# THE EFFECTS OF TAMARINDUS INDICA SEED EXTRACT AS ANTI-SNAKE VENOM AGAINST DABOIA RUSSELLI, NAJA KAOUTHIA AND OPHIOPHAGUS HANNAH IN MICE

BY

# SORAYA ISMAIL

A thesis submitted in fulfilment of the requirement for the degree of Doctor of Philosophy (Medical Sciences)

Kulliyyah of Medicine International Islamic University Malaysia

**JUNE 2017** 

### **ABSTRACT**

Globally, snakebite cases are estimated to be around 5 million annually affecting mainly the residents of poorer counties like Africa and Asia, and in 2009 WHO has categorised it as a 'neglected tropical disease'. Currently the standard treatment for snake envenomation is the use of anti-snake venom (ASV) therapy. However this is expensive and not readily available in smaller hospitals in the developing world. Herbal medicine has been and is still in use in some cultures for the treatment of snakebite and one such plant is Tamarindus indica. This plant is found in many countries where snake envenomation is also prevalent. This study was conducted to evaluate the potential of using *T. indica* seed extract (TSE) to inhibit the effects of snake venom of three snakes; namely Naja kaouthia, Ophiophagus hannah and Daboia russelli. The testa of tamarind seed was used and it underwent ethanolic soxhlet extraction to obtain TSE. The inhibition of the activity of the following enzymes i.e phospholipase A<sub>2</sub> (PLA<sub>2</sub>), proteinase and phosphomonoesterase (PME) in vitro by the three snake venoms with TSE was studied. SDS-PAGE experiment was conducted to observe the effects of TSE on venom proteins. In vivo acute subcutaneous (SC) toxicity of TSE in ICR mice was conducted. Study on the inhibition of lethality was conducted on each of the three snake venoms when SC TSE was injected into mice. Venom concentration and site were fixed but TSE concentration, time and site of injection were manipulated. Findings from venom enzymatic inhibition studies showed that, TSE was able to significantly reduce (p<0.05) all three venom enzymatic activities i.e PLA<sub>2</sub>, proteinase and PME. SDS-PAGE experiment showed that venom protein bands were disrupted when venom reacted with TSE. No signs of toxicity were observed over a period of 4 weeks when mice were exposed to SC TSE 60 mg/20 g body weight except for skin ulcers. Histological examination on liver, both kidneys and skin at the site of SC injection showed no changes compared to the control group injected with SC distilled water. TSE was able to increase the survival rate of ICR mice when exposed to each of the three snake venoms regardless of the site of injecting SC TSE. Mice injected with N. kaouthia or D. russelli venom, had increased 24 hour survival rate when SC TSE was given at 15 minutes; and of mice injected with O. hannah venom the 24 hour survival rate increased with higher TSE concentration when given sooner. In conclusion, SC TSE was safe to be injected up to 60 mg/20 g and has the potential to delay the effects of venom from N. kaouthia, O. hannah and D. russelli.

# خلاصة البحث

تقدر حالات لدغ الأفاعي على الصعيد العالمي بحوالي 5 ملايين سنويا، مما يؤثر بشكل كبير على سكان البلدان الفقيرة مثل افريقيا وآسيا. وقد صنفت منظمة الصحة العالمية عام 2009 لدغ الأفاعي ضمن "أمراض المناطق الاستوائية المهملة". وحاليا فإن المعيار القياسي للعلاج من حالات التسمم من اللدغات السامة يكمن في استخدام ترياق مضاد لسموم الأفاعي . ولكن هذا أمر مكلف و غير متوفر بسهولة في المستشفيات الصغيرة في العالم النامي. وقد استخدمت الأدوية العشبية في الماضي لعلاج لدغات الأفاعي ومن بينها نبات التمر هندي كأحد هذه الوسائل. وقد تم العثور على هذا النبات في العديد من البلدان حيث التأثر بزعاف الحشرات السامة لا تزال سائدة أيضا. وقد أجريت هذه الدراسة لمعرفة إمكانية استخدام مستخلص بذور روسلي. لقد استخدم غلاف بذور التمر الهندي والذي تم استخراجه عن طريق السوكسليت المثيلي وذلك للحصول على مستخلص التمر هندي. ولقد استخدم في المختبر مستخلص بذور وذلك للمصول على مستخلص التمر هندي. ولقد استخدم في المختبر مستخلص بذور وذلك لسموم ثلاثة أفاعي. وقد أجري الفصل الكهربائي لهلام متعدد الأكريلاميد لملاحظة وذلك السموم ثلاثة أفاعي. وقد أجري الفصل الكهربائي لهلام متعدد الأكريلاميد لملاحظة تأثير مستخلص التمر هندي على بروتينات سموم الأفاعي. هذا وقد أجريت دراسة التأثير الحاد لحقن الفئر ان تحت الجلد بمستخلص التمر هندي.

