MicroRNA (miRNA) PROFILE IN ACUTE MYOCARDIAL INFARCTION (AMI) OF YOUNG ADULTS IN KUANTAN, PAHANG

BY

NURUL ASHIKIN MUHAMMAD MUSA

A thesis submitted in fulfillment of the requirement for the degree of Doctor of Philosophy in Medical Sciences (Biochemistry)

Kulliyyah of Medicine International Islamic University Malaysia

SEPTEMBER 2022

ABSTRACT

Acute myocardial infarction (AMI) is a severe form of coronary heart disease where Malaysians are getting AMI at younger age compared to well-developed countries. MicroRNAs (miRNAs) are implicated in AMI pathogenesis, but no study looked at their profiling or involvement in young population. The present study aims to profile the miRNAs expressions in healthy controls (aged 18 to 45 years), young AMI (YAMI) (aged \leq 45 years), and mature AMI (MAMI) (aged \geq 46 years) patients with matching criteria and to determine the effect of the dysregulated miRNAs on the target mRNAs as well as the pathways involve in the pathogenesis of AMI. This study was conducted on twenty Malay males for each group in Kuantan, Pahang. Total RNA was extracted from plasma and the miRNA expression profiling was carried out on the BGISEQ500 SE50 sequencing platform with BGI sequencing libraries. The sequence data were analyzed using Gene Ontology (GO) to determine the role of the differentially expressed genes, followed by the Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analysis for identification of the biological pathways in YAMI against MAMI. The top six dysregulated miRNAs identified during sequencing were validated using quantitative reverse-transcription polymerase chain reaction (qRT-PCR) between the groups. ANOVA and unpaired T-test were used to analyze the differences of miRNAs and gene expression between the three groups. This study revealed that majority AMI patients were smokers, where YAMI patients had higher BMI, SBP, DBP and TG while MAMI patients had higher FBG than the rest of the group. A total of 1599 miRNAs were differentially expressed in AMI (YAMI and MAMI) patients compared to healthy controls, where 1288 were upregulated and 311 were downregulated (FDR ≤ 0.001). However, when YAMI patients were compared to MAMI patients, 1497 miRNAs were found to be dysregulated, of which 1090 miRNAs were upregulated, and 407 miRNAs were downregulated (FDR \leq 0.001). The top ten upregulated miRNAs were miR-552, miR-4446-3p, miR-432-5p, miR-548j-5p, miR-219, miR-982, miR-181a-2-3p, miR-654-5p, miR-58 and miR-548k; while the top ten downregulated were miR-16-5p, miR-1064, miR-431-5p, miR-790 miR-1177, miR-201, miR-105, miR-518, miR-419 and miR-1103. This study also discovered ten novel miRNAs: miR-4446-3p, miR-982, miR-58, miR-548k, miR-1064, miR-790, miR-1177, miR-201, miR-419, and miR-1103. The validation of the top six dysregulated miRNAs between YAMI and MAMI patients revealed the upregulation of miR-423-5p by 2.08fold (p = 0.040) and downregulation of miR-431-5p by 33.90-fold (p = 0.034), and miR-378a-5p by 34.61-fold (p = 0.040). For these 1497 differentially expressed miRNAs, 34,195 target genes were predicted by GO analysis. The functional analysis demonstrated 11,199 GO terms found to be involved in biological processes, 12,012 in cellular components, and 10,984 in molecular functions were significantly enriched (p < 0.05). The target genes that were mapped to the signal transduction pathway in KEGG revealed 346 classes were enriched. In conclusion, miRNAs are differentially expressed between young and mature AMI, ten of which are novel. Three biological pathways, and aldarate metabolism, collecting duct acid secretion glycosaminoglycans biosynthesis – heparin sulfate/heparin were identified but their involvements in the regulatory mechanisms on gene expression in Young AMI need further evaluation.

