LOW-COST AND RAPID PROTOTYPING OF ELECTROCHEMICAL MICROFLUIDIC BIOSENSORS

BY

MOHD AFIQ BIN MOHD ASRI

A thesis submitted in fulfilment of the requirement for the degree of Master of Science (Electronics Engineering)

Kulliyyah of Engineering International Islamic University Malaysia

MARCH 2021

ABSTRACT

Electrochemical microfluidic biosensor is a widely used category of bioanalytical microdevices, with applications ranging from home-use glucometers to advanced blood analysis devices. They enable powerful microscale analyses in biology, physics and chemistry. Conventionally, the methods to fabricate these devices are either screen-printing, inkjet printing, or cleanroom-based photolithography. All these methods have slow iteration times, and cleanroom facilities are especially expensive and are limited in access to researchers in low-and-middle-income (LMIC) countries. In this thesis, a low-cost, accessible and rapid fabrication process of electrochemical microfluidic biosensors has been developed. This work leverages the accessibility of consumer-grade electronic craft cutters as the primary tool for patterning of sensor electrodes and microfluidic circuits, while commodity materials such as gold leaf, conductive silver ink, double-sided tape, vinyl sticker, plastic transparency films, and fabric adhesives are used as its base structural materials. The process enables fabrication of gold electrodes with dimensions as small as 450 µm and gaps of 110 µm, silver electrodes with dimensions as small as 600 µm, and fluid microchannels as small as 300 µm. Micro-volume hydrogen peroxide concentration measurements were performed as validation of biosensor performance, which achieved a limit of detection of 0.713 mM and sensitivity of 82.002 μ A mM⁻¹ cm⁻² from 2 μ L samples. The rapid process allows an iterative design-build-test cycle in less than 2 hours. This method is applicable in typical university laboratories and costs less than RM2100 to set up, enabling lower access barriers into the biosensor field for academic and industry researchers in low-resource settings.

خلاصة البحث

إنَّ المستشعرات الحيوية الكهروكