ANTI-CANCER ACTIVITY OF TRITERPENOIDS ISOLATED FROM *LUVUNGA SCANDENS* AGAINST MCF-7 CELLS

BY

PUTRI NUR HIDAYAH AL-ZIKRI BINTI MOHAMAD AKIL

A thesis submitted in fulfilment of the requirement for the degree of Master in Pharmaceutical Science (Pharmaceutical Technology)

Kulliyyah of Pharmacy International Islamic University Malaysia

JUNE 2015

ABSTRACT

Luvunga scandens belong to the family of Rutaceae which usually inhabits tropical and moist environment. This plant is known as 'Mengkurat Jakun' among locals and used traditionally to treat fever and fatigue via decoction. The study aimed to elucidate the compounds from L. scandens stem, which were isolated using bioactivity guided isolation together with their mechanism of action on human breast cancer cell (MCF-7). The bioactivity-guided isolation of cytotoxic agent of the stem of L. scandens resulted in the isolation and characterization of a new triterpenoid from this species, 3oxotirucalla-7,24-dien-21-oic-acid (2), along with a known triterpenoid, flindissol (1). The isolation was conducted using chromatographic techniques on silica gel and sephadex LH-20. The structures of the isolated compounds were elucidated on the basis of spectroscopic analysis including UV, IR, NMR, MS and 2D NMR. The cytotoxic activity of the plant on the proliferation of MCF-7 cell line was evaluated by MTT, WST-1 assay, scanning electron microscope (SEM), flow cytometry and RTqPCR. The cytotoxic evaluation of the extracts showed that IC₅₀ value of the dichloromethane (LSC-SD) and methanol (LSC-SM) extracts from the stem were 75.0 and 77.0 µg/mL respectively. Whereas IC₅₀ value of n-hexane (LSF-H), nhexane:DCM (LSF-HD), DCM (LSF-D) and DCM:MeOH (LSF-DM) fraction extracts were 97.5, 35, 98.9 and 94 µg/mL respectively. The IC₅₀ value of MeOH (LSF-M) fraction could not be extrapolated since none of the concentration of this extract was able to reduce the proliferation activity to 50 %. Compound 1 and 2 showed potent cytotoxicity against MCF-7 cell line with IC₅₀ values of 13.8 µM and 27.5 respectively after 24 h of treatment. Doxorubicin was used as a positive control drug with IC₅₀ values of 6.21 µM. Morphological analysis of the cells surface exhibit the apoptosis results after 24 h treated with LSC-SD extract, 1 and 2, where the cells were rounded up, shrank and lost contact with neighboring cells. Apoptosis of MCF-7 cells treated with both compounds were confirmed by flow cytometry. The results show that compound 1 and 2 exert their anti-proliferative effect on MCF-7 cells through inhibiting cell cycle where the distribution of cells at Sub G₁ phase was 7.7 and 9.3 % respectively. Further evident of apoptosis induction by compound 1 and 2 towards MCF-7 cell lines were assayed by RT-qPCR on the expression of PUMA, caspase-8 and caspase-9. In this assay, both compounds showed the increased of the expression of PUMA, caspase-8 and caspase-9 gene. In conclusion, both triterpenoids, 3-oxotirucalla-7,24-dien-21-oic-acid (2) and flindissol (1) have a potential to be developed as anticancer agents against breast cancer.