ملخص البحث

احتشاء العضلة القلبية الحاد (Acute myocardial infarction, AMI) هو مرض قلبي حاد من أمراض القلب التاجية، يصاب به الماليزيون في سن أصغر مقارنة بالدول المتقدمة. الحمض النووي الريبوزي الدقيق (miRNAs) يساهم بشكل رئيسي في التسبب بمذا المرض، ولا توجد دراسة سابقة قامت بتحديد دوره أو دراسة سماته في فئة الشباب. استهدفت الدراسة الحالية تحديد سمات الدmiRNAs وتعبيراته الجينية في المجموعات التالية: الأصحاء كمجموعة ضابطة (18-45 عاماً)، ومجموعتين من مرضى الـAMI وهي، مجموعة المرضى الشباب YAMI (> 45 عاماً) ومجموعة المرضى الكبار MAMI (< 46 عاماً)، المرضى ذو المعايير المطابقة تم دراستهم لتحديد تأثير خلل التنظيم في الmiRNAs على الحمض المستهدف من الmRNA و كذلك المسارات المشاركة في التسبب بالمرض. أجريت هذه الدراسة على 20 رجلاً في كل مجموعة، في مدينة كوانتن بإقليم بمانج. أستخلص حمض الرنا من البلازما وحُددت سمات التعبير الجيني لـ miRNAs على منصة التسلسل BGISEQ500 SE50 وعبر مكتبة بيانات التسلسل. أُجري تحليل بيانات التسلسل باستخدام الأونتولوجيا الجينية (Gene Ontology, GO) لتحديد دور الجينات، متبوعًا بتحليل موسوعة كيوتو للجينات والجينوم الإثرائي لتحديد المسارات البيولوجية في مجموعة الـYAMI مقابل مجموعة الـMAMI . تم التحقق من صحة أفضل ستة miRNAs ذات خلل في التعبير الجيني وتحديدها أثناء النسخ باستعمال تفاعل البوليمراز المتسلسل الكمي للنسخ العكسي بين المجموعات. واستخدم إحصاء تحليل التباين واختبار "تي" غير المقيد لتحليل الاختلافات في miRNAs والتعبير الجيني بين المجموعات الثلاث. كشفت هذه الدراسة أن غالبية مرضى AMI كانوا مدخنين، حيث كان مؤشر كتلة الجسم أعلى لدى YAMI ، وضغط الدم الانقباضي والانبساطي عالى، وارتفاع في مستوى دهن ثلاثي الجليسريد. بينما الMAMI كان لديهم معدل سكر الصيام أعلى في الدم مقارنة ببقية المجموعات. تم التعبير جينياً عن إجمالي 1599 حمضاً من الmiRNAs بشكل تفاضلي في مرضى الك AMI) AMI و MAMI) مقارنةً بالمجموعة الضابطة، حيث كان التعبير بالتنظيم الرفعي 1288 وكان التعبير بالتنظيم التخفيضي 311 (معدل الاكتشاف الخاطئ <0.001). ومع ذلك، عند مقارنة المرضى الـYAMI بالـMAMI، تم العثور على 1497 حمض من miRNAs غير منتظم التعبير، منها كان التعبير بالتنظيم الرفعي 1090 حمضاً، و 407 حمضاً كان التعبير بالتنظيم التخفيضي (معدل الاكتشاف الخاطئ < 0.001).ال miRNAs المنظمة رفعيا وفي المراتب العشر الأولى كانت: -miR-552, miR-4446-3p, miR-432-5p, miR-548j-5p, miR-219, miR-982, miR-181a-2-3p, miR-654-5p, miR-58 and miR-548k بينما العشرة الأحماض الأخرى من miR-16-5p, miR-1064, miR-431-5p, miR-790, المعبرة بالتنظيم التخفيضي هي: miR-16-5p, miR-1064, miR-431-5p, miR-790, miR-1177, miR-miR-201, miR-105, miR-518, miR-419 and miR-1103. اكتشفت هذه الدراسة أيضاً عشرة أحماض من miRNAs وهي: -miR-4446-3p, miR-982, miR-58, miR-548k, miR وهي: -miR-4446-3p - 11031064, miR-790, miR-miR-1177, miR-201, miR-419, and miR-فعالية عالية من الـmiRNAs ذات خلل تعبيري بين المرضى YAMI و MAMI كانت بالتنظيم الرفعي ل 92-423 miR-423 بمقدار 2.08 طي (miR-431-5p)، أما بالتنظيم التخفيضي فكانت ل miR-431-5p بمقدار 23.9 طي (0.034 =p)، و-miR-378a-5p بقدار 34.61 طي (9- 0.04). بالنسبة إلى هذه ال1497 حمضاً من miRNAs المعبر عنها تفاضلياً، تم توقع استهداف 34195 جيناً بواسطة تحليل اله. GO. أظهر التحليل الوظيفي أن 11199 مصطلحاً من مصطلحات GO وجدت أنها مشاركة في العمليات البيولوجية، و 12012 في المكونات الخلوية، و10984 في الوظائف الجزيئية كانت غنية بشكل أكبر (0.05 > p). كشفت الجينات المستهدفة التي تم تحديدها لمسار تحويل الإشارة في موسوعة كيوتو للجينات والجينوم أنه تم إثراء 346 فئة. اختصاراً، التعبير الجيني عن miRNAs عبرت بشكل تفاضلي بين مرضى الYAMI والMAMI، عشرة منها جديدة. تم تحديد ثلاثة مسارات بيولوجية وهي: أيض الأسكوربات والألدارات، وإفراز حمض القناة، والتخليق الحيوي للجليكوز أمينو جليكان - كبريتات الهيبارين/الهيبارين، ولكن مشاركتها في الآليات التنظيمية للتعبير الجيني في مجموعة المرضى الشباب تحتاج إلى مزيد من الدراسة. تم تحديد ثلاثة مسارات بيولوجية، أيض الأسكوربات والألدارات، وإفراز حمض القناة، والتخليق الحيوي للجليكوز أمينو جليكان - كبريتات الهيبارين/الهيبارين، لكن مشاركتها في الآليات التنظيمية للتعبير الجيني في مجموعة المرضى الشباب تحتاج إلى مزيد من الدراسة.