وقد تمت دراسة تثبيط فتك سمية ثلاثة أفاعي بحقن مستخلص التمرهندي تحت جلد فئران التجارب. وقد ثُبتت جرعات سموم الأفاعي وكذلك أماكن حقنها؛ مع تغيير جرعات، ووقت وأماكن حقن مستخلص التمر هندي. وأظهرت نتائج تثبيط فاعلية انزيمات سموم الأفاعي على مقدرة مستخلص التمر هندي على اضعاف تأثير سُمّية انزيمات الأفاعي الثلاثة (انزيمات الفوسفوليبد أ2، بروتيوليتك، والفوسفومونواستيريز). وقد أظهرت تجارب فصل الرحلان الكهربائي لهلام الأكريلاميد عُطل رحلان سموم الأفاعي عند استعمال مستخلص التمر هندي. ولمدة 4 أسابيع لم يُلاحظ ظهور أعراض لتمسمم الفئران المحقونة بمستخلص التمر هندي بجرعة 60 ملج/20ملج باستثناء التقرحات الجلدية. ولم يُظهر الفحص النسيجي للكبد والكُليتن ومواضع الحقن تحت الجلد أي تغييرات بالمقارنة بفئران مجموعة التحكم والتي حقنت بالماء المُقطّر . وقد رفع مستخلص التمر هندي معدل بقاء الفئر ان على قيد الحياة عند تعرضها لسموم كل من الثلاثة أفاعي بغض النظر عن موقع الحقن تحت الجلد. ارتفع معدل بقاء الفئر ان على قيد الحياة لمدة 24 ساعة التي حُقنت بسموم أفعى الأميوفيجس هانا وذلك عند التعجيل بحقنها بجرعة عالية من مستخلص التمر هندي. أما الفئر إن التي حُقنت بسموم أفعى الناجاكوثيا والدابيا روسلي فقد زاد معدل بقائها على قيد الحياة لمدة 24 ساعة عند حقنها بمستخلص التمر هندي كل 15 دقيقة. و إستنتجت الدر إسة أن لدي مستخلص التمر هندي قدرة على تأخير آثار سموم أفاعي الأميوفيجَس هانا، والناجاكوثيا والدابيا روسلى عند حقنها تحت الجلد بمستخلص التمر هندى بجرعات تتراوح بين 60 ملج الى 20 ملج.

# **APPROVAL PAGE**

Pakeer Oothuman Syed Ahamed Supervisor	-
	-
Khin Maung Maung Co-Supervisor	
M. Taher Bakhtiar Internal Examiner	-
Stephen Ambu External Examiner	-
Rafidah Hanim Mokhtar External Examiner	-
Wan Mohd Azizi Wan Sulaiman Chairman	-

# **DECLARATION PAGE**

I hereby declare that this thesis is the result of my	y own investigations, except where
otherwise stated. I also declare that it has not been p	reviously or concurrently submitted
as a whole for any other degrees at IIUM or other in	nstitutions.
Soraya binti Ismail	
Signature	Date

# INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

# DECLARATION OF COPYRIGHT AND AFFIRMATION OF FAIR USE OF UNPUBLISHED RESEARCH

# THE POTENTIAL OF TAMARINDUS INDICA SEED EXTRACT AS ANTI-SNAKE VENOM AGAINST DABOIA RUSSELLI, NAJA KAOUTHIA AND OPHIOPHAGUS HANNAH IN MICE

I declare that the copyright holders of this thesis are jointly owned by the student and IIUM.

Copyright © 2017 Soraya binti Ismail and International Islamic University Malaysia. All rights reserved.

No part of this unpublished research may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder except as provided below

- 1. Any material contained in or derived from this unpublished research may be used by others in their writing with due acknowledgement.
- 2. IIUM or its library will have the right to make and transmit copies (print or electronic) for institutional and academic purposes.
- 3. The IIUM library will have the right to make, store in a retrieved system and supply copies of this unpublished research if requested by other universities and research libraries.

By signing this form, I acknowledged that I have read and understand the IIUM Intellectual Property Right and Commercialization policy.

Affirmed by Soraya binti Ismail		
Signature	 Date	

This thesis is dedicated to my loving family;

To my respectful and loving parents; Haji Ismail Bazain and Allahyarharmah Hajjah Khadijah Haji Mohd Said, thank you very much for allowing me to pursue my dreams and supporting me all the way. Without your blessing I will never be where I am today. Thank you for believing in me.

To my sweet husband; Dr. Ahmad Faidzal Othman, in you soft embrace I find the strength; and in your kind words I find the wisdom. I am blessed to have you in my life.

To my adorable children; Ahmad Nadzrein Iskandar, Ahmad Faredz Idham & Ahmad Redza Iezlan, you are my pride and joy. Soar the sky and live your dream.

In all of you I find love and sincerity, warmth and comfort, peace and serenity.

Thank you for being part of the worderful journey of my life. I love you all.

### **ACKNOWLEDGEMENT**

In the name of Allah, most gracious, most merciful.

Alhamdulillah, praise be to Allah the Almighty, for His will this thesis is able to be completed. My sincere appreciation to my respected supervisor; Professor Dr. Pakeer Oothuman b Syed Ahamed and co-supervisor; Dr. Khin Maung Maung. Thank you very much for the knowledge shared and guidance given in seeing me through PhD journey. My sincere appreciation too to Assoc. Professor Imad Matloub Dally Al Ani for assisting me with my histopathological examination and Assoc. Professor Dr. Emad Mohamad Nafie Abdel Wahab for helping write the Arabic translation of the abstract.

Many thanks to Encik Ramali b Awang; Director, Department of Landscape, Kuantan Municipal Council and Teruntum Mini Zoo staff for allowing and facilitating the milking of snake venom from the snakes available. Many thanks too to the laboratory staff from Kulliyyah of Pharmacy, Kulliyyah of Science and Kulliyyah of Medicine for assisting in laboratory work conducted.

