COPYRIGHT[©] INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

ANTICANCER STUDY OF PORCUPINE BEZOAR EXTRACTS ON HUMAN BREAST AND LUNG CANCER CELLS

BY

AL'AINA YUHAINIS BT FIRUS KHAN

A thesis submitted in fulfilment of the requirement for the degree of Master Degree of Health Science

Kulliyyah of Allied Health Science International Islamic University Malaysia

JULY 2017

ABSTRACT

Breast and lung cancer have highest mortality rate among female and male respectively. Even though with the availability of advanced treatment, the mortality rate is still increasing. Thus, this study aims to elucidate the potential Malayan porcupine bezoar (PB) in exhibiting anticancer effects on the breast (MCF7) and lung (A549) cancer cells. Porcupine Bezoar extracts were tested with optimized $IC5_0$ for its potential ability in inhibiting cell growth, changing morphology, inducing apoptosis and cell arrest. The PB extract showed the growth inhibitory effect with IC_{50} value at 19.0 µg/ml (MCF7) and 13.5 µg/ml (A549) respectively. The morphology assessment upon PB extract treatment at 72 hours displayed possible apoptosis features of pyknosis and karyorrhexis. Further investigation with Annexin-V/7AAD revealed PB treated cell induced early and late apoptosis with a presence of dead cells in 72 hours. Further test has shown, PB may have altered cell cycle regulation and arrest in G1 for both cells. Possible signaling pathway at a molecular level for both cells was investigated. The result revealed induction of apoptosis following intrinsic pathway by initiating the release of *Cytochrome C* from mitochondria thereby activating caspase cascade. The above claims were supported by downregulation of Bcl-2 which suggest PB extracts triggered apoptosis. Furthermore, the investigation was conducted on cell cycle regulator genes, revealed the up-regulation of p21 and down-regulation of cyclin D suggesting cell arrest in G1 phase. Additionally, finding demonstrate PB extracts was selective in inducing significant effect on cancer cells MCF7 and A549 compared to normal fibroblast 3T3-L1 and HGF-1. This research report unveil the potential medicinal properties of porcupine bezoar in anticancer perspectives.

خلاصة البحث

أعلى معدل وفيات بين الإناث والذكور، سرطان الثدي وسرطان الرئة على التوالي. وعلى الرغم من توافر العلاج المتقدم، فإن معدل الوفيَّات ما زال في ازدياد. و هكذاً، تهدف هذه الدراسة إلى توضيح فعالية (porcupine bezoar (PB الماليزي في تأثيره كمضاد لسرطان الثدي (MCF7) والرئة(A549) . تم اختبار مستخلصات porcupine bezoar مع IC₅₀ الأمثل لقدرتها المحتملة في تثبيط نمو الخلايا السرطانية، وتغيير التشكل، وتحريض الخلايا على القتل المبرَّمج وتوقيف الخلايا. أظهر مستخلص PB التأثير التثبيطي على نمو الخلايا مع قيمة IC50 عند 19.0 ميكروغرام / مل (MCF7) و 13.5 ميكروغرام / مل (A549) على التوالي. تقييم مورفولوجيا الخلايا المعالجة ب PB خلال 72 ساعة عرض ملامح موت الخلايا المبرمج مع أشكال من pyknosis and karyorrhexis. كشف مزيدا من التحقيق مع أنيكسين 7AAD / V المعالجة ب PB وقتا مبكرا ومتأخرا في موت الخلايا المبرمج مع وجود خلايا ميتة في 72 ساعة. وقد أظهرت اختبارات أخرى، أن PB غيرت تنظيم دورة الخلية وحدث الاعتقال في G1 لكلا من الخلايا. وقد تم التحقيق في مسار الإشارات المحتملة على المستوى الجزيئي لكل من الخلايا. وكشفت النتيجة تحريض موت الخلايا المبرمج بعد المسار الجو هري من خلال الشروع في الافراج عن السيتوكروم C من الميتوكوندريا وبالتالي تفعيل caspase cascade. وقد دعمت النتائج المذكورة أعلاه حدوث تناقص في تنظيم Bcl-2 التي تشير إلى أن مستخلصات PB تسبب في موت الخلايا المبرمج. وتعلاوة على ذلك، أجري التحقيق على الجينات المنظمة لدورة الخلية، وكشف عن تنظيم أعلى من P21 والتناقص في تنظيم cyclin D مما يشير إلى اعتقال الخلايا في مرحلةG1 . بالإضافة إلى ذلك، تعتبر مستخلصات PB انتقائية في إحداث تأثير كبير على الخلايا السرطانية MCF7 و A549 مقارنة مع الخلايا الليفية العادية T3-L1 3 و HGF-1. هذا التقرير البحثي كشف النقاب عن الخصائص الطبية المحتملة ل porcupine bezoar من وجهات النظر المضادة للسر طان.