ملخص البحث

Luvunga scandens والتي تنمو في العادة بمحيط استوائى ورطب. هذه النبتة تسمى كذلك 'Mengkurat Jakun' ضمن السكان المحليّين وهي تستخدم تقليديًّا لعلاج الحمي والتعب عن طريق استخلاصها بالغلى في الماء. الهدف من هذه الدراسة كان معرفة تأثير المركبات والتي تمّ استخلاصها من L. scandens ثمّ عزلها باستخدام تقنية جذع العزل عن طريق النشاط الحيوي الموجّه بالإضافة إلى معرفة آلية العمل على خلايا الثدي السرطانية البشرية (MCF-7) تقنية العزل عن طريق النشاط الحيوي الموجّه للعامل السام بالنسبة للخلايا L. scandens أدّى إلى عزل وتمييز ثلاثي لجذع تيربنويد جديد من (2) 3-oxotirucalla-7,24-dien-21-oic-acid، بالإضافة إلى آخر معروف (1) flindissol، سلالة عملية العزل تمّت بواسطة تقنيات كروماتوغرافية على جيل السيليكا LH-20. بنية المركبات المعزولة تمّ تحليلها والسيفاديكس باستخدام الأشعة الطيفيّة بالإضافة إل VD NMR ،MS ،NMR ،IR ،UV النشاط السامّ للخلايا الخاص بالنبتة ضدّ MCF-7 تمّ تقييمه بتقنيات WST-1 ،MTT، ميكروسكوب التحليل الالكتروني خلايا تقنية التدفّق الخلوي وتقنية RT-qPCR، التقييم السام للخلايا الخاص بالمستخلصات أظهر IC50 الخاصة بمستخلصات الديكلوروميثان (LSC-SD) والميثانول بأنّ قيم -LSC (SM من الجذع كانت تعادل 77.0 و 75.0 مكغ \backslash مل على التوالى. حيث كانت قيمة IC_{50} الخاصة بـ ن هيكسان، (LSF-H)، ن-هيكسان (DCM (LSF-HD) وكذا (LSF-DM) من أجزاء المستخلصات تعادل 97.5، 35، 98.9 و 94 مكغ \مل على التوالي. قيمة IC50 الخاصة بجزء الميثانول (LSF-M) لم يتمّ تقدير قراءتما بما أنّ تركيزات هذا المستخلص لم تتمكّن من تخفيض نشاط تكاثر الخلايا إلى % المركبات 1 و 2 أظهرت فعالية عند النشاط السامّ للخلايا ضدّ خلايا MCF-7 بقيم IC50 تعادل 50. 13.8 مكم و 27.5 على التوالي بعد 24 ساعة من العلاج. تمّ استخدام دوكسوروبيسين كدواء دليل يكابي بقيم IC_{50} تعادل 6.21 مكم. التحليل المورفولوجي لسطح الخلايا أظهر نتائج موت الخلايا بعد IC_{50} ساعة من العلاج LSC-SD و 2 بمستخلص. حيث كانت الخلايا كرويّة، منكمشة وفقدت التواصل مع الخلايا المجاورة لها. موت T-MCF المبرمج المعالجة بكلا المركبين تمّ تأكيده بواسطة تقنية التدفّق الخلوي. النتائج تظهر بأنّ المركبين 1 و 2 خلايا. يطرحان خصائص مضادة للتكاثر على خلايا MCF-7 عن طريق منع الدورة الخلوية حيث كان تقسيم الخلايا عند مرحلة 7.7 Sub G₁ و 9.3 بالمئة على التوالي. الدليل الموالي على وجود موت مبرمج للخلايا من طرف المركبين 2 و 1 ضدّ خلايا تمّت معايرته عن طريق تقنية RT-qPCR حول التعبير عن PUMA، الكاسباس 8 والكاسباس 9. كخلاصة فإنّ الترابتيربنويدات -3 oxotirucalla-7,24-dien-21-oic-acid (2)، والفلينديسول (1) يملكان خصائص يمكن تطويرها كعلاج ضد السرطان وبالأخص سرطان الثدي.

ABSTRAK

Luvunga scandens tergolong dalam keluarga Rutaceae yang kebiasaannya mendiami persekitaran tropika dan lembap. Pokok ini dikenali sebagai 'Mengkurat Jakun' dalam kalangan penduduk tempatan dan digunakan secara tradisional untuk mengubati demam dan keletihan dengan meminum air rebusan. Kajian ini bertujuan untuk menguraikan sebatian dari L. scandens batang yang diasingkan berdasarkan pengasingan berpadukan bioaktiviti bersama-sama tindak balas sebatian terhadap sel kanser payudara manusia (MCF-7). Pengasingan berpandukan bioaktiviti daripada batang pokok L. scandens menghasilkan pemencilan dan pencirian triterpenoid baru, 3-oxotirucalla-7,24-dien-21-oic-asid (2) serta triterpenoid yang diketahui iaitu (1). Pengasingan telah dijalankan dengan menggunakan teknik kromatografik di atas gel silica dan sephadex LH-20. Struktur sebatian yang telah diasingkan itu telah diuraikan berdasarkan analisis spektroskopi termasuk UV, IR, NMR, MS dan 2D NMR. Kesan sitotoksik daripada pokok ini terhadap pertumbuhan sel MCF-7 telah dijalankan menggunakan ujian MTT, WST-1, scanning electron microscope (SEM), flow cytometry dan RT-qPCR. Ujian sitotoksik telah menunjukan nilai IC₅₀ diklorometana (LSC-SD) dan metanol (LSC-SM) ekstrak dari batang masing-masing adalah 75.0 dan 77.0 µg/mL. Manakala nilai IC₅₀ n-heksana (LSF-H), n-heksana: DCM (LSF-HD), DCM (LSF-D) dan DCM: MeOH (LSF-DM) ekstrak masing-masing adalah 97.5, 35, 98.9 dan 94 µg/mL. Nilai IC₅₀ daripada MeOH (LSF-M) tidak boleh dikenalpasti kerana ekstrak ini tidak dapat merencat 50 % daripada aktiviti pertambahan sel. Sebatian 1 dan 2 menunjukkan kesan sitotoksik yang kuat terhadap sel MCF-7 dengan nilai IC₅₀ 13.8 dan 27.5 µM selepas 24 jam rawatan. Doxorubicin telah digunakan sebagai ubat kawalan positif dengan nilai IC₅₀ 6.21 µM. Analisis morfologi permukaan sel selama 24 jam menunjukkan kesan apoptosis selepas rawatan dengan ekstrak LSC-SD, sebatian 1 dan 2, di mana sel-sel itu telah membulat, mengecut dan kehilangan hubungan dengan sel bersebelahan. Apoptosis daripada sel MCF-7 yang dirawat dengan kedua-dua sebatian telah disahkan dengan flow cytometry. Keputusan menunjukkan bahawa sebatian 1 dan 2 memberi kesan anti-proliferasi pada sel MCF-7 melalui pengencatan kitaran sel di mana pembahagian sel di fasa Sub G₁ adalah masing-masing 7.7 dan 9.3 %. Bukti yang lagi jelas tentang apoptosis oleh sebatian 1 dan 2 terhadap sel MCF-7 telah diuji oleh RT-qPCR pada ekspresi gen PUMA, caspase-8 dan caspase-9. Dalam pengujian ini kedua-dua sebatian menunjukkan peningkatan ekspresi pada gen PUMA, caspase-8 dan caspase-9. Kesimpulannya, keputusan kajian menunjukkan triterpenoids, 3-oxotirucalla-7,24dien-21-oic-acid (2) dan flindissol (1) berpotensi untuk dijadikan agen menentang kanser payudara.