Many thanks to the Research Management Center, IIUM for funding the study (EDW B 14-211-1096). Lastly, my warmest appreciation to all my lecturers, staff, fellow postgraduate students and friends alike who have taught me and supported me along the way. May Allah bless all of you.

# TABLE OF CONTENTS

Abstract			ii
Arabic Tra	nslation	1	iii
Copyright	Page		vi
Dedication			vii
Acknowled	dgemen	t	viii
List of Figu	ures		xix
List of Abb	oreviatio	ons	xxxii
List of Def	initions		xxiii
CHAPTE		: INTRODUCTION	
1.1		ground	
1.2		ication of Study	
1.3		arch Question	
1.4	• •	thesis	
1.5		ctive	
	1.5.1	General Objective	6
	1.5.2	Specific Objectives	6
		): LITERATURE REVIEW	
2.1		es	
		Identification of Snakes	
	2.1.2	Snakes in Malaysia	
2.2		e Envenomation	
		Global Prevalence of Snake Envenomation	
	2.2.2	Prevalence of Snake Envenomation in Malaysia	
2.3	_	nosis of Envenomation	
	2.3.1	Symptoms of Snake Envenomation	
2.4		ment of Snake Envenomation	
2.5		e Venom	
		Elapidae Venom	
	2.5.2	Viperidae Venom	
	2.5.3	Composition of Snake Venom	
_	2.5.4	Snake Venom Proteins	
2.6		al Medicine and Snakebite Management	
	2.6.1	Plants with Anti-snake Venom Properties	
	2.6.2	Phytoconstituents in Traditional Plants Active Against Snake	
	_	Envenomation	
2.7		urindus indica	
	2.7.1	T. indica Pharmacological Properties	
	2.7.2	T. indica Whole Seed	
	2.7.3	T. indica Seed Testa	47

2.8	Method of Plant Extraction	48
2.9	Method of Evaluating Extract for Anti-Snake Venom Properties –	
	in vitro	50
2.1	0 Method of Evaluating Extracts for Toxicity – <i>in vivo</i>	52
	2.10.1 <i>T. indica</i> Seed Extract - Acute Subcutaneous Toxicity Study	
2.1	1 Method of Evaluating Extract For Anti-snake Venom Properties –	
	in vivo	53
CHAPTI	ER THREE: METHODOLOGY	54
3.1		
3.2		
3.3	· · · · · · · · · · · · · · · · · · ·	
3.4	· · · · · · · · · · · · · · · · · · ·	
٥.	3.4.1 Animal Grouping	
3.5	1 0	
3.6		
3.7		
J.,	3.7.1 Harvesting T. indica Fruit and Collecting Seeds	
	3.7.2 <i>T. indica</i> Seed Extraction	
2 (	3.7.2 1. <i>Indica</i> Seed Extraction	
3.0	3.8.1 SDS-PAGE Experiment	
	3.8.2 Inhibition of Venom PLA <sub>2</sub> Enzymatic Activities	
	3.8.3 Inhibition of Proteinase Enzymatic Activity	
3.9	3.8.4 Inhibition of Phosphomonoesterase Enzymatic Activity  Methods – Phase 2 – Tamarind Seed Extract <i>In vivo</i> Toxicity	04
3.5	Experiment	65
2 1	10 Methods – Phase 3 – <i>In vivo</i> Inhibition of Lethality Experiment	
3.1	3.10.1 Venom Selection	
	3.10.2 Venom Preparation	
	3.10.4 Inhibition of Lethality	69
CILADTI	ED EATID, DILACE 1 IN WITTO EXPEDIMENT VI	
	ER FOUR: PHASE 1 – <i>IN VITRO</i> EXPERIMENT - VE YMATIC INHIBITION EXPERIMENT	
<b>ENZ</b> : 4.1		
4.1		
4.2		
	23	74
4.0	4.2.4 Conclusion for SDS-PAGE	
4.3	Phospholipase A <sub>2</sub> Enzyme Inhibition	//
	4.3.1 Methodology for Phospholipase A <sub>2</sub> Inhibition	/8
	4.3.2 Results for Phospholipase A <sub>2</sub> Inhibition	
	4.3.3 Discussion for Phospholipase A <sub>2</sub> Inhibition	
	4.3.4 Conclusion for Phospholipase A <sub>2</sub> Inhibition	
4.4	Proteinase Enzyme Inhibition	
	4.4.1 Methodology for Proteinase Inhibition	
	4.4.2 Results for Proteinase Inhibition	
	4.4.3 Discussion for Proteinase Inhibition	89