APPROVAL PAGE

The thesis of Nurul Ashikin Muhammad Musa has been approved by the following:

Nor Zamzila Abdullah Supervisor
Norlelawati A. Talib
Co-supervisor
Aszrin Abdullah
Co-supervisor
Azarisman Shah Mohd Shah
Co-supervisor
Sirajudeen Kuttulebbai Naina Mohamed Salam
Internal Examiner
Wan Rohani Wan Taib
External Examiner
-
Muhammad Lokman Md. Isa Chairman
Chamban

DECLARATION

I hereby declare that this thesis is the result of my own investigations, except where otherwise stated. I also declare that it has not been previously or concurrently submitted as a whole for any other degrees at IIUM or other institutions.

Nurul Ashikin Muhammad Musa

G:	D /
Signature	Date

INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

DECLARATION OF COPYRIGHT AND AFFIRMATION OF FAIR USE OF UNPUBLISHED RESEARCH

MicroRNA (miRNA) PROFILE IN ACUTE MYOCARDIAL INFARCTION (AMI) OF YOUNG ADULTS IN KUANTAN, PAHANG

I declare that the copyright holder of this thesis is Nurul Ashikin Muhammad Musa.

Copyright © 2022 International Islamic University Malaysia. All rights reserved.

No part of this unpublished research may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder except as provided below

- 1. Any material contained in or derived from this unpublished research may only be used by others in their writing with due acknowledgement.
- 2. IIUM or its library will have the right to make and transmit copies (print or electronic) for institutional and academic purpose.
- 3. The IIUM library will have the right to make, store in a retrieval system and supply copies of this unpublished research if requested by other universities and research libraries.

By signing this form, I acknowledged that I have read and understand the IIUM Intellectual Property Right and Commercialization policy.

Affirmed by Nurul Ashikin Muhammad Musa		
Signature	Date	



ACKNOWLEDGEMENTS

All glory is due to Allah, the Almighty, whose Grace and Mercies have been with me throughout the duration of my programme. Although, it has been tasking, His Mercies and Blessings on me ease the herculean task of completing this thesis.

I am most indebted to by supervisor, Assoc. Prof. Dr. Nor Zamzila Abdullah and my co-supervisor, Assoc. Prof. Dr. Norlelawati A. Talib, whose enduring disposition, kindness, promptitude, thoroughness and friendship have facilitated the successful completion of my work. I put on record and appreciate their detailed comments, useful suggestions and inspiring queries which have considerably improved this thesis. Their brilliant grasp of the aim and content of this work led to their insightful comments, suggestions and queries which helped me a great deal. Despite their commitments, they took time to listen and attend to me whenever requested. The moral support they extended to me is in no doubt a boost that helped in building and writing thedraft of this research work. I am also grateful to my co-supervisor, Asst. Prof. Dr. Aszrin Abdullah and Prof. Dr. Azarisman Shah Mohd Shah, whose support, and cooperation contributed to the outcome of this work.

Apart from my supervisor and co-supervisors, I am so thankful to Asst. Prof. Dr. Aida Nur Sharini Mohd Shah, Emergency Physician of Sultan Ahmad Shah Medical Centre, IIUM and Dr. Badli Sham Baharun, the Head of Emergency Department, Hospital Tengku Ampuan Afzan and all their staffs for their support and help in recruitment of patients for this research. I am also thankful to all the staffs at IIUM Kuantan Campus and Klinik Kesihatan Bandar Kuantan for their help in recruitment of control subjects. My gratefulness also goes to Asst. Prof. Dr. Norbaiyah Mohamed Bakrim and Asst. Prof. Dr. Wan Fatein Nabeila Wan Omar for their help in this research project. I am also highly indebted to Asst. Prof. Dr. Wan Muhamad Salahudin Wan Salleh, Asst. Prof. Dr. Nour El Huda Abd Rahim and Assoc. Prof. Dr. Roslina Abdul Rahim for their help and support during this study period.

Lastly, my gratitude goes to my beloved husband, Syahwarni Noor Shah and my dear mother, Salamah Radin Sumadi, for their prayers, understanding and endurance while I was away.

Once again, we glorify Allah for His endless mercy on us, one of which is enabling us to successfully round off the efforts of writing this thesis. Alhamdulillah.