APPROVAL PAGE

I certify that I have supervised and read this stuto acceptable standards of scholarly presentation quality, as a thesis for the degree of (Pharmaceutical Technology)	on and is fully adequate, in scope and
	Muhammad Taher Supervisor
	Deny Susanti Co-Supervisor
	Solachuddin Jauhari Arief Ichwar Co-Supervisor
I certify that I have read this study and that in standards of scholarly presentation and is fully thesis for the degree of Master of Phar Technology)	y adequate, in scope and quality, as a
	Qamar Uddin Ahmed Internal Examiner
	Masa Aki Ikeda External Examiner
This thesis was submitted to the Department accepted as a fulfilment of the requirem Pharmaceutical Science (Pharmaceutical Technology)	ents for the degree of Master of
	Juliana Md Jaffri Head, Department of Pharmaceutical Technology
This thesis was submitted to the Kulliyyah fulfilment of the requirements for the degree (Pharmaceutical Technology)	•
	Siti Hadijah Shamsuddin Dean, Kulliyyah of Pharmacy

DECLARATION

I hereby declare that this thesis is the result of my own investigations, except where

otherwise stated. I also declare that it has not been previously or concurrently
submitted as a whole for any other degrees at IIUM or other institutions.
Putri Nur Hidayah Al-Zikri Binti Mohamad Akil

Signature.....

Date

INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

DECLARATION OF COPYRIGHT AND AFFIRMATION OF FAIR USE OF UNPUBLISHED RESEARCH

Copyright ©2015 by Putri Nur Hidayah Al-Zikri. All rights reserved.

ANTI-CANCER ACTIVITY OF TRITERPENOIDS ISOLATED FROM LUVUNGA SCANDENS AGAINST MCF-7 CELLS

No part of this unpublished research may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder except as provided below.

- 1. Any material contained in or derived from this unpublished research may be used by others in their writing with due acknowledgement.
- 2. IIUM or its library will have the right to make and transmit copies (print or electronic) for institutional and academic purposes.
- 3. The IIUM library will have the right to make, store in a retrieval system and supply copies of this unpublished research if requested by other universities and research libraries.

Affirmed by Putri Nur Hidayah Al-Zikri Binti Mohamad Akil		
Signature	Date	

ACKNOWLEDGEMENTS

In the name of Allah, the Most Benevolent and the Most Merciful. First and foremost, praise to Allah, the Great and Almighty surrounded me under His auspices during my Master study, in the Kulliyyah of Pharmacy, International Islamic University Malaysia (IIUM). I would like to express my sincere gratitude to my supervisor, Assoc. Prof. Dr. Muhammad Taher for his advices, support, guidance and encouragement during experimental work and thesis writing. I would also like to thank to my co-supervisor Assoc. Prof. Dr. Deny Susanti (Kulliyyah of Science) and Assoc. Prof. Dr. Solachuddin (Kulliyyah of Dentistry) for their kind assistance and guidance. All of your valuable advices and guidance to finish my study will not be forgotten. Special thanks to my husband, Shahurin Abd. Rahman for his understanding, prayer and unconditional moral support. My heartiest gratitude is for my parents Mohamad Akil Jaapar and Puteri Algunda Taib, my siblings, Mohammad Svaiful Amri, Putri Nur Ain, Mohammad Syaiful Amir, Mohammad Syaifullah and Puteri Nursyafigah. All this would not be possible without their cares, loves and prayers. I expressed my gratefulness to Sr. Wastuti Hidayati, Anugerah Budipramata Adina and Tengku Muhamad Faris Syafiq for helping and accompany me during my experimental work. Other labmates, Huwaida Abdul Azis, Hanisuhana, Sama Nazia and Afnan, thanks for your support and our times together. To all my friends, Asia Sassi, Hafizah, Izzati, Nadia Hanis, Huda, Farah Dayana, Hawa, Laili, Ain, Fathin, Mariam Sa'adah, Syafinaz, Aya, Fahmi, Anas, Mahmood, Khairul, Azad, Hassan, Fahim and all postgraduate, thanks for your friendship and supports. Their friendship made my life a wonderful experience throughout the course. This study could not have been possible without cooperation, support and help of administrative staffs, Sis Haslina and Sis Syuhaiha, Science Officer, Sis Zaililah, Sis Zian, Sis. Wahida and Sis. Rusianti, technical and non-technical staff of Kulliyyah of Pharmacy, Bro.Dzadil, Bro. Faris, Bro. Hanif and Bro. Razif. Special thanks to Sis, Hazan Harvanti for her help in terms of consumable items, En. Fazlin Rezali from SIRIM Berhad to fulfill my request for NMR analysis, Sis. Afzan, Science Officer, and all staffs and imolec group members of ICRACU for the interesting seminar, workshop and demonstration. I would like to thank all from the depths of my heart who contributed directly or indirectly to this study. Lastly, very special thanks to Ministry of Education Malaysia for the scholarship, Ministry of Science, Technology and Innovation (MOSTI) for the partially support via Science Fund with research grant no. 02-01-08-SF0110. Thank you all!