APPROVAL PAGE

I certify that I have supervised and read this study and that in my opinion, it conforms to acceptable standards of scholarly presentation and is fully adequate, in scope and quality, as a thesis for the degree of Master of Health Science.

Ridhwan Abdul Wahab Supervisor

I certify that I have read this study and that in my opinion it conforms to acceptable standards of scholarly presentation and is fully adequate, in scope and quality, as a thesis for the degree of Master of Health Science.

Radiah Abdul Ghani Internal Examiner

Rapeah Suppian External Examiner

This thesis was submitted to the Department of Biomedical Science and is accepted as a fulfilment of the requirement for the degree of Master of Health Science

> Mohd Shukri Baba Head, Department of Biomedical Science

This thesis was submitted to the Kulliyyah of Allied Health Science and is accepted as a fulfilment of the requirement for the degree of Master of Health Science.

Wan Azdie Bin Mohd. Abu Bakar Dean, Kulliyyah of Allied Health Science

DECLARATION

I hereby declare that this thesis is the result of my own investigations, except where otherwise stated. I also declare that it has not been previously or concurrently submitted as a whole for any other degrees at IIUM or other institutions.

Al'aina Yuhainis Bt Firus Khan

Signature

Date

INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

DECLARATION OF COPYRIGHT AND AFFIRMATION OF FAIR USE OF UNPUBLISHED RESEARCH

ANTICANCER STUDY OF PORCUPINE BEZOAR EXTRACTS ON HUMAN BREAST AND LUNG CANCER CELLS

I declare that the copyright holder of this thesis are jointly owned by the student and IIUM

Copyright © 2016 (Al'aina Yuhainis Bt Firus Khan). All rights reserved.

No part of this unpublished research may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder except as provided below

- 1. Any material contained in or derived from this unpublished research may be used by others in their writing with due acknowledgement.
- 2. IIUM or its library will have the right to make and transmit copies (print or electronic) for institutional and academic purposes.
- 3. The IIUM library will have the right to make, store in a retrieved system and supply copies of this unpublished research if requested by other universities and research libraries.

By signing this form, I acknowledged that I have read and understand the IIUM Intellectual Property Right and Commercialization policy.

Affirmed by Al'aina Yuhainis Bt Firus Khan

Signature

Date

ACKNOWLEDGEMENTS

First and foremost, I thank Allah for giving me the strength and endowing me with health, patience, and knowledge to complete this work. Nothing more to ask from what Allah already gave.

I am grateful to my supervisor Dr. Ridhwan Abdul Wahab for accepting me as his student, guide and advise me throughout the journey without make me fell stress or burden.

Utmost thanks to my parent Firus Khan, Rodziyah Mahmud and all my siblings Farezal, Fazlizul, Fakhrul Ikhzan, Fakhrul Afdhal, Maslinda, Masliza, Aliyaa for supporting me from far.

I acknowledge, with deep gratitude and appreciation, for all microbiology laboratory in KOM staff, ICRACU staff for helping me with technical issues in laboratory throughout the study. Special thanks to Dr which guide me in specific aspect Dr. Hamzah (fluorescence microscope), Dr. Mardhiah (qPCR), and Dr. Radiah (thesis correction). Without Dr's help and guidance, I don't think I can finish my study.

I would like to express my gratitude to friends Vignesh, Tara, Afiqah misran, and Rosyafirah that always with me throughout my master journey for the ups, down, cry and joy we have together. The journey will be boring without all of you.

Not to forget all Dr. Ridhwan students; Master's (Habibah, Shakirah, and Faizah); FYP, 2015 (Syikin, Nazihah, and Syafiqah); and FYP, 2014 (Amy, Thoher, Syafiq, Syiela and Aina) for the memory. Many things I learn from all of you and I appreciate the moments with all of you.