TABLE OF CONTENTS

Abstract		ii
	ı Arabic	
Approval F	age	iv
	1	
Copyright.		vi
100	lgements	
	les	
	ıres	
	previations	
CHAPTE	R ONE: INTRODUCTION	
1.1	Background and Justification	
1.2	Research Question	
1.3	General Hypothesis	
1.4	Specific Hypotheses	
1.5	General Objective	4
1.6	Specific Objectives.	4
		_
	R TWO: LITERATURE REVIEW	
2.1	Acute Myocardial Infarction (AMI)	
	2.1.1 An Overview	
	2.1.2 Definition of AMI	
	2.1.3 Classification of AMI	
	2.1.4 Prevalence, Mortality and Morbidity	
	2.1.5 Aetiology and Risk Factors of AMI	
	2.1.6 Pathogenesis of AMI	
	2.1.7 AMI in Young Adults	
	2.1.8 Cut-off Age of 'Young'	
2.2	MicroRNA (miRNA)	14
	2.2.1 An Overview	
	2.2.2 Biogenesis of miRNA	16
	2.2.3 miRNA Binding and Its Function	
	2.2.4 Factors Influencing miRNA Expression and Its Half-Life	
2.3	miRNA and AMI	
	2.3.1 AMI and Apoptosis	
	2.3.2 AMI and Necrosis	24
	2.3.3 AMI and Autophagy	
2.4	Rationale of the Study	27
2.5	Conceptual Framework	28
OTT 1 555		
	R THREE: MATERIALS AND METHODS	
3.1	Material	
	3.1.1 Equipment	
	3.1.2 Reagents and Disposable Materials	
3.2	Study Design	
3 3	Type of Sampling	31

3.4	Study Period	32	
3.5	Study Population		
3.6	Sample Size	32	
3.7	Selection of Subjects		
	3.7.1 Control Subjects	33	
	3.7.1.1 Inclusion Criteria		
	3.7.1.2 Exclusion Criteria		
	3.7.2 AMI Patients		
	3.7.2.1 Inclusion Criteria for Young AMI		
	3.7.2.2 Exclusion Criteria for Young AMI		
	3.7.2.3 Inclusion Criteria for Mature AMI		
	3.7.2.4 Exclusion Criteria for Mature AMI		
3.8	Sample and Data Collection		
5.0	3.8.1 Data From Questionnaire		
	3.8.2 Blood Sample Collection		
3.9	Phases of the Study		
	Determination of Lipid Parameters		
5.10	3.10.1 Total Cholesterol (TC)		
	3.10.1.1 Principle		
	3.10.1.2 Procedure		
	3.10.2 Triglyceride (TG)		
	3.10.2.1 Principle		
	3.10.2.2 Procedure		
	3.10.3 High Density Lipoprotein Cholesterol (HDL-C)		
	3.10.3.1 Principle		
	3.10.3.2 Procedure		
2 1 1	3.10.4 Low Density Lipoprotein Cholesterol (LDL-C)		
3.11	Determination of Fasting Blood Glucose		
	3.11.1 Principle		
2 10	3.11.2 Procedure		
3.12	Extraction of Total RNA From Plasma		
	3.12.1 Principle		
2 12	3.12.2 Procedure		
3.13	miRNA Analysis		
	3.13.1.1 Small RNA Sequencing		
	3.13.1.2 Principle		
	3.13.1.3 Procedure		
	3.13.1.3.1 Experiment Pipeline		
	3.13.1.3.2 Bioinformatics Pipeline		
	3.13.1.3.3 Data Filtering		
	3.13.1.3.4 Reads Mapping		
	3.13.1.3.5 sRNA Classification		
	3.13.1.3.6 sRNA Prediction		
	3.13.1.3.7 sRNA Expression		
	3.13.1.3.8 Target Gene Prediction	53	
	3.13.1.3.9 Screening Differentially Expressed Scores		
	(DESs) With Differentially Expressed Gene		
	Sequence (DEGseq)		
	3.13.1.3.10 Hierarchical Clustering Analysis		
	3.13.1.3.11 Gene Ontology (GO) Enrichment Analysis	54	