TABLE OF CONTENTS

Abstract	ii
Abstract in Arabic	iii
Abstract in Bahasa Malaysia	iv
Approval page	
Declaration	
Copyright Page	vii
Acknowledgements	
List of Tables	
List of Figures	
List of Equations	
CHAPTER 1: INTRODUCTION	1
1.1 General Overview	
1.2 Objective of the Study	
1.2.1 General Objective.	
1.2.2 Specific Objectives	
1.3 Experimental Design	
1.5 Experimental Design	,
CHAPTER 2: LITERATURE REVIEW	4
2.1 Natural Product As Anti-Cancer Agent	
2.1.1 Secondary Metabolites in Natural Products	
2.2 Cancer	
2.2.1 Epidemiology of Cancer	
2.2.2 Breast Cancer	
2.2.2.1 Breast Adenocarcinoma Cell Line (MCF-7)	
2.2.3 Cell Culture	
2.2.3 Cell Culture in Cancer Research	
2.2.4 Cell Cycle	
2.2.5 Cell Death (Apoptosis, Necrosis and Autophagy)	
2.3 Plant Description	
2.3.1 Rutaceae Family	
2.3.2 Genus Luvunga	
<u> </u>	
2.3.3 Traditional Usage	
2.3.4 Phytochemical and Biological Activities	∠1
CHARTER 2. DHATE CHEMICERA OF LUMINGA CCAMPENC	24
CHAPTER 3: PHYTOCHEMISTRY OF LUVUNGA SCANDENS	
3.2 Materials and Methods	
3.2.1 General	
3.2.2 Chemicals and Reagents	
3.2.3 Plant Collection	
3.2.4 Extraction	
3.2.5 Qualitative Phytochemical Screening Procedures	
3.2.5.1 Detection of Alkaloids	
3.2.5.1.1 Mayer's test	27

3.2.5.2 Detection of Terpenoids and Steroids	
3.2.5.2.1 Libermann-Burchard test	
3.2.5.3 Detection of Flavonoids	
3.2.5.3.1 Alkaline Reagent test	
3.2.5.4 Detection of Saponins	28
3.2.5.4.1 Froth test	28
3.2.6 Fractionantion Procedures	28
3.2.7 Isolation Procedures	29
3.2.7.1 Spectroscopic Analysis	30
3.2.7.1.1 Infrared (IR)	
3.2.7.1.2 Ultraviolet (UV)	
3.2.7.1.3 Mass Spectrometry (MS)	31
3.2.7.1.4 Nuclear Magnetic Resonance (NMR)	
3.2.8 Flowchart of Extraction, Fractionation and Isolation	
3.3 Results and Discussion	
3.3.1 Plant Extraction.	33
3.3.2 Qualitative Phytochemical Screening of <i>L. scandens</i> Extracts.	
3.3.3 Fractionation by Vacuum Liquid Chromatography	
3.3.4 Isolation by Column Chromatography	
3.3.4.1 Compound 1	
3.3.4.1.1 UV spectrum	
3.3.4.1.2 FTIR spectroscopy	
3.3.4.1.3 ¹ H NMR	39
3.3.4.1.4 ¹³ C NMR	
3.3.4.1.5 ¹ H- ¹ H COSY	
3.3.4.1.6 ¹ H- ¹³ C HMQC	
3.3.4.1.7 HRMS	
3.3.4.2 Compound 2	
3.3.4.2.1 UV spectrum	
3.3.4.2.2 FTIR spectroscopy	
3.3.4.2.3 ¹ H NMR	
3.3.4.2.4 ¹³ C NMR	
3.3.4.2.5 ¹ H- ¹ H COSY	
3.3.4.2.6 ¹ H- ¹³ C HMQC	
3.3.4.2.7 HRMS	
3.3.4.2./ TRNS	34
CHAPTER 4: CYTOTOXICITY OF LUVUNGA SCANDENS ON	MCE 7
CELLS	
4.1 Introduction	
4.2 Materials and Methods	
4.2.1 General	
4.2.2 Chemicals and Reagent	
4.2.3.1 Sterilization of Equipments	
4.2.3.2 Thawing and Propagation of Cells	
4.2.3.2.1 Thawing of Cells	
4.2.3.2.2 Propogation of Cells	
4.2.3.3 Subculturing Cells	
4.2.3.4 Counting of Cells	60