TABLE OF CONTENTS

Abstract	
Abstract in Arabic	
Approval Page	iv
Declaration	
Acknowledgements	
Table of Contents	viii
List of Tables	xii
List of Figures	
List of Abbreviations	XV
List of Symbols	xvi
CHAPTER ONE: INTRODUCTION	1
1.1 Background of the study	
1.2 Statement of the problem	
1.3 Research objectives	
1.4 Research questions	
1.5 Research hypotheses	
1.6 Significance of the study	
1.7 Research framework	
1.8 Chapter summary	
CHAPTER TWO: LITERATURE REVIEW	
2.1 Cancer	
2.1.1 Pathogenesis of cancer	
2.2 Breast and lung cancer	
2.3 Cancer therapy (chemoprevention and anticancer)	
2.3.1 Mechanism of anticancer through cell cycle arrest	
2.3.1.1 Cell cycle phases and checkpoints	
2.3.1.2 Role of p21	
2.3.1.3 Anticancer action of cell cycle arrest	
2.3.2 Mechanism of anticancer through apoptosis	
2.3.2.1 Morphology of cells	28
2.3.2.2 Apoptosis induction and its molecular pathway	
2.3.2.3 Anticancer action of apoptosis induction	
2.4 Porcupine bezoar	
2.4.1 Origins and history	
2.4.2 Medicinal properties of bezoar	
2.5 Chapter summary	43
CHAPTER THREE: MATERIALS AND METHOD	44
3.1 Materials	
3.1.1 Porcupine bezoar	
3.1.2 Cell lines.	
3.1.3 Chemicals and reagents	
C C	

3.1.4	4 Commercial kits and consumables	45
3.1.	5 Laboratory apparatus and equipment	45
3.1.0	5 Computer application and software	45
3.1.7	7 Primers	45
3.2 Met	hodology	45
3.2.	1 Preparation of PB extracts	45
	3.2.1.1 Preparation of stock solution porcupine bezoar extracts.	45
	3.2.1.2 Preparation of working solution of PB extracts for	
	experiments	46
3.2.2	2 Cell culture maintenance	47
	3.2.2.1 Cell culture techniques	47
	3.2.2.2 Thawing cells	47
	3.2.2.3 Growing cell	47
	3.2.2.4 Changing media	48
	3.2.2.5 Subculturing cells	48
	3.2.2.6 Cryopreservation of cells	48
	3.2.2.7 Measuring cell viability using Trypan blue exclusion	
	assay method (TBEA)	
	3.2.2.8 Seeding of cells for treatment	50
3.2.	3 Cell optimization	
3.2.4	4 Determination of 50 % inhibitory concentration (IC ₅₀)	51
	3.2.4.1 MTT assay	
3.2.	5 Anti-proliferative assay	52
3.2.0	6 Morphological observations	53
	3.2.6.1 Morphological analysis by inverted microscope	53
	3.2.6.2 Live/dead analysis under fluorescence microscope	53
	3.2.6.3 Cytoskeleton analysis under fluorescence microscope	54
3.2.7	7 Study in normal cells	54
3.2.3	8 Determination of apoptosis using Annexin-V assay	. 56
	3.2.8.1 Sample preparation	56
	3.2.8.2 Apoptosis analysis	
3.2.9	O Cell cycle analysis using Propidium Iodide Staining	57
	3.2.9.1 Sample preparation	57
	3.2.9.2 Cell cycle analysis	
3.2.	10 Gene expression analysis using Quantitative Polymerase	
	Chain Reaction (qPCR)	58
	3.2.10.1 Generating primer	58
	3.2.10.2 RNA extraction	
	3.2.10.3 Measuring RNA concentration and purity	60
	3.2.10.4 Measuring RNA integrity	
	3.2.10.5 cDNA reverse transcription	
	3.2.10.6 Optimization of annealing temperature and melt curve	
	for primer	61
	3.2.10.7 Optimization amplification efficiency	
	3.2.10.8 Quantitative polymerase chain reaction (qPCR) of	
	the genes	64
	3.2.10.9 Analysing gene expression	
	3.2.10.10 Statistical analysis	