	3.13.1.3.12 Pathway Enrichment Analysis	55
	3.13.2 Validation of Selected Dysregulated miRNAs	56
	3.13.2.1 Selected miRNAs	
	3.13.2.2 qRT-PCR	56
	3.13.2.2.1 Principle	56
	3.13.2.2.2 Procedure	57
	3.13.2.2.3 miRNAs Expression Analysis	60
	3.13.3 mRNA Analysis	
	3.13.3.1 Genes Selected for mRNA Expression	61
	3.13.3.1.1 Principle	
	3.13.3.1.2 Procedure	
3.14	Statistical Analysis	66
CHAPTEI	R FOUR: RESULTS	
4.1	Study Population	67
	4.1.1 Demographic and Clinical Characteristics of Participants	
	Selected for sRNA-seq	67
	4.1.2 Demographic and Clinical Characteristics of Participants	
	Selected for qRT-PCR	
4.2	1	72
	4.2.1 miRNA Expression Profile of Healthy Controls Versus AMI	
	(Young AMI and Mature AMI) Patients	
	4.2.1.1 Hierarchical Clustering	
	4.2.1.2 Distribution of Expressed miRNAs	
	4.2.1.3 Twenty Most Differentially Expressed miRNAs	75
	4.2.2 miRNA Expression Profile of Healthy Controls Versus	
	Young AMI Patients	
	4.2.2.1 Hierarchical Clustering	
	4.2.2.2 Twenty Most Differentially Expressed miRNAs	78
	4.2.3 miRNA Expression Profile of Young AMI Versus Mature	
	AMI Patients	
	4.2.3.1 Hierarchical Clustering	
	4.2.3.2 Distribution of Expressed miRNAs	
	4.2.3.3 Twenty Most Differentially Expressed miRNAs	81
4.3	Pathways Involve in Pathogenesis of AMI in Young AMI Group	0.2
	Based on Dysregulated miRNAs	82
	4.3.1 Gene Ontology (GO) Analysis of Differentially Expressed	00
	miRNAs in Young AMI Group	82
	4.3.2 Kyoto Encyclopedia of Genes and Genomes (KEGG)	0.4
4.4	Pathway Analysis	84
4.4	Validation of Dysregulated miRNAs Expression in Healthy Controls,	0.5
	Young AMI and Mature AMI Patients	85
	4.4.1 miRNAs Expression Between Heathy Controls and AMI	0.5
	(Young AMI and Mature AMI) Patients	03
	4.4.2 miRNAs Expression Between Heathy Controls and Young AMI Patients	97
		0/
	4.4.3 miRNAs Expression Between Young AMI and Mature AMI	QO
15	Patients	00
4.3	Mature AMI Patients	89
	TRECORDER OF THE PROPERTY OF T	177

4.6	Summary of the Results	91
CHAPTE	R FIVE: DISCUSSION	93
5.1	Overview of the Study	93
5.2	General Characteristics of the Study Population	93
5.3	miRNA Expression Profile in Healthy Controls and AMI Patients	94
	5.3.1 Proposed Role of Dysregulated miRNAs in Pathogenesis of	
	AMI	
5.4	Validation of miRNAs Expression That Were Significantly	
	Dysregulated in Healthy Controls, Young AMI, and Mature AMI	
	Patients	
5.5	Pathways Involve in Pathogenesis of AMI in Young AMI Group	
	Based on Dysregulated miRNAs	103
5.6	mRNA Expression of the Dysregulated miRNAs in Young AMI and	
	Mature AMI Patients	105
5.7		
5.8	Impact of the Study	
CHAPTE	R SIX: CONCLUSION	108
REFERE	NCES	109
APPENDI	X I: INFORMATION SHEET AND CONSENT FORM	135
APPENDI	X II: CASE RECORD FORM AND QUESTIONNAIRE	148
	X III: RAW DATA ON RNA CONCENTRATION OF ALL	
	STUDY SAMPLES	
APPENDI	X IV: LIST OF PUBLICATIONS AND AWARDS	

LIST OF TABLES

Table 2.1	Classification of AMI.	7
Table 2.2	Various miRNAs and Their Role in Pathophysiological Pathways in AMI	26
Table 3.1	The Selected miRNAs and Their Target Sequences.	56
Table 3.2	Reverse Transcription Reaction Setup Per Sample.	58
Table 3.3	Reverse Transcription Reaction Temperature Cycling Protocol.	58
Table 3.4	cDNA Dilution for miRCURY LNA miRNA Custom PCR Panels.	59
Table 3.5	Reaction Setup Per Sample for miRCURY LNA miRNA Custom PCR Panels.	59
Table 3.6	PCR Cycling Condition for miRCURY LNA miRNA Custom PCR Panels.	60
Table 3.7	Target Genes and Reference Genes for mRNA Expression.	61
Table 3.8	Genomic DNA Removal Reaction Components.	62
Table 3.9	Reverse Transcription Reaction Components.	62
Table 3.10	gDNA Elimination and RT Temperature Protocol.	64
Table 3.11	Reaction Setup.	64
Table 3.12	Real-Time Cycler Conditions.	65
Table 4.1	Demographic and Baseline Clinical Characteristics of Participants in sRNA-seq.	68
Table 4.2	Demographic and Baseline Clinical Characteristics of Participants in qRT-PCR.	71
Table 4.3	Differentially Expressed miRNAs Between Healthy Controls and AMI (Young and Mature AMI) Patients in Small-RNA Sequencing.	75
Table 4.4	Differentially Expressed miRNAs Between Healthy Controls and Young AMI Patients in Small-RNA Sequencing.	78
Table 4.5	Differentially Expressed miRNAs Between Young AMI and Mature AMI Patients in Small-RNA Sequencing.	81

Table 4.6	Validation of miRNAs That Were Dysregulated Between Healthy Controls and AMI (Young AMI and Mature AMI) Patients	86
Table 4.7	Validation of miRNAs That Were Dysregulated Between Healthy Controls and Young AMI Patients	88
Table 4.8	Validation of miRNAs That Were Dysregulated Between Young AMI and Mature AMI Patients	89
Table 4.9	Summary of Known and Novel Dysregulated miRNAs in This	92