	4.4.4 Conclusion for Proteinase Inhibition	90
4.5	Phosphomonoesterase Enzyme Inhibition	90
	4.5.1 Methodology for Phosphomonoesterase Inhibition	
	4.5.2 Results for Phosphomonoesterase Inhibition	
	4.5.3 Discussion for Phosphomonoesterase Inhibition	
	4.5.4 Conclusion for Phosphomonoesterase Inhibition	
4.6	<u>-</u>	
	Experiment	97
		> ,
CHAPTE	R FIVE: PHASE 2 – TAMARIND SEED EXTRACT TOX	ICITY
EXPE	RIMENT – IN VIVO EXPERIMENT	98
5.1	Introduction	98
5.2	Methodology	98
	5.2.1 Animals	98
	5.2.2 Acute Subcutaneous Toxicity Experiment	98
	5.2.3 Histological Examination	
	5.2.4 Statistical Analysis	
5.3	Results	
	5.3.1 Mortality Rate	
	5.3.2 Signs of Toxicity	
	5.3.3 Body Weight	
	5.3.4 Gross Histological Observation	
	5.3.5 Microscopic Histological Changes	
5.4		
7 7	Conclusion	119
5.5	Conclusion	119
CHAPTE	R SIX: PHASE 3 - LETHALITY EXPERIMENT - IN	VIVO
CHAPTE	R SIX: PHASE 3 - LETHALITY EXPERIMENT – <i>IN</i>	<i>VIVO</i> 121
CHAPTE EXPE 6.1	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT Introduction	<i>VIVO</i> <b>121</b> 121
CHAPTE EXPE 6.1	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT Introduction Venom Preparation	<i>VIVO</i> <b>121</b> 121 122
<b>CHAPTE EXPE</b> 6.1  6.2	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT  Introduction  Venom Preparation	<i>VIVO</i> 121 121 122 122
CHAPTE EXPE 6.1	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT	<i>VIVO</i> 121 122 122 123
<b>CHAPTE EXPE</b> 6.1  6.2	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT	VIVO 121 121 122 122 123 125
<b>CHAPTE EXPE</b> 6.1  6.2	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT  Introduction Venom Preparation	VIVO 121 122 122 123 125 128
CHAPTE EXPERIENCE 6.1 6.2 6.3	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT	VIVO 121 122 122 123 125 128 130
<b>CHAPTE EXPE</b> 6.1  6.2	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN  RIMENT	VIVO 121 122 122 123 125 128 130 131
CHAPTE EXPERIENCE 6.1 6.2 6.3	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN  RIMENT	VIVO 121 122 122 123 125 128 130 131 132
CHAPTE EXPERIENCE 6.1 6.2 6.3	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN  RIMENT	VIVO 121 122 122 123 125 128 130 131 132 135
CHAPTE EXPERIENCE 6.1 6.2 6.3	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN  RIMENT	VIVO 121 122 122 123 125 128 130 131 132 135 143
CHAPTE EXPERIENCE 6.1 6.2 6.3	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN  RIMENT	VIVO 121 122 122 123 125 128 130 131 132 135 143 149
CHAPTE EXPERIENCE 6.1 6.2 6.3	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN  RIMENT	VIVO 121 122 122 123 125 128 130 131 132 135 143 149 150
CHAPTE EXPERIENCE 6.1 6.2 6.3	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT	VIVO 121 122 122 123 125 128 130 131 132 135 143 149 150 150
CHAPTE EXPERIENCE 6.1 6.2 6.3	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT	VIVO 121 122 122 123 125 128 130 131 132 135 143 149 150 151
CHAPTE EXPERIENCE 6.1 6.2 6.3	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT	VIVO 121 122 122 123 125 128 130 131 132 135 143 149 150 151 163
CHAPTE EXPERIENCE 6.1 6.2 6.3	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT	VIVO 121 122 122 123 125 128 130 131 132 135 143 149 150 151 163
CHAPTE EXPE 6.1 6.2 6.3 6.4 6.5	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT	VIVO 121 122 122 123 125 128 130 131 132 135 143 149 150 151 163 165
CHAPTE EXPE 6.1 6.2 6.3 6.4 6.5	R SIX: PHASE 3 - LETHALITY EXPERIMENT – IN RIMENT	VIVO 121 122 122 123 125 128 130 131 132 135 143 149 150 151 163 165

7.3 Sug	gested Further Studies
REFERENCES	
APPENDIX A:	PHYLOGENY OF SNAKES - ORDER SQUAMATA, SUBORDER SERPENTES (OPHIDIA)181
APPENDIX B:	HIGHER TAXA IN EXTANT REPTILES 182
APPENDIX C:	THE ELAPID AND VIPERID SNAKES OF MALAYSIA PENINSULAR, SABAH, SARAWAK AND BORNEO) 184
APPENDIX D:	DICHOTOMOUS IDENTIFICATION KEY TO THE VENOMOUS TERRESTRIAL SNAKES IN MALAYSIA 185
APPENDIX E:	SNAKEBITE MANAGEMENT GUIDE FOR HEALTHCARE PROVIDERS IN MALAYSIA 187
APPENDIX F:	GUIDE TO ANTIVENOM APPROPRIATE FOR MALAYSIA
APPENDIX G:	ANTISNAKE VENOM STOCK PILE CALCULATION 193
APPENDIX H:	LIST OF MEDICINAL PLANTS WITH ANTI-SNAKE VENOM PROPERTIES
APPENDIX I:	VERNACULAR NAMES OF TAMARINDUS INDICA 198
APPENDIX J:	PLANT VOUCHER 202
APPENDIX K:	PREPARATION OF EGG YOLK SUSPENSION AS SUBSTRATE FOR PHOSPHOLIPASE A2 ASSAY 204
APPENDIX L:	ANIMAL ETHICS APPROVAL
APPENDIX M:	ACUTE SUBCUTANEOUS TOXICITY ASSESSMENT OF TAMARIND SEED EXTRACT IN ICR MICE 206
APPENDIX N:	XYLAZINE : KETAMINE ANAESTHESIA 207
APPENDIX O:	PREPARATION FOR 10% FORMAL SALINE 208
APPENDIX P:	TAMARIND SEED TOXICITY STUDY – SKIN CHANGES AND BODY WEIGHT CHANGES 209
APPENDIX Q:	3 MINUTES THESIS COMPETITION, NATIONAL LEVEL
APPENDIX R:	PUBLICATION IN INTERNATIONAL MEDICAL JOURNAL MALAYSIA (IMJM), VOLUME 15, 2016 211
APPENDIX S:	PUBLICATION IN INTERNATIONAL MEDICAL JOURNAL MALAYSIA (IMJM), VOLUME 15, 2016 212