CHAPTER FOUR: RESULTS	66
4.1 Suppressive effects of PB extracts on MCF7 and A549 cells	66
4.1.1 Determination of PB extracts concentration that effectively	
inhibits 50% of cells	66
4.1.2 Effects of PB extract treatment in the IC ₅₀ concentration on	
cell proliferation activity in MCF7 and A549 cells	68
4.2 The effect of PB extracts on normal cells of human gingival	
fibroblast (HGF) and mouse fibroblast (3T3-L1) cells line	70
4.3 Effects of PB extracts on the morphology alteration of A549 and	
MCF7 cells	76
4.3.1 Live /Dead Analysis on MCF7 and A549	76
4.3.2 Phase contrast inverted microscope analysis on MCF7 and	
A549 morphology	
4.3.3 Cytoskeleton analysis on MCF7 and A549 morphology	88
4.4 Flow cytometry analysis of PB extracts treatment on MCF7 and	
A549	92
4.4.1 Effects of PB extracts treatment at the IC ₅₀ concentration	
on apoptotic activity of cells	92
4.4.2 Effects of PB extracts treatment at the IC ₅₀ concentration on	
cell cycle arrest on MCF7 and A549 cells	96
4.5 Gene expression analysis using Quantitative Polymerase chain	
reaction analysis of PB extracts on MCF7 and A549	99
4.5.1 Effects of PB extracts treatment at the IC_{50} concentration on	
gene expression related to apoptosis induction MCF7 and	
A549 cells.	99
4.5.2 Effects of PB extracts treatment at the IC_{50} concentration on	
gene expression related to cell cycle arrest on MCF7 and	104
A549 cells	
4.6 Chapter summary	108
CHAPTER FIVE: DISCUSSION	100
5.1 General discussion	
5.2 Limitation of the study	
5.3 Chapter summary	
5.5 Chapter summary	123
CHAPTER SIX: CONCLUSION AND FUTURE STUDY	124
6.1 Conclusion.	
6.2 Future study	
REFERENCE	128
APPENDIX A	138
APPENDIX B	140
APPENDIX C	
APPENDIX D	
APPENDIX E	
APPENDIX F	145
APPENDIX G	147

APPENDIX H	
APPENDIX I	
APPENDIX J	
APPENDIX K	
APPENDIX L	
APPENDIX M	
APPENDIX N	
APPENDIX O	

LIST OF TABLES

Table 2.1	Summary of Breast Cancer And Lung Cancer Statistic	14
Table 2.2	Summary of Different Actions Targeted By Anti-Cancer Agents.	18
Table 2.3	Cyclins And CDK Found In Homo Sapiens.	22
Table 2.4	List of Prototype Drugs With Mechanism of Action on Cell Cycle Interference.	27
Table 2.5	Summary of Difference Morphology Between Apoptosis, Oncosis, Paraptosis And Necrosis.	34
Table 3.1	List of Cell Lines.	44
Table 3.2	Reactivity Grade For Cytotoxicity Study.	55
Table 3.3	Components For Reverse Transcription Reaction.	61
Table 3.4	Reaction Components For PCR Reaction In Each Well.	62
Table 3.5	Protocol For Standard Cycling.	63
Table 4.1	Proportion of Apoptotic Cells Distribution of (A) MCF7 And (B) A549 Cells By Annexin V/7-AAD Staining.	95
Table 4.2	Proportion of Cell Cycle Distribution of (A) MCF7 And (B) A549 Cells.	98

LIST OF FIGURES

Figure 2.1	Role of Genes And Environment In Cancer Development.	
Figure 2.2	Carcinogenesis of Normal Cells Into Malignant Tumor.	
Figure 2.3	Summary of Carcinogenesis Stage And Chemoprevention Target of Phytochemicals Action.	
Figure 2.4	Example of Herbs With Chemopreventive And Anticancer Efficacy.	
Figure 2.5	Phase of Cell Cycle With Its Regulatory Cyclin/Cdks Complex.	
Figure 2.6	Schematic of P21 Mechanism of Action. P21 Approach In Arresting G1 Phase (Cyclin/CDK Deactivation) And G2 Phase (DNA Replication Inhibition) Due To P53 Induction.	26
Figure 2.7	Morphology Schematic of Cells That Undergo Apoptosis.	30
Figure 2.8	Figure 2.8 Pathways of Cell Death Leading To Necrosis.	
Figure 2.9	Overview of Intrinsic And Extrinsic Pathway of Apoptosis.	38
Figure 2.10	Pendant of Porcupine Bezoar.	43
Figure 4.1	 Cell Growth Inhibition Effects In The MCF7 (A) And A549 (B) Cells Treated With PB Extracts. 	
Figure 4.2	Cell Proliferation Inhibition Effects In The MCF7 (A) And A549 (B).	69
Figure 4.3	Cytotoxicity Effects of PB Extracts On Normal Mouse Cells 3T3 Fibroblast.	71
Figure 4.4	Analysis of PB Extracts On Normal Mouse Cells 3T3 Fibroblast Morphology.	72
Figure 4.5	Cytotoxicity Effects of PB Extracts On Normal Human Gingival Fibroblast Cells (HGF-1).	74
Figure 4.6 Figure 4.7	Analysis of PB Extracts On Normal Human Gingival Fibroblast (HG-1) Morphology. Live/Dead Analysis of MCF7 Cells Detected Under A Fluorescence Microscope.	75 78