LIST OF FIGURES

Figure 2.1	Pathophysiology of AMI.	10
Figure 2.2	Biogenesis of miRNA.	16
Figure 2.3	miRNAs Implicated in Plaque Destabilization.	21
Figure 2.4	Conceptual Framework.	29
Figure 3.1	The Study Flow.	37
Figure 3.2	The Phases of the Study.	38
Figure 3.3	Small-RNA Experiment Process.	48
Figure 3.4	Bioinformatics Analysis Pipeline for Small RNA Sequencing.	50
Figure 4.1	Initial Screening for Differentially Expressed miRNAs in Various Groups.	72
Figure 4.2	Hierarchical Clustering of Differentially Expressed miRNAs in Healthy Controls and AMI (Young and Mature AMI) Patients.	73
Figure 4.3	Volcano Plot of Differential miRNA Expression in Controls and AMI (Young AMI and Mature AMI) Patients.	74
Figure 4.4	Hierarchical Clustering of Differentially Expressed miRNAs in Healthy Controls and Young AMI Patients.	76
Figure 4.5	Volcano Plot of Differential miRNA Expression in Controls and Young AMI Patients.	77
Figure 4.6	Hierarchical Clustering of Differentially Expressed miRNAs in Young AMI and Mature AMI Patients.	79
Figure 4.7	Volcano Plot of Differential miRNA Expression in Young AMI and Mature AMI Patients.	80
Figure 4.8	GO Analysis of Differentially Expressed miRNAs That Covers Three Domains: Biological Process, Cellular Components, and Molecular Function.	83
Figure 4.9	Scatter Plot of Enriched KEGG Pathway Analysis of Differentially Expressed miRNAs Between Young AMI and Mature AMI Patients.	84
Figure 4.10	Differentially Expressed miRNAs in Healthy Controls and AMI (Young AMI and Mature AMI) Patients.	86

Figure 4.11	Differentially Expressed miRNAs in Healthy Controls and Young AMI Patients.	87
Figure 4.12	Differentially Expressed miRNAs in Young AMI and Mature AMI Patients.	88
Figure 4.13	Amplification Curve (A) and Melt Curve (B) of the Target and Housekeeping Genes at Various Temperatures in Gradient Analysis.	90



LIST OF ABBREVIATIONS

ACS Acute coronary syndrome

ACTB Actin B

ADAM10 A disintegrin and metalloproteinase 10

ADP Adenosine diphosphate

AGO Argonaut Ago 2 Argonaut 2

AIM2 Absent in melanoma 2
AKI Acute kidney injury

AMI Acute myocardial infarction

AMPK Adenosine monophosphate-activated protein kinase

ATG4B Autophagy-related 4B
ATG7 Autophagy-related 7
ATP Adenosine triphosphate
AUC Area under the curve
BCL-2 B-cell lymphoma 2

BGI Beijing Genome Institute

BMI Body mass index

BNIP3 BCL-2/adenovirus E1B 19 k-Da protein-interacting protein 3

C5ARI Complement C5A receptor inhibitor

CABG Coronary artery bypass grafting

CAD Coronary artery disease
CD40 Cluster of differentiation 40

CD40L Cluster of differentiation 40 ligand

CDC Center for Disease Control

cDNA Complementary deoxyribonucleic acid

CHD Coronary heart disease circANXA2 Circular RNA ANXA2

cTn Cardiac troponin

CVD Cardiovascular disease
DBP Diastolic blood pressure

DEG Differentially expressed gene

DGCR8 DiGeorge syndrome critical region 8

DNA Deoxyribonucleic acid

dNTPs Deoxynucleotide triphosphate

dsRNA Double-stranded RNA

DUSP-JNK1/2 Dual-specificity protein phosphatase 1-c-jun N-terminal kinase 1

or 2

ECG Electrocardiogram
ECM Extracellular matric
ED Emergency Department

EDTA Ethylenediamine tetraacetic acid

FBG Fasting blood glucose FOLR3 Folate receptor 3

G6P-DH Glucose-6-phosphate dehydrogenase

GAG Glycosaminoglycans

GAPDH Glyceraldehyde 3-phosphate dehydrogenase

gDNA Genomic deoxyribonucleic acid

GDP Gross domestic product

GK Glycerol kinase

GMR Glutamate metabotropic receptor

GO Gene Ontology

GPO Glycerol phosphate oxidase

GRM4 Glutamate metabotropic receptor 4

GZMB Granzyme B

H/R Hypoxia/reperfusion

H₂O Water

H₂O₂ Hydrogen peroxide

H9c2 Clonal cell or cell line derived from rat heart tissue

HDL High-density lipoprotein

HIF-1 α Hypoxia inducible factor 1 alpha

HIPK3 Homeodomain-interacting protein kinase 3

HK Hexokinase

I/R Ischaemia/reperfusion

IC Intercalated cell IFN-γ Interferon-gamma

IHD Ischaemic heart disease

IIUM International Islamic University Malaysia

IL-1 β Interleukin 1 betaIL-29Interleukin 29IQRInterquartile range

KEGG Kyoto Encyclopedia of Genes and Genomes

KKM Kementerian Kesihatan Malaysia

KLF2 Kruppel-like factor 2 LBBB Left bundle branch block **LDL** Low-density lipoprotein **LRR** Leucine -rich repeat