4.2.3.5 Cryopreservation of cells	61
4.2.4 Cell Line	61
4.2.5 MCF-7 Growth Curve Determination	62
4.2.6 Cell Proliferation Assay	62
4.2.6.1 MTT Assay	
4.2.6.2 WST-1 Assay	63
4.2.7 Morphological Staining of Apoptosis	64
4.2.8 Statistical Analysis	66
4.3 Results And Discussion	67
4.3.1 MCF-7 Growth Curve	67
4.3.2 Cell Proliferation Assay	68
4.3.2.1 MTT Assay	
4.3.2.1.1 Cytotoxic Activity of <i>L. scandens</i> Extracts	69
4.3.2.1.2 Cytotoxic Activity of <i>L. scandens</i> Fraction Extra	cts 70
4.3.2.1.3 Cytotoxic Activity of <i>L. scandens</i> Isolated	
Compounds	74
4.3.2.2 WST-1 Assay	
4.3.3 Morphological Analysis of MCF-7	82
CHAPTER 5: CELL CYCLE ANALYSIS	
5.1 Introduction	85
5.2 Materials and Methods	86
5.2.1 General	
5.2.2 Cell Cycle Staining Protocol	
5.3 Results and Discussion	88
CHAPTER 6: GENE EXPRESSION ANALYSIS	
6.1 Introduction	
6.2 Materials and Methods	
6.2.1 General	
6.2.2 Chemicals and Reagent	
6.2.3 Cell Culture and Sample Preparation	
6.2.4 Total RNA Extraction and cDNA Analysis	
6.2.4.1 Phase Separation	
6.2.4.2 RNA Precipitation	
6.2.4.3 RNA Wash	
6.2.4.4 RNA Resuspension	
6.2.4.5 Spectrophotometric Analysis	
6.2.5 Quantitative Real-Time RT-PCR Analysis	
6.2.5.1 Reverse-Transcription PCR	
6.2.5.2 Quantitative RT-PCR of the Genes	
6.2.6 Statistical Analysis	
6.3 Results and Discussion	
6.3.1 RNA concentration and purity	
6.3.2 Quantitative RT-PCR of the Genes	
6.3.2.1 Effects of compound 1 and 2 on <i>PUMA</i> Gene Expression	1.103
6.3.2.2 Effects of compound 1 and 2 on Caspase-8 Gene	104
Expression	104

6.3.2.3 Effects of compound 1 and 2 on Caspase-9 Gene	
Expression	105
CHAPTER 7: CONCLUSION	108
7.1 General	
7.2 Recommendation for Future Works	110
REFERENCES	111
APPENDIX I: UV SPECTRUM OF COMPOUND 1	119
APPENDIX II: ¹ H NMR OF COMPOUND 1	120
APPENDIX III: ¹³ C NMR OF COMPOUND 1	121
APPENDIX IV: HMBC OF COMPOUND 1	122
APPENDIX V: UV SPECTRUM OF COMPOUND 2	123
APPENDIX VI: ¹ H NMR OF COMPOUND 2	124
APPENDIX VII: ¹³ C NMR OF COMPOUND 2	
APPENDIX VIII: HMBC OF COMPOUND 2	126
APPENDIX IX: STATISTICAL ANALYSIS RESULTS OF STEM EXTR	
APPENDIX X: STATISTICAL ANALYSIS RESULTS OF F	RACTION
EXTRACTS	128
APPENDIX XI: STATISTICAL ANALYSIS RESULTS OF CO)MPOUND
1 AND 2	129
APPENDIX XII: HISTOGRAM GRAPH OF COMPOUND 1	
APPENDIX XIII: HISTOGRAM GRAPH OF COMPOUND 2	131
APPENDIX XIV: HISTOGRAM GRAPH OF POSITIVE CONTROL	132
APPENDIX XV: HISTOGRAM GRAPH OF UNTREATED CELLS	133
APPENDIX XVI: MELT CURVE ANALYSIS OF PUMA GENE	134
APPENDIX XVII: MELT CURVE ANALYSIS OF CASPASE-8 GENE	135
APPENDIX XVIII: MELT CURVE ANALYSIS OF CASPASE-9 GENE	136
APPENDIX XIX: PUBLICATION IN BRAZILIAN JOURI	
PHARMACOGNOSY	137

LIST OF TABLES

<u>Table No.</u>		Page No.
2.1	List of plant derivatives used in cancer therapy	5
2.2	Differences in morphology, biochemical, physiologic and pathologic role between apoptosis and necrosis	19
3.1	Extracts form L. scandens	33
3.2	Qualitative phytochemical screening of <i>L. scandens</i> extracts	33
3.3	Fractions of L. scandens (LSF) from DCM extract	34
3.4	Isolated compounds from L. scandens stem	35
3.5	FTIR spectral data of compound 1 (cm ⁻¹)	37
4.1	Concentration of samples used in Morphological Analysis	64
4.2	IC ₅₀ value of <i>L. scandens</i> fraction DCM extracts	71
5.1	DNA content of MCF-7 cell, untreated and treated with different compounds and drug for 24 h as determined by PI-stained flow cytometry cell cycle analysis	88
6.1	Volume of each sample and DMEM for RT-PCR treatment in 10 cm ² dish with total volume of 10 mL	96
6.2	Components for the reverse transcription reaction	99
6.3	List of primers	100
6.4	Components for PCR	101
6.5	RT-qPCR reaction under optimal amplification conditions	101
6.6	RNA concentration of each samples and control	102