Figure 4.8	Live/Dead Analysis of A549 Cells Detected Under A Fluorescence Microscope.	79
Figure 4.9	Analysis of PB Extract's Effect On MCF7 Demonstrated Using Inverted Microscope.	81
Figure 4.10	Analysis of MCF7 Cell Morphology Upon Treated With PB Extracts.	83
Figure 4.11	Analysis of PB Extracts Effect On A549 Demonstrated Using Inverted Microscope.	85
Figure 4.12	Analysis of A549 Cell Morphology Upon Treated With PB Extracts.	87
Figure 4.13	Cytoskeleton Analysis of MCF7 Cells. Cells Were Stained With Hoechst 33342 /Actin Phalloidin And Observed Under A Fluorescence Microscope.	89
Figure 4.14	Cytoskeleton Analysis of A549 Cells. Cells Were Stained With Hoechst 33342 /Actin Phalloidin And Observed Under A Fluorescence Microscope.	91
Figure 4.15	Effect of PB Extracts Upon Apoptosis Induction On (A) MCF7 And (B) A549.	93
Figure 4.16	Proportion of Cell Cycle Distribution of MCF7 (A) And A549 (B).	
Figure 4.17	Effect of PB Extracts On The Apoptotic Targeted Genes On MCF7 Using qPCR Analysis.	101
Figure 4.18	Effect Of PB Extracts On The Apoptotic Targeted Genes On A549 Using qPCR Analysis.	103
Figure 4.19	9 Effect of PB Extracts On The Cell Cycle Targeted Genes On MCF7 Using qPCR Analysis.	
Figure 4.20	Effect Of PB Extracts On The Cell Cycle Targeted Genes On A549 Using qPCR Analysis.	107
Figure 6.1	The Summary Finding of PB Extract Effect On MCF7 And A549 Cells.	126

LIST OF ABBREVIATIONS

ATCC	American type culture collection
A549	Lung cancer cells
BLAST	Basic local alignment search tool
Bax	Bcl-2-associated X protein
Bcl2	B-cell lymphoma 2
BID	BH3-interacting-domain death
CASPASE	Cysteine aspartic acid protease
CDKs	Cyclin-dependent kinases
CDKI	Cyclin dependent kinases inhibitors
CDNA	Complementary DNA
CGM	Complete growth media
CIP/KIP	CDK Interacting protein/kinase inhibitor protein
CO2	Carbon dioxide
ddH2O	double-distilled water
DNA	Deoxyribonucleic acid
DMEM	Dulbecco"s Modified Eagle Medium
DMSO	Dimetyl sulphoxide
Е	Efficiency
FADD	Fas-associated death domain protein
FasL	Fas ligand
FBS	Fetal bovine serum
IC50	Inhibition concentration (reduces the effect by 50%)
IAP	Inhibitor of apoptosis
GAPDH	Glyceraldehyde-3-phosphate dehydrogenase
GC	Guanine-cytosine (DNA base pairing)
KIP	Kinase inhibitory proteins
LC50	Lethal concentration
mRNA	messenger RNAs
MIQE	Minimum information for quantitative polymerase chain
	reaction publication experiments
MCF7	Breast cancer cells
NCBI	National center for biotechnology
PB	Porcupine bezoar
PBS	Phosphate buffer saline
PCD	Programmed cell death
PCR	Polymerase chain reaction
PI	Propidium Iodide
qPCR	Quantitative polymerase chain reaction
RNA	Ribonucleic acid
RT	Real-Time
7AAD	7-Aminoactinomycin D