LVH Left ventricular hypertrophy

MADB Bis dimethylaniline disodium salt

MARCH6 Membrane-associated ring-CH finger protein 6

 Mg^{2+} Magnesium ion

MI Myocardial infarction

miRNA **MicroRNA**

MLKL Mixed lineage kinase domain-like

MMPs Matrix metalloproteinases

MOH Ministry of Health

MREC Medical Research Ethical Committee

mRNA Messenger RNA

MSCs Mesenchymal stem cells

MYBL2 MYB proto-oncogene like 2

NAD Nicotinamide adenine dinucleotide

NADH Nicotinamide adenine dinucleotide hydride

National Cardiovascular Disease – Percutaneous Intervention NCVD-PCI

NEMO Nuclear factor-kappa B essential modulator

NF-κB Nuclear factor kappa B NF1B Nuclear factor 1B

NLRP3 Nucleotide-binding domain (NOD-), leucine-rich repeat (LRR),

pyrin domain-containg-3

NOD Nucleotide-binding and oligomerization domain

NOS Nitric oxide synthase

Non-ST elevation myocardial infarction **NSTEMI**

P53 or TP53 Tumour protein 53

PCI Percutaneous intervention

PCSK9 Proprotein convertase subtilisin kexin 9

piRNA Piwi-interacting ribonucleic acid

Pre-miRNA Precursor miRNA Pri-miRNA Primary miRNA

PTCH1 Protein patched homolog 1

QC Quality control

qRT-PCR Quantitative Reverse Transcription Polymerase Chain Reaction qRT-PCR Quantitative reverse-transcription polymerase chain reactions

RAB22A Ras-related protein Rab-22A **RBBB**

Right bundle branch block

RBP RNA binding protein

RIPK Receptor-interacting serine/threonine-protein kinase

RISC RNA induced silencing complex

RNA Ribonucleic acid RNAi RNA interference

SASMEC Sultan Ahmad Shah Medical Center

SBP Systolic blood pressure

SCN4A Sodium voltage-gated channel type 4 alpha

SD Standard deviation

SDF Stromal cell-derived factor

SELT Selenoprotein T shRNA Short hairpin RNA

siRNA Small interfering ribonucleic acid

SIRT3 Sirtuin 3

SMCs Smooth muscle cells

snoRNA Small nucleolar ribonucleic acid

SPSS Statistical Package for Social Sciences

sRNA-seq Small RNA sequencing

STEMI ST elevation myocardial infarction

TAK TGF- β activated kinase 1

TC Total cholesterol

TEH Total health expenditure

TG Triglyceride

TGF- β Transforming growth factor beta

TLR4 Toll-like receptor 4

TRIM55 Tripartite motif-containing protein 55

UDP-G Uridine diphosphate-glucose
UMI Unique Molecular Identifier

URL Upper Reference Limit
UTR Untranslated region

VCMCs Vascular smooth muscle cells

VDR Vitamin D receptor
VF Ventricular fibrillation

WHO World Health Organization

ZEB1 Zinc finger E-box binding homeobox 1

μL microliter

CHAPTER ONE

INTRODUCTION

1.1 BACKGROUND AND JUSTIFICATION

Acute myocardial infarction (AMI) is the lethal manifestation of coronary heart disease (CHD), also known as ischaemic heart disease (IHD), with high morbidity and mortality (Roth et al., 2017; WHO, 2021b). It occurs when there is an injury to a part of the heart muscle tissue due to lack of oxygenation, which is usually caused by a partial or total blockage of the coronary blood flow (Thygesen et al., 2018). AMI is the leading cause of death globally and predicted to remain so for the next 20 years (Roth et al., 2017). According to the World Health Organization (WHO), in 2019, 17.9 million people died from cardiovascular disease (CVD), which accounted for 32% of all global deaths, and of these deaths, 85% were due to AMI and stroke (WHO, 2021b). About three fourth of deaths occurred in low- and middle-income countries (WHO, 2021b). This makes CHD, particularly AMI, a serious major public health issue around the world.

In Malaysia, AMI is also the leading cause death. According to the Department of Statistics Malaysia, IHD accounted for 17% of 109,155 medically certified deaths in 2020 (Department of Statistics Malaysia, 2021). In addition to its mortality burden, IHD is also the leading cause of morbidity and loss of quality of life and exerts heavy economic costs with yearly increment in total health expenditure. In 2018, the Malaysian government spent RM60,339 million on total expenditure on health (TEH), which is equivalent to 4.17% of gross domestic product (GDP), rising to RM64,306 million, equivalent to 4.26% of GDP in 2019 (National Health Accounts Malaysia, 2021). With the continuous rise of prevalence and incidence of IHD, the Malaysian Government may need to bear further financial strain on the health care expenditure.