# LIST OF TABLES

Table 1.1	Global prevalence of snake envenomation (Chippaux, 2008)	3
Table 2.1	General morphological characteristics to identify snakes	8
Table 2.2	Venomous snakes in peninsular Malaysia (Das et al., 2015; Lim, 1982)	12
Table 2.3	Some examples of medically significant Elapidae in Malaysia (Das et al., 2015)	14
Table 2.4	Examples of medically significant Viperidae in Malaysia (Das et al., 2015)	14
Table 2.5	General morphological differences between Elapidae and Viperidae found in Malaysia (Das et al., 2015)	15
Table 2.6	Summary of the published research on snakebite cases in Malaysia.	20
Table 2.7	Types of toxin in snake venom (Tan, 2005)	23
Table 2.8	The general signs and symptoms of snake envenomation (Ismail, 2015)	24
Table 2.9	Some of the proteins and enzymes present in snake venoms (Tan, 2005)	35
Table 2.10	Top ten families of plants with the largest number of species used as anti-snake venom (Molander et al., 2012)	37
Table 2.11	An overview of the size and taxonomic distribution of medicinal plants used in top 5 countries (Molander et al., 2012)	39
Table 2.12	Phenolic compounds in plants and the corresponding inhibitory actions on the effects of snake envenomation (Guimaraes et al., 2014)	40
Table 2.13	List of isolated constituents with anti-snake venom activity (Makhija & Khamar, 2010)	41
Table 2.14	Different parts of the <i>Tamarindus indica</i> plant and its pharmacological uses. (Isha & Milind, 2006; Meher et al., 2014)	44

Table 2.15	al., 2010)	47
Table 2.16	Past researches on tamarind seed extract as anti-snake venom	48
Table 2.17	Main methods of plant extraction and the general outline of the procedures (Handa et al., 2008)	49
Table 3.1	Parameter observed for tamarind seed extract <i>in vivo</i> toxicity experiment.	67
Table 3.2	General overview of lethality experiment.	69
Table 4.1	Materials used for SDS-PAGE.	72
Table 4.2	Grouping for venom phospholipase $A_2$ enzymatic and inhibitory experiment.	79
Table 4.3	Grouping for venom proteinase enzymatic and inhibitory experiment.	86
Table 4.4	Venom and tamarind seed extract (TSE) concentration and volume for venom phosphomonoesterase enzymatic and inhibitory experiments	92
Table 4.5	Grouping for venom phosphomonoesterase enzymatic and inhibitory experiments.	92
Table 5.1	Summary of acute subcutaneous tamarind seed extract toxicity experiment activities	102
Table 6.1	Past studies on some snake venom LD <sub>50</sub> in mice	125
Table 6.2	Sequence of tests to establish subcutaneous $LD_{100}$ for $Naja$ kaouthia	126
Table 6.3	Sequence of tests which took place to establish subcutaneous $LD_{100}$ for <i>Ophiophagus hannah</i>	127
Table 6.4	Sequence of tests which took place to establish subcutaneous LD <sub>100</sub> for <i>Daboia russelli</i>	128
Table 6.5	$Naja \ kaouthia$ venom concentration adjustments to achieve $LD_{100}$	129
Table 6.6	Ophiophagus hannah venom concentration adjustments to achieve $LD_{100}$	129
Table 6.7	Daboia russelli venom concentration adjustments to achieve LD <sub>100</sub>	130

Table 6.8	The 24 hour survival rate after receiving treatment with tamarind seed extract following exposure to 11.45 μg/20 g Naja kaouthia venom	135
Table 6.9	The 24 hour survival rate after receiving treatment with tamarind seed extract following exposure to 31.5 $\mu g/20$ g <i>Ophiophagus hannah</i> venom	137
Table 6.10	The 24 hour survival rate after receiving treatment with tamarind seed extract following exposure to 95.0 $\mu g/20$ g Daboia russelli venom	139
Table 6.11	Survival rate after receiving $30 \text{ mg}/20 \text{ g}$ tamarind seed extract treatment $30 \text{ minutes post-exposure}$ to $11.45 \mu\text{g}/20 \text{ g}$ Naja kaouthia venom	141
Table 6.12	Survival rate after receiving $10 \text{ mg}/20 \text{ g}$ tamarind seed extract treatment $30 \text{ minutes post-exposure to } 31.5  \mu\text{g}/20 \text{ g}$ Ophiophagus hannah venom	142
Table 6.13	Survival rate after receiving 30 mg/20 g tamarind seed extract treatment 30 minutes post-exposure to 95.0 µg/20 g <i>Daboia russelli</i> venom	142
Table 6.14	Previous studies on tamarind seed extract with anti-snake venom properties	143
Table 6.15	Previous studies on various plant extract with anti-snake venom properties	144