LIST OF FIGURES

Figure No.		Page No.
1.1	Flow chart of the study	3
2.1	Characteristic of standard growth pattern of culture cells	11
2.2	Schematic presentation of the proposed mechanisms of cellular reduction of MTT and WST-1	14
2.3	Schematic mechanism of the MTT reduction	14
2.4	Schematic mechanism of the WST-1 reduction	15
2.5	Division of cells in cell cycle control system	16
2.6	Differences in the process of apoptosis, autophagy and necrosis cell death process	18
2.7	L. scandens plant	21
2.8	Chemical structure of 3-epi-skimmiarepin A	22
2.9	Chemical structure of 21,23-epoxy-7α,21-dihydroxyapotirucalla-14,24-dien-3-one	22
2.10	Chemical structure of 3-epi-flindissol	23
3.1	Voucher specimens of L. scandens plant	26
3.2	Flow chart of extraction and isolation of L. scandens	32
3.3	Chemical structure of compound 1	35
3.4	TLC result of compound 1	36
3.5	UV spectrum of compound 1	37
3.6	FTIR spectrum of compound 1	38
3.7	¹ H NMR spectrum of compound 1 in CDCl ₃	39
3.8	¹³ C NMR spectrum of compound 1 in CDCl ₃	40
3.9	¹ H- ¹ H COSY NMR spectrum of compound 1 in CDCl ₃	41

3.10	¹ H- ¹ H COSY NMR spectrum of compound 1 in the range of 0.6-2.7 ppm	42
3.11	¹ H- ¹³ C HMQC NMR spectrum of compound 1 in CDCl ₃	43
3.12	HRMS spectrum of compound 1	44
3.13	Chemical structure of compound 2	45
3.14	TLC pattern of compound 2	46
3.15	UV spectrum of compound 2	47
3.16	FTIR spectrum of compound 2	48
3.17	¹ H NMR spectrum of compound 2 in CDCl ₃	49
3.18	¹³ C NMR spectrum of compound 2 in CDCl ₃	50
3.19	¹ H- ¹ H COSY NMR spectrum of compound 2 in CDCl ₃	51
3.20	¹ H- ¹ H COSY NMR spectrum of compound 2 in the range of 0.85-1.85 ppm	52
3.21	¹ H- ¹³ C HMQC NMR spectrum of compound 2 in CDCl ₃	53
3.22	¹ H- ¹³ C HMQC NMR spectrum of compound 2 in the range of 5-55 ppm for ¹³ C atoms and 0.7-2.5 ppm for ¹ H atoms	53
3.23	HRMS spectrum of compound 2	54
4.1	Sample processing chart for SEM	66
4.2	Growth curve of the MCF-7 cell line	67
4.3	Cytotoxicity activity of DCM (LSC-SD) extract of <i>L. scandens</i> stems against MCF-7 cell lines by MTT assay	69
4.4	Cytotoxicity activity of MeOH (LSC-SM) extract of <i>L. scandens</i> stems against MCF-7 cell lines by MTT assay	70
4.5	Cytotoxicity activity of <i>n</i> -hexane (LSF-H) fraction extract against MCF-7 cell lines by MTT assay	71

4.6	Cytotoxicity activity of <i>n</i> -hexane: DCM (LSF-HD) fraction extract against MCF-7 cell lines by MTT assay	72
4.7	Cytotoxicity activity of DCM (LSF-D) fraction extract against MCF-7 cell lines by MTT assay	72
4.8	Cytotoxicity activity of DCM: MeOH (LSF-DM) fraction extract against MCF-7 cell lines by MTT assay	73
4.9	Cytotoxicity activity of MeOH (LSF-M) fraction extract against MCF-7 cell lines by MTT assay	73
4.10	Cytotoxicity activity of <i>L. scandens</i> compound 1 against MCF-7 cell lines by MTT assay	74
4.11	Cytotoxicity activity of <i>L. scandens</i> compound 2 against MCF-7 cell lines by MTT assay	75
4.12	Cytotoxicity activity of Doxorubicin as positive control drug against MCF-7 cell lines by MTT assay	75
4.13	Reduction of purple tetrazolium salt of MTT to a yellow formazan dye	76
4.14	MCF-7 cell lines treated with different concentration of compound 1 observed under the phase-contrast inverted microscope with magnification of 20x	77
4.15	MCF-7 cell lines treated with different concentration of compound 2 observed under the phase-contrast inverted microscope with magnification of 20x	78
4.16	Cytotoxicity activity of compound 1 against MCF-7 cell lines by WST-1 assay	79
4.17	Cytotoxicity activity of compound 2 against MCF-7 cell lines by WST-1 assay	80
4.18	Reduction of red tetrazolium salt of WST-1 to a dark vellow/ orange formazan dve	81

4.19	Scanning electron photomicrograph of the surface morphology of MCF-7 cells treated with different concentration of compound 1 for 24 h under magnification of 2.00k x	82
4.20	Scanning electron photomicrograph of the surface morphology of MCF-7 cells treated with different concentration of compound 2 for 24 h under magnification of 2.00k x	83
4.21	Scanning electron photomicrograph of the surface morphology of MCF-7 cells treated with IC ₅₀ value of DCM extract (LSC-SD), Doxorubicin as positive control drug and untreated cells grown in DMEM for 24 h	84
5.1	DNA content frequency histogram of MCF-7 cells treated with IC ₅₀ value of compound 1 for 24 h	89
5.2	DNA content frequency histogram of MCF-7 cells treated with IC ₅₀ value of compound 2 for 24 h	90
5.3	DNA content frequency histogram of MCF-7 cells treated with IC ₅₀ value of Doxorubicin for 24 h	90
5.4	DNA content frequency histogram of MCF-7 cells were grown with untreated media for 24 h	91
6.1	Effects of compound 1 and 2 on PUMA mRNA expression	104
6.2	Effects of compound 1 and 2 on caspase-8 mRNA expression	105
6.3	Effects of compound 1 and 2 on <i>caspase-9</i> mRNA expression	106