In Asia, particularly Malaysia, people are getting AMI at younger age compared to well-developed countries with the age range of 55.9 to 59.5 years in 2006 to 2010, and 55.0 to 59.3 years in 2012 to 2016 versus 63.4 to 68 years in well-developed countries (Lee et al., 2021; Lu & Nordin, 2013). According to the Malaysian National

Cardiovascular Disease-Percutaneous Coronary Intervention (NCVD-PCI) database, between 2007 to 2009, the prevalence of young AMI under the age of 45 years old is about 16% (Zuhdi et al., 2013) while between 2017 and 2018, the prevalence of AMI under the age of 50 is approximately 23% (Wan Ahmad, 2021). The consequences of AMI can be devastating particularly at "young" age due to the huge impact on patient's psychology, ability to work and socioeconomic burden. Besides, as this young patient may be the main income provider of the family, the repercussion following AMI can also affect multiple dependents.

One of the major factors for developing AMI in this young population is genetic predisposition. A family history of IHD is considered as one of the most relevant risk factors for developing early onset of AMI as positive family history was reported to be higher in young AMI than older AMI patients (Ambroziak et al., 2020; Ge et al., 2017; Lei & Bin, 2019; Venkatason et al., 2019). Therefore, a deeper molecular understanding on the pathological processes of AMI is crucial for both basic cardiovascular and clinical research. Since the study of IHD in young population is important in the era of preventive cardiology, this knowledge is also vital in developing the framework in primary and secondary prevention in the future.

MicroRNAs (miRNAs) are short, single stranded, noncoding RNAs that regulate gene expression post-transcriptionally by binding to the untranslated region (UTR) of the target messenger RNA (mRNA) (Wang et al., 2015). In AMI, miRNAs might affect the atherogenesis, a precursor for AMI by affecting the genes that regulate endothelial stability and atherosclerotic plaque destabilization. miRNAs might also affect the genes involved in the pathogenic pathway of AMI including apoptosis, necrosis, and autophagy. However, information on these theoretical roles of miRNA in young AMI is scarce. Therefore, it is important to dissect further miRNAs involvement in the pathogenesis of AMI in this young population.

Specific miRNAs are postulated to be involved in various stages of AMI pathogenesis in cell culture and animal studies (Ge et al., 2019; Guo et al., 2020; Han, Chen, Su, Zheng, Chen, Sun, Wu, Jiang, Xu, & Yang, 2019; Hao et al., 2020; Huang et al., 2020; Huangfu et al., 2020; Li et al., 2019b; Shi et al., 2020; Shin, Choi, Moon, Lee,

Park, Lee, Seo, Han, Lim, & Lee, 2019; Wang et al., 2020; Zhang et al., 2019a, 2019b). However, their complex regulatory mechanisms have not been completely understood (Schulte et al., 2017b). Though there are few studies in human looking at the involvement of these miRNAs in AMI but none of them studied on young AMI patients. There is a possibility that different miRNAs may be involved in the pathogenesis of AMI in this young population. Understanding the pathogenesis of AMI in this young group is very important in providing accurate diagnosis and prompt management of the disease. The discovery of miRNAs in the AMI pathogenesis in this young population could lead to their potential usage as novel biomarkers for detection of early cardiac injury, providing prognosis and predicting development of complications following AMI as well as for therapeutic intervention. Therefore, this warrants further studies in this area.

1.2 RESEARCH QUESTION

How miRNAs involve in the pathogenesis of AMI in young AMI group in our population and how do they affect the expression and translation of genes related to the pathophysiology of AMI in this young population?

1.3 GENERAL HYPOTHESIS

There is an involvement of miRNAs in AMI event of young adults in our population.

1.4 SPECIFIC HYPOTHESES

- i. There are specific miRNA profiles that are present in AMI patients in our population.
- ii. There are different pathways that involve in the pathogenesis of AMI in Young AMI group based on the significantly dysregulated miRNAs.
- iii. The miRNAs are differently dysregulated in Young AMI and Mature AMI.
- iv. The mRNA expressions of the dysregulated miRNAs in AMI event are differently dysregulated between Young AMI and Mature AMI.

1.5 GENERAL OBJECTIVE

The general objective of this study was to investigate the involvement of miRNAs in AMI of young adults in Kuantan, Pahang.

1.6 SPECIFIC OBJECTIVES

- v. To profile miRNAs in Young AMI and Mature AMI patients.
- vi. To identify the pathway involves in pathogenesis of AMI in Young AMI group based on the dysregulated miRNAs.
- vii.To compare the miRNAs that are dysregulated between Young AMI, Mature AMI, and Control group.
- viii.To measure the mRNA expressions of dysregulated miRNAs in AMI event between Young AMI and Mature AMI.