# LIST OF FIGURES

Figure 1.1	Scientific classification of snakes (adapted from www.itis.gov. Downloaded on 20 Mac 2015)	2
Figure 1.2	Map showing the global distribution of snakebite mortality (Chippaux, 2008)	3
Figure 2.1	Morphological identification of Elapidae/Colubridae head shape and scalation (Ismail, Das, & Weinstein, 2013)	9
Figure 2.2	Morphological identification chracteristics of pit-viper head shape and scalation. (a) Presence of the Loreal pit and elliptical pupil. (b) Triangular shape of head and scalation pattern. (c) Diagramatic illustration of pit viper chracteristics (Gardner, 2005; Ismail et al., 2013).	10
Figure 2.3	Morphological identification of snakes using features of the tail. (a) – (c) are characteristics of venomous snakes and (d) is characteristics of non-venomous snakes. (a) Rattle snake showing its rattle-like tail, (b) 'rattle-like' tail end, (c) single anal plate (i), single pattern scale (ii), (d) divided anal plate (iii) double pattern scale (iv) (taken from www.snake protection.com and www.quora.com on 12 Sept. 2015).	11
Figure 2.4	(a) Diagrammatic illustration of the snake venom gland and venom duct in the fangs. (b) Scanning electron micrograph of snake fangs with deep grooves (Gopalakrishnakone et al., 1990).	16
Figure 2.5	Diagrammatic illustration of Viperidae venom gland. The venom is synthesized at the venom gland (mg). It is then moves to the primary duct (pd) through the accessory gland (ag) and secondary duct (sd). The venom exits into the fang pocket sheath (s) then travels through the fangs groove or canal (f). Red arrows show the flow of venom into fangs (taken from: http://jrscience.wcp.muohio.edu Downloaded on 4 Jan. 2016).	16
Figure 2.6	The snakebite pattern with fang marks to identify species of venomous snake and non-venomous snakes of Malaysia. (taken from http://www.prn.usm.my. Downloaded on 12 Sept. 2015).	
		22

Figure 2.7	Illustration of general symptoms of snake envenomation at different organ systems. (taken from en.wikipedia.org /wiki/File:Snake_bite_symptoms.png. Downloaded on 3 Dec. 2015)	25
Figure 2.8	Anti-snake venoms available in Malaysia imported from QSMI Thailand and CSL Ltd. Australia. ( <i>Pictures taken from http://www.saovabha.com and from 'Pharmacy Bullettin hsmanjung .moh.gov.my'</i> . Downloaded on the 28 Dec. 2015).	30
Figure 2.9	Formula to calculate the holding quantities of anti-snake venom at any one time in one hospital in Malaysia (Ismail, 2013).	31
Figure 2.10	Tamarind tree and fruit used for this study.	43
Figure 2.11	Tamarind used for this study (a) fruit on the tree, (b) ripe tamarind fruit, (c) shell removed from fruit exposing the pulp, and (d) tamarind seed.	45
Figure 3.1	Conceptual framework showing the cause and effect relationship between snake envenomation and the death outcome. Venom enzymes that were studied are herein*.	55
Figure 3.2	Study design and general layout plan of the study.	56
Figure 3.3	General layout of animal grouping for in vivo study.	59
Figure 3.4	Tamarind fruit harvesting process	62
Figure 3.5	Tamarind seed being processed (a-g) to obtain powdered ethanolic extraction	63
Figure 3.6	Mice grouping for the study of tamarind seed extract toxicity experiment	66
Figure 3.7	Dorsal view of a mice illustrating the site where the venom and tamarind extract were given. For same site, both venom and extract were injected at A. For different sites, venom was given at A followed by extract at B.	70
Figure 4.1	Protocol for SDS_PAGE experiment carried out for this study	73

Figure 4.2	Gel from SDS-PAGE showing the effects of tamarind seed extract (TSE) on snake venom protein. Well 1: 17.14 μg <i>Daboia russelli</i> venom only, Well 2: 17.14 μg <i>D. russelli</i> with 4.76 μg TSE, Well 3: 17.14 μg <i>Naja kaouthia</i> venom only, Well 4: 17.14 μg <i>N. kaouthia</i> with 4.76 μg TSE, Well 5: 17.14 μg <i>Ophiophagus hannah</i> venom only, Well 6: 17.14 μg <i>O. hannah</i> with 4.76 μg TSE, Well 7 & 8: empty, Well 9: TSE I (167μg/100μl) only, Well 10: TSE II (250μg/100μl) only.	74
Figure 4.3	Pharmacological effects of snake venom PLA <sub>2</sub> (Mackessy, 2010)	78
Figure 4.4	Venom phospholipase $A_2$ enzymatic and inhibitory experiment protocol	81
Figure 4.5	Figure 4.5 Effect of tamarind seed extract on venom phospholipase A <sub>2</sub> enzyme activity of <i>Ophiophagus hannah</i> , <i>Naja kaouthia</i> and <i>Daboia russelli</i> . Result is the mean of triplicate determination. Vertical lines represent SD. Blue bars represent 100% venom phospholipase A <sub>2</sub> enzyme activity and coloured bars represent the tamarind seed extract inhibited venom phospholipase A <sub>2</sub> enzyme activity.	82
Figure 4.6	Venom proteinase enzymatic and inhibitory experiment protocol.	87
Figure 4.7	Effect of tamarind seed extract on venom proteinase enzyme activity of <i>Ophiophagus hannah</i> , <i>Naja kaouthia</i> and <i>Daboia russelli</i> . Result is the mean of triplicate determination. Vertical lines represent SD. Blue bars represent 100% venom proteinase enzyme activity and coloured bars represent tamarind seed extract inhibited venom proteinase enzyme activity.	89
Figure 4.8	Venom phosphomonoesterase enzymatic and inhibitory experiments protocol.	93
Figure 4.9	Effect of tamarind seed extract on venom phosphomonoesterase enzyme activity of <i>Ophiophagus hannah</i> , <i>Naja kaouthia</i> and <i>Daboia russelli</i> . Result is the mean of triplicate determination. Vertical lines represent SD. Blue bars represent 100% venom phosphomonoesterase enzyme activity and coloured bars represent tamarind seed extract inhibited venom phosphomonoesterase enzyme activity.	94