LIST OF SYMBOLS

α	Alpha
β	Beta
Δ	Delta
Cq	Quantification cycle
Ct	Threshold cycle
g	Gram
G	Gravity
G	Gap
hpf	Hours post fertilization
Μ	Mitosis
S	Synthesis
μl	Microliter
µg/ml	Microgram per millilitre
°C	Degree Celsius
%	Percent
-	То
>	More than
<	Less than
±	Plus-minus
X	Times
=	Equal to
*	Statistical significance denotation

CHAPTER 1

INTRODUCTION

1.1 BACKGROUND OF THE STUDY

Cancer is one of major concern nowadays as it is continuously reported to be the second leading cause of death worldwide (Hashim et al. 2016; Park et al. 2008; Siegel, Miller, and Jemal 2016; Xu et al. 2016). Study reported cancer is expected to grow worldwide, especially in underdeveloped countries which contribute to 82% of the world's population (Alberg, Brock, and Samet 2005; American Cancer Society 2014; Park et al. 2008; Siegel et al. 2016; Torre et al. 2015; Yip, Taib, and Mohamed 2006). Additionally, a study by Torre et.al reported in 2012 there were 14.1 million new cancer cases and 8.2 million deaths due to cancer condition globally. In Asia alone, the incidence from 6.1 million in 2008 to 10.6 million in 2030 of cancer cases was estimated to expand as Asia contributed 60% of the world total population (Sankaranarayanan, Ramadas, and Qiao 2014).

In Malaysia, the cumulative cancer risks in peninsular Malaysia were up to 18% of 26,089 cancer patients, in which 54.7% were females and 45.3% were males (Onn and Stats 2002). The statistic also showed that the risk of getting cancer is very high with a probability of 1 from 4 Malaysian will have the potential of getting cancer in their lifetime. Furthermore, the report revealed that the highest occurrence cancer for males was lung cancer by 13.9%, while for the woman was breast cancer 30.4%. Different incidence was documented in 2016 where the leading cancer for males was

prostate (21%) followed by lung or bronchus cancer by 14.0% and for woman it is breast cancer by 29.0% subsequently lung or bronchus with 13.0% (Siegel et al. 2016).

In addition, the occurrence of cancer is becoming an enormous burden when available cancer therapies, such as surgery, chemotherapy and radiotherapy, are showing defective prognosis and various side effects (American Cancer Society 2014; Mariotto et al. 2007; De Moor et al. 2013; Siegel et al. 2013). Therefore, the exploration of anti-cancer agent with minimal toxicity and highly specific becomes urgent to avert increasing cancer cases throughout the world every year.

1.2 STATEMENT OF THE PROBLEM

Researchers had been struggling after years in finding new potential anticancer agents to manage cancer patients as current treatment leaves side effects for a lifetime and are not targeted specifically on the cancer cells (Amin et al. 2009). Thus, exploration on alternative strategy using a natural resources is crucial to overcome cancer treatment challenges.

One of long forgotten natural resources that once known as the prince of antidote are porcupine bezoar (PB). Bezoar is a stone which consists of lump undigested organic and inorganic material, which hardened into calcareous concretions in the gastrointestinal tract (Barroso 2014; Duffin 2013; Mori and Sforzi 2013). It is reported that bezoar can be existed in any kind of mammals, however the most famous bezoar is PB which is due to its medicinal value. Porcupine bezoar was documented of its medicinal value as early 8th century from Persian (Barroso 2014). Between the years of 968 and 977AD, Abu Mansur Muwaffak in his famous work Materia Medica mentioning about bezoars, its medicinal values and it was categorized under precious stone (Duffin 2013). It was described to have the ability to treat pestilent disease such as cholera, plague, malignant diseases, quartan fevers, acute febrile illness, small pox, measles, jaundice, bilious, kidney stones, pleurisy, palpitations of the heart, heart diseases, epilepsy, intestinal worms, the bloody flux, internal abscesses, leprosy, intestinal obstructions, and chickenpox (Duffin, 2013& Barroso, 2014).

Despite all these medicinal values, no scientific investigation has been done with regard to its medicinal value. Additionally, there are PB consumers in Malaysia which believe PB can cure cancer. Therefore, this study intended to discover the PB extracts effects on cancer using in vitro model to understand its effect and mechanism.