LIST OF EQUATIONS

Equation No.	<u>I</u>	Page No.
3.1	% of extract = $\frac{\text{Extract weight}}{\text{Stems powder weight}} \times 100$	27
	Distance to a like the second	
3.2	$R_f = \frac{\text{Distance travelled by compound}}{\text{Distance travelled by solvent}}$	30
3.3	Degree of Unsaturation = $\frac{2C + 2 + N - X - H}{2}$	31
4.1	Cells/mL =	
	$\frac{\text{Number of cells counted}}{\text{Number of square counted}} \times 10^4 \times \text{Dilution factor}$	61
4.2	Total cells = Cells/mL Volume of original suspension	
		61
4.3	Percentage viability = $\frac{\text{Viable cells}}{\text{Total number of cells}} \times 100$	62
4.4	Viability of cells = $ \left[\frac{OD_{Sample} - OD_{Blank}}{OD_{Control} - OD_{Blank}} \right] \times 100 $	68
6.1	Concentration of RNA = $A_{260} \times Dilution \times 40$ sample (µg/mL)	98

CHAPTER ONE

INTRODUCTION

1.1 GENERAL OVERVIEW

Most plants consist of phytochemicals or known as secondary metabolites which have disease preventive properties. Plants produce these chemicals to protect themselves from any possible harm in the ecological environment. Recent scientific researches show that phytochemicals have the potential to protect human against various diseases including cancer, heart disease, diabetes and high blood pressure (Liu, 2004; Lodish, Berk, Zipursky, Matsudaira, Baltimore, & Darnell, 2000).

Phytochemicals are present in smaller quantities in plants which include alkaloids, steroids, flavonoids, terpenoids, tannins and many others. Nearly about 50% of drugs used in medicine are plant origin and only a small fraction of plants with medicinal activity has been assayed. Based on the previous research studies and findings, phytochemicals have been reported to play an important role in the prevention of cancer. It can be proven via revelation of new anticancer compound, Taxol. Taxol is originally isolated from the stembark of *Taxus brevifolia*. It is a new antitumor drug approved by FDA for the treatment of breast, ovarian and non-small-cell lung carcinomas (Peteros & Uy, 2010).

Breast cancer is the second most frequently diagnosed cancer in the developed and developing countries. Breast cancer occurs due to the out-of-control growth of normal cells in breast. Instead of dying like normal cells, breast cancer cells can grow and invade other tissues. Different types of cancer can behave very differently. They

grow at different rates and respond to different treatments. That is why people with cancer need treatment that is targeted their own kind of cancer.

This research study was focused to evaluate the effect of plant-derived terpenoids specifically known as triterpenoids from *Luvunga scandens* against human breast adenocarcinoma (MCF-7) cell lines. These plant-derived triterpenoids were targeted to inhibit the breast cancer cells via apoptosis pathway.

1.2 OBJECTIVES OF THE STUDY

1.2.1 General Objective

To evaluate the cytotoxic effects of *L. scandens* on human breast adenocarcinoma (MCF-7) cell line through bioactivity-guided isolations of active compounds.

1.2.2 Specific Objectives

- To evaluate the cytotoxic activity of L. scandens's stem extracts against
 MCF-7 cells.
- 2. To isolate anticancer compounds from the active fractions.
- To observe the morphological changes of MCF-7 cells treated with different concentrations of isolated compounds.
- 4. To analyze cell cycle profile of MCF-7 treated with *L. scandens* compounds.
- 5. To determine the gene expression of *PUMA*, *caspase-8* and *caspase-9* genes at mRNA level in MCF-7 cell lines treated with *L. scandens* compounds.

1.3 EXPERIMENTAL DESIGN

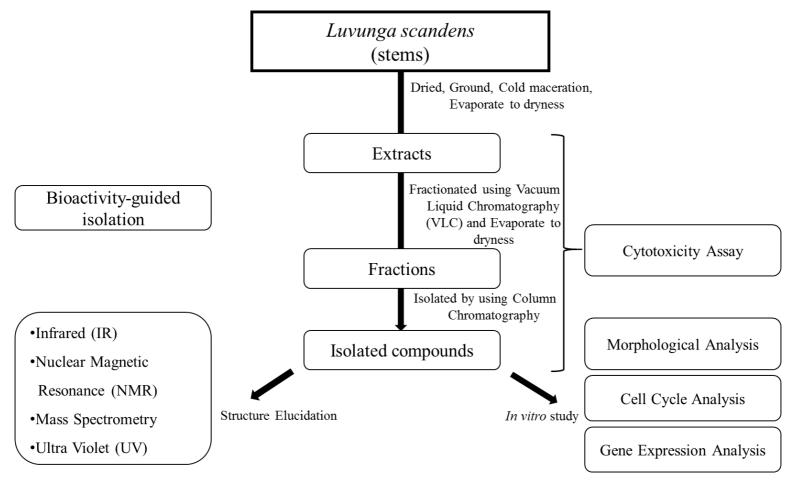


Figure 1.1 Flow chart of the study.