Figure 4.10	Schematic representation of adenosine generation by venom enzymes from DNA/RNA hydrolysis. Venom enzymes contained in ovals*, and bold letters <sup>#</sup> indicate end products released upon enzyme actions (Mackessy, 2010).	96
Figure 4.11	Schematic representation of adenosine generation from ATP hydrolysis by venom enzymes. Venom enzymes contained in ovals, and bold letters indicate end products released upon enzyme actions (Mackessy, 2010).	96
Figure 5.1	Grouping for acute subcutaneous tamarind seed extract toxicity experiment	100
Figure 5.2	(a) Diagrammatic representation of a mice dorsally illustrating the site where the fur was clipped and tamarind seed extract (TSE) given subcutaneously. (b) and (c) show mice dorsally clipped ready for subcutaneous administration of TSE and their tail marked for the purpose of individual identification.	101
Figure 5.3	Flowchart of tissue processing procedure	105
Figure 5.4	Flowchart of Haematoxylin – Eosin (H&E) staining procedure	107
Figure 5.5	Graph of mean±SD mice skin ulcer surface area for toxicity experiment when tamarind seed extract 45 mg/20 g and 60 mg/20 g were given subcutaneously.	110
Figure 5.6	Photographs showing skin ulcer progression (a-d) in mice treated with TSE 45 mg/20 g over 28 days.	111
Figure 5.7	Photographs showing skin ulcer progression in mice treated with TSE 60 mg/20 g (a-d) over 28 days.	112
Figure 5.8	Graph of mean±SD mice body weight for toxicity experiment when tamarind seed extract 45 mg/20 g and 60 mg/20 g were given subcutaneously.	113
Figure 5.9	Histological examination of Haematoxylin – Eosin (H&E) stained kidney section of ICR mice. Normal kidney tissue architecture seen in all images. There were mild congestion of blood in interstitial tissue (b-d). B - Bowman's capsule, CT – convoluted tubule. Magnification 20x.	115
Figure 5.10	Histological examination of Haematoxylin – Eosin (H&E) stained liver section of ICR mice. Normal liver architecture seen in all images. Mild hydropic changes seen as blood congested in CV and vacuolation of hepatocyte (c). CV –	
	central vein, H – hepatocyte. Magnification 20x.	116

Figure 5.11	Histological examination of Haematoxylin – Eosin (H&E) stained skin sections of ICR mice. Normal skin architecture seen in all images. Magnification 20x.	117
Figure 6.1	Light yellow colour of lyophilized <i>Daboia russelli</i> venom from Myanmar.	122
Figure 6.2	(a) Venom milking process of <i>Naja kaouthia</i> by zoo staff, (b) Lyophilized <i>N. kaouthia</i> venom, (c) Venom milking process of <i>Ophiophagus hannah</i> , (d) Lyophilized <i>O. hannah</i> venom.	123
Figure 6.3	Dorsal view of a mice illustrating the site where the venom and tamarind seed extract (TSE) were injected subcutaneously. For same site: both venom and TSE given at (A). For different sites: venom was given at (A) and TSE was given at (B).	132
Figure 6.4	Mice grouping to study the inhibition of venom lethality with fixed venom concentration and fixed treatment interval but tamarind seed extract treatment concentration changed.	133
Figure 6.5	Mice grouping to study the inhibition of venom lethality with fixed venom concentration and fixed tamarind seed extract treatment concentration but treatment time interval changed.	134
Figure 6.6	Mice grouping to study the inhibition of venom lethality after receiving TSE 30 mg/20 g treatment 30 minutes post-exposure to venom	134
Figure 6.7	Grouping of mice for histological examination to observe the effects of TSE 30 mg/20 g in mice injected with venom.	150
Figure 6.8	Figure 6.8 Histological examination of Haematoxylin – Eosin (H&E) stained kidney section of ICR mice exposed to <i>Naja kaouthia</i> . CT – convulated tubule; B – Bowman's capsule. Magnification 40x.	152
Figure 6.9	Histological examination of Haematoxylin – Eosin (H&E) stained kidney section of ICR mice exposed to <i>Ophiophagus hannah</i> . CT – convulated tubule; B – Bowman's capsule. Magnification 40x.	153
Figure 6.10	Histological examination of Haematoxylin – Eosin (H&E) stained kidney section of ICR mice exposed to <i>Daboia russelli</i> . No specimen was available for the venom only group at 1 week since all mice died within 48 hours of exposure to venom. CT – convulated tubule; B – Bowman's capsule.	
	Magnification 40x.	154