1.3 RESEARCH OBJECTIVES

Considering the inadequate information on the PB and its efficacy as anticancer agents, the general aim of this study is to explore the anticancer property of PB using in vitro models. The specific objectives are:

- i. To determine 50% inhibitory concentration (IC_{50}) of PB on human breast cancer cells (MCF7) and human lung cancer cells (A549).
- ii. To determine the effect of PB extracts on normal cell lines, mouse embryo fibroblast (3T3-L1) and normal human gingival fibroblast (HGF-1).
- iii. To observe apoptosis related morphological changes upon treatment with PB extracts on MCF7 and A549 cells.
- iv. To investigate the effect of PB extracts on apoptosis and its markers (*Bax, Bcl-*2, *Bid, caspases, Cytohrome C*) on MCF7 and A549 cells.
- v. To investigate the effect of PB extracts on cell cycle regulation and its markers (cyclins, cyclin dependent kinases) on MCF7 and A549 cells.

1.4 RESEARCH QUESTIONS

- i. Does *IC*₅₀ of PB extracts induce significant effects on MCF7 and A549 cells?
- ii. Do PB extracts induce significant toxicity to 3T3-L1 and HGF-1?
- iii. Does PB extracts able to induce apoptosis morphology on MCF7 and A549 cells?
- iv. Does IC_{50} of PB extracts induce significant effects of apoptosis on MCF7 and A549 cells?
- Does *IC*₅₀ of PB extracts induce significant effects of arresting cell cycle on MCF7 and A549 cells?

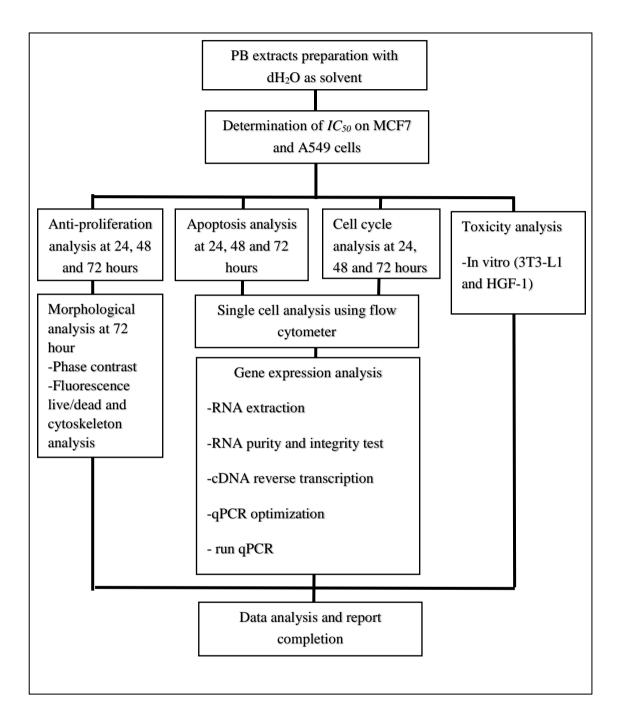
1.5 RESEARCH HYPOTHESES

- i. PB extracts induce significant 50% inhibitory at low concentration.
- ii. PB extracts can be toxic to 3T3-L1 and HGF-1 only at extremely high concentration.
- iii. PB extracts able to induce morphological changes of apoptosis on MCF7 and A549 cells.
- iv. PB extracts trigger apoptosis on MCF7 and A549 cells.
- v. PB extracts promote cell cycle arrest on MCF7 and A549 cells.

1.6 SIGNIFICANCE OF THE STUDY

For the first time, the present study will provide scientific data on the effect of PB extract on cancer cells, MCF7 and A549. The cell cycle profile changes and type of cell death induced by PB extract also crucial as a part of anticancer candidate development. In addition, the molecular part of the result will provide more information in understanding of how PB extracts react towards MCF7 and A549 cells at the molecular level. Furthermore, a study done in normal cells and preliminary findings in zebrafish will provide better understanding on the therapeutic and toxic dosage of PB extracts. Overall, this study will provide a better insight of PB and the strategy that can be established to direct the study further as potential anticancer agents in the future.

1.7 RESEARCH FRAMEWORK



1.8 CHAPTER SUMMARY

This chapter has presented and discussed the background of the study consisting of the global, Asia and Malaysia statistic on cancer cases that occurred previously. Explained in the research background and problem statement section the importance of finding anticancer agents which give justification to conduct the study. Additionally, the statement of the problem was discussed, as this study was undertaken to investigate PB effects on breast and lung cancer cells. The research objectives, questions and hypotheses were also outlined in this chapter, followed by the significance of study and finally the research framework used in this study was also presented.