CHAPTER TWO

LITERATURE REVIEW

2.1 NATURAL PRODUCT AS ANTI-CANCER AGENT

Natural products, especially from plants, have been used for the treatment of various diseases for thousands of years. They have also played an important role in the development of several clinical useful anticancer agents (Shoeb, 2006). Over 60% of currently used anti-cancer agents are derived in one way or another from natural sources, including plants, marine organisms and microorganisms (Cragg & Newman, 2005). Example of natural product based drug is doxorubicin or known as Adriamycin derived from *Streptomyces peucetius*, the daunomycin producing microorganism (Arcamone, Cassinelli, Fantini, Grein, Orezzi, Pol, & Spalla, 1969), have been used in oncologic practice since the late 1960. It is a powerful drug in the fight against cancer that includes breast and esophageal carcinomas, osteosarcoma, Kopsi's sarcoma and others (Singal & Iliskovic, 1998). Other plant derivatives used in cancer therapy are listed in Table 2.1.

2.1.1 Secondary Metabolites in Natural Products

Bioactive natural products play a vital role in order to find the novel therapeutic agents. Over 40 % of the medicines have been originated from natural products. Plant and marine sources are among the natural products from the nature and have existed since the beginning of life. This is the evidence that works related to natural products continue to develop and involve the researchers from various scientific backgrounds throughout the world (Hadi, Duru, & Martin-Diana, 2013).

Table 2.1 List of plant derivatives used in cancer therapy.

Semisynthetic analogs of plant derivatives	Species and Genus name	Experiments on various cancer cells	References
Vindesine and vinorelbine	Catharanthus roseus	Leukemia, lymphomas advanced testicular cancer, breast cancer, lung cancer and Kaposi's sarcoma.	Cragg & Newman (2005)
Taxol [®]	Taxus brevifolia Nutt, T. baccata	Metastatic breast, ovarian, lung, prostate cancer and lymphoid malignancies	Kingston (2007)
Taxotere [®]	T. brevifolia Nutt, T. baccata	Used in patients resistant to Paclitaxel	Hait, Rubin, Alli, & Goodin (2007)
Topotecan	Camptotheca acuminate	Epithelial ovarian cancer and small cell lung cancer	Creemers, Bolis, Gore, Scarfone, Lacave, Guastalla, Despax, Favalli, Kreinberg, & Van Belle (1996)
Irinotecan Exatecan	C. acuminate C. acuminate	Metastatic and colorectal cancer Potential anti-tumor activity both <i>in vitro</i> and <i>in vivo</i>	Fuchs, Mitchell, & Hoff (2006) Ishii, Iwahana, Mitsui, Minami, Imagawa, Tohgo, & Ejima (2000)
Beberine	Hvdrastis canadensis L., Berberineeris sp & Arcungelisia flaw	Osteosarcoma, lung, liver, prostate and breast cancer	Patil, Kim, & Jayaprakasha (2010)
Beta-lapachone	Tabebuia avellanedae	Breast, prostate, pancreatic cancer and promyelocytic leukemia	Li, Li, Yu, & Pardee (2000)
Curcumin	Curcuma longa	Colorectal cancer, multiple myeloma and pancreatic cancer	Sa, Das, Banerjee, & Chakraborty (2010); Goel, Kunnumakkara, & Aggarwal (2008)

Semisynthetic analogs of plant derivatives	Species and Genus name	Experiments on various cancer cells	References
Diadzein and Genistein	Lupinus species, Vicia faba, Glycine max, Psoralea corylifolia	Genistein inhibits ovarian and breast cancers and also chemically induced cancers of stomach, bladder, lung, prostate, colon and blood.	Kaufman, Duke, Brielmann, Boik, & Hoyt (1997); Moon, Wang, & Morris (2006); Dixon & Ferreira (2002)
Flavopiridol	Amoora rohituka and Dysoxylum binectariferum	Colorectal, non-small cell lung cancer, renal cell carcinoma, non-Hodgkin's lymphoma, chronic lymphocytic leukemia and also solid tumor	Mans, da Rocha, & Schwartsmann (2000)
Harringtonine and Homoharringtonine	Cephalotaxus harrintonia, Cephalotaxus hainanensis and Cephalotaxus qinensis	Acute myeloid leukemia and chronic myeloid leukemia	Cragg & Newman (2005); Efferth, Li, Konkimalla, & Kaina (2007)
Pandimex TM	Saponins of gengseng	Advance cancer of breast, colon-rectum, lung, pancreas and solid tumor	Pan, Chai, & Kinghorn (2010)
Perillyl alcohol	Many plant species like mints, cherries, lavenders and many others	Non small lung cancer, prostate cancer, colon and breast cancer	Pan et al. (2010)
Schischkinnin	Centaurea schischkinii	Colon cancer lines in vitro	Shoeb, Celik, Jaspars, Kumarasamy, MacManus, Nahar, Thoo-Lin, & Sarker (2005)
Silvestrol	Aglaia foveolata Panell	Prostate, breast and lung cancers	Kinghorn, Carcache de Blanco, Chai, Orjala, Farnsworth, Soejarto, Oberlies, Wani, Kroll, & Pearce (2009); Kim, Hwang, Su, Chai, Mi, Kinghorn, Wild, & Swanson (2007)