Figure 6.11	Histological examination of Haematoxylin – Eosin (H&E) stained liver section of ICR mice injected with <i>Naja kaouthia</i> venom. CV – central vein. Magnification 40x.	156
Figure 6.12	Histological examination of Haematoxylin – Eosin (H&E) stained liver section of ICR mice injected with <i>Ophiophagus hannah</i> venom. CV – central vein. Magnification 40x.	157
Figure 6.13	Histological examination of Haematoxylin – Eosin (H&E) stained liver section of ICR mice injected with <i>Daboia russelli</i> venom. No specimen was available for the venom only group at 1 week since all mice died within 48 hours of exposure to venom. CV – central vein. Magnification 40x.	158
Figure 6.14	Histological examination of Haematoxylin – Eosin (H&E) stained skin section of ICR mice injected with <i>Naja kaouthia</i> venom. Magnification 40x.	160
Figure 6.15	Histological examination of Haematoxylin – Eosin (H&E) stained skin section of ICR mice injected with <i>Ophiophagus hannah</i> venom. Magnification 40x.	161
Figure 6.16	Histological examination of Haematoxylin – Eosin (H&E) stained skin section of ICR mice injected with <i>Daboia russelli</i> venom. No specimen was available for the venom only group at 1 week since all mice died within 48 hours of exposure to venom. Magnification 40x.	162

# LIST OF ABBREVIATIONS

AMSEM Advanced workshop on marine and snake envenomation

management

H & E Haematoxylin - Eosin

HPLC High performance liquid chromatography

IM Intramuscular

IV Intravascular

LD<sub>50</sub> 50% of lethal dose

LD<sub>100</sub> 100% of lethal dose

MHD Minimum haemorrhagic dose

MPT *Mimosa pudica* extract (*Mimosa pudica* tannin)

OECD Organization for Economic Co-operation and Development

PLA<sub>2</sub> Phospholipase A<sub>2</sub>

PME Phosphomonoesterase

rpm Revolution per minute

SC Subcutaneous

SDS-PAGE Sodium dodecyl sulphate-polyacrylamide gel electrophoresis

SMVPs Snake venom metalloproteinases

TSE Tamarind seed extract

### LIST OF DEFINITIONS

### Antivenom

A purified fraction of immunoglobulins or its fragments fractionated from the plasma of animals that have been immunized against venom from one or several species of snakes

### Venom

The toxic secretion of a specialized venom gland which in the case of the snake, it is delivered through the fangs and provokes deleterious effects. Venoms usually comprise many different protein components of variable structure and toxicity. Venom enters the victims via 'injection'.

### Toxin

A toxic substance, especially a protein, which is produced by living cells or organism and is capable of causing disease when introduced into the body tissues. It is also often capable of introducing neutralizing antibodies or antitoxins. Toxin enters the victims via ingestion.

Calloselasma rhodostoma

Synonym to i) *Agkistrodon rhodostoma* Common name: Malayan pit-viper

Daboia russelli

Synonym to: Vipera russelli

Common name: Russell's viper, Chain viper

Daboia russelli siamensis Synonym to: Vipera russelli

Common name: Eastern Russell's viper, Siamese Russell's viper

Naja kaouthia

Synonym to: Naja naja kaouthia, Naja-naja

Common name: Monocled cobra

It was formerly treated as a subspecies of Indian cobra (Naja naja)

Naja sumatrana

Synonym to: *Naja naja sputatrix* Common name: Spitting cobra

Ophiophagus hannah Synonym to: Hamadryad Common name: King cobra

# **CHAPTER ONE**

## INTRODUCTION

### 1.1 BACKGROUND

Snakes have been the subject of fascination, fear and myths all through history. In ancient Egyptian times, the cobra was worshipped and its replica was used to decorate the crowns of the Pharaohs (Belluccio & Johnson, 1992). In ancient Greek too, Asclepius the demi-god of medicine carried a pole with two snakes entwined on it (Wilcox & Whitham, 2003). The Greek mythology associates snake with being deadly and dangerous. The infamous Medusa had snakes in place of hair (Graves, 1990). In Hindu mythology, the cobra occupies the revered position i.e around Lord Shiva's neck and is protected from enemy, eagle or garuda (Lochtefeld, 2002). In ancient Chinese astrology, the snake represents a month in the Chinese zodiac calendar (Eberhard, 2006).

The scientific classification of snake is shown below in Figure 1.1. The pylogeny of snakes have gone through many updates through the years. The latest update uses snake classifications by Pyron et al (2011). This classification is available in Appendix A.

Globally, over 3400 species of snakes have been documented (Uetz & Hosek, 2015a). From that, peninsular Malaysia has over 141 known species of land and sea snakes. Of these, 26 species of land snakes (Das, Ahmed, & Liat, 2015) and 21 species of sea snakes are venomous (Lim, 1982). The medically significant venomous snakes