatory Element
T	Thymine
TNF- α	Tumor Necrosis Factor- α
TZD	Thiazolidinedione
WHO	World Health Organization

LIST OF SYMBOLS

α	alpha
β	beta
γ	gamma
p	the probability of obtaining the result
*	statistical significance denotation
n	sample sizes
$^{\circ}\text{C}$	degree Celcius
®	registered trademark

CHAPTER ONE

INTRODUCTION AND LITERATURE REVIEW

1.1 THE GROWING HEALTH PROBLEM OF DIABETES MELLITUS

Diabetes mellitus (DM) is now being recorded as one of the major threat to human health in the 21st century (Zimmet et al., 2001). DM has an influenced on the quality of life of sufferers and caused a wide range of problems from threatened live sustain to death (Zimmet, 2011). The high rates of morbidity and mortality of DM are resulted from the long-term progression of microvascular and macrovascular complications that caused several organs damage. Thus, people with DM are at a risk in serious health problems such as blindness, renal failure, and/or neuropathy, foot ulcers, amputation, cardio vascular and peripheral vascular diseases (World Health Organization [WHO], 1999). The chronic hyperglycemic condition has been known to accelerate the onset and progression of the DM and its complications (Brownlee, 2001; Reusch, 2003).

1.1.1 Epidemiology, Definition and Classification of Diabetes Mellitus

The worldwide prevalence of patient with DM is projected to rise from 2.8 % in 2000 to 4.4 % by 2030, which is affecting 366 million people. Most of the cases that is 298 million will occur in developing countries. This escalating trend is seen in the age between 45 and 64 years. It happens due to numerous factors such as escalation of population aging, urbanization, the high prevalence of obesity and lack of physical activity (Wild et al., 2004).

In Asia, people with DM present moderate obesity and chronic diabetic complications since juvenile which then leads to a shorter life expectancy compared to people in developed countries. People in Asia also have a highly genetic susceptibility to develop the type 2 DM, which is characterized by β -cell failure and abdominal obesity (Yoon, et al., 2006). In tandem with the rising prevalence of obesity and metabolic syndrome in certain ethnic groups of developing countries such as in South Asians, Hispanics and sub-Saharan Africans, the prevalence of the metabolic syndrome is also rapidly increasing in East Asia and China (Misra and Khurana, 2008). In Malaysia, the highest prevalence according to race is in the Asian Indians (28.8 %), followed by the Malays (24.2 %) and the Chinese (14.8 %) (Tan et al., 2004).

The Malaysian National Health Morbidity Survey III (NHMS III) in 2006 affirmed that the overall prevalence of DM in Malaysia among adults of ≥ 30 years old had risen from 8.3 % to 14.9 %. The data seemed to be associated with the increase of obesity (Hussein, 2008).

DM is defined as a chronic metabolic disease, which occurs when the level of blood glucose is higher than normal, resulting from dysfunction and/or lack of insulin produced by pancreatic β -cell and leads to distinctive complications (WHO, 1999). The current classification of DM consists of four main types (American Diabetes Association [ADA], 2004). The type 1, previously refers to insulin-dependent diabetes (IDDM), is characterized by absolute insulin deficiency due to β -cell destruction either immune-mediated or idiopathic. More than 90 % of type 1 DM resulting from the autoimmune islet cell destruction (Carver and Abrahamson, 2009). The type 2, a non-insulin-dependent diabetes (NIDDM), is a relative insulin deficiency owing to insulin secretion defect and insulin resistance (Wright, 2003). The third group consists of other less common types of DM that are caused by or associated with certain

specific conditions and/or syndromes. The fourth group considered as diabetes diagnosed during pregnancy which is called gestational diabetes (GDM) (ADA, 2004).

Diagnosis of type 2 DM is based on the fasting plasma glucose (FPG) and/or 2-hour plasma glucose (2hPG) concentration measurement during an oral glucose tolerance test (OGTT). The limits of diabetes level are FPG and 2hPG concentration at 7.0 and 11.1 mmol/L respectively (Drouin et al., 2009). The ADA Expert Committee identified the category of pre-diabetes state include both impaired fasting glucose (IFG) and/or impaired glucose tolerance (IGT). The current parameters from ADA revealed that a FPG concentration of 100 to 125 mg/dL (≥ 5.6 and < 7.0 mmol/L) is considered to as IFG and that of more than 126 mg/dL is indicated as diabetes condition. IGT is considered if 2hPG concentration at 140 to 199 mg/dL (≥ 7.8 and < 11.1 mmol/L), meanwhile the concentration > 200 mg/dL is considered as diabetes (ADA, 2004).

1.1.2 Complications of Diabetes Mellitus and Metabolic Aspect Related to Insulin Resistance

DM complications are the leading causes of morbidity and mortality of patients and require costly treatment (Umpierrez et al., 2002). In the United States, the estimation of total direct and indirect diabetes costs for 25.8 million people with diabetes reached \$174 billion in 2007 (Centers for Disease Control [CDC], 2011). According to ‘The Cost of Diabetes in Europe – Type II (CODE-2) study’, the total medical cost for patients with both microvascular and macrovascular complications was increased up to 250 % compared to those with no complications (Williams et al., 2002).

Diabetic ketoacidosis (DKA) and non-ketotic hyperosmolar state (NKHS) are two types of acute diabetes complications that correlated to absolute or relative insulin

deficiency (Tripathi and Srivastava, 2006). The extreme hyperglycemic emergencies stages can cause lethargy and coma, with 2–5 % and 15 % of the mortality rates for DKA and NKHS respectively (Umpierrez et al., 2002).

Long-term hyperglycemia leads to the damage of several tissues of the body and produces chronic complications, such as retinopathy, neuropathy and nephropathy in microvascular disorders. The macrovascular problems include atherosclerosis, coronary artery, peripheral vascular and cerebrovascular diseases. Other chronic complications of nonvascular are gastroparesis, sexual dysfunction and skin changes (Tripathi and Srivastava, 2006).

Insulin resistance occurs in the majority of people with the metabolic syndrome (MetS). Reaven (1988) noted that the MetS or syndrome X refers to cluster of various metabolic risk factors that include abdominal obesity, hypertension, glucose intolerance or hyperglycemia and dyslipidemia. Dandona et al. (2005) postulated that the insulin resistance was induced by proinflammatory state of obesity through cytokine TNF- α factor and promoted clinical and biochemical manifestations of the MetS.

Insulin resistance is an underlying factor in this syndrome and it is also a characteristic feature of most of type 2 diabetic obese patients. In addition to that it is correlated with the increase risk of metabolic and cardiovascular cluster of disorders, such as coronary heart disease (CHD) and cardiovascular disease (CVD) (Kassi et al., 2011; Levesque and Lamarche, 2008). However, Insulin resistance syndrome or that associated with hyperinsulinemia are independent risk factors for CVD (Grundy et al., 2004). It is pathophysiologically occurs in insulin-resistant or hyperinsulinemic persons which do not develop to type 2 DM (Figure.1.1) (Reaven, 2005).

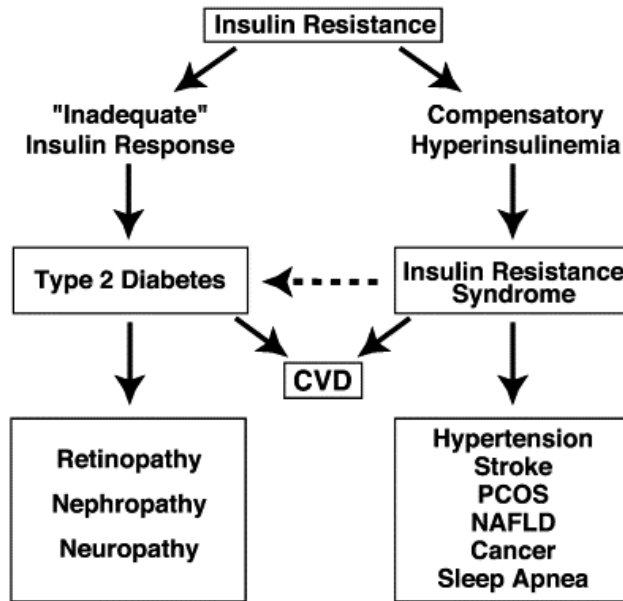


Figure 1.1: Clinical syndromes associated with insulin resistance. Insulin resistance leads to type 2 diabetes and other syndromes that linked to the compensatory hyperinsulinemia. The long-term of hyperglycemia produced microangiopathic complications. PCOS, polycystic ovary syndrome; NAFLD, non-alcoholic fatty liver disease; CVD, cardiovascular disease. (Retrieved from Reaven, 2005).

1.1.3 Interactions between Insulin Resistance and Insulin Secretion in The Development of Glucose Intolerance

The progressive failure of the pancreatic β -cell to compensate insulin resistance is the common pathway responsible for the development of type 2 DM (Kahn, 1998). The hypersecretion of insulin compensatory results in the expansion of β -cell mass and increase the expression of β -cell glucose metabolism enzymes (Cavaghan et al., 2000). In insulin resistance, β -cell may be exhausted due to over secreted insulin to maintain euglycemia. The β -cell failure will results in increase plasma free fatty acid (FFA) concentration, due to insufficient of insulin suppression on the plasma FFA. The elevation of plasma FFA concentration leads to increase hepatic glucose production (Reaven, 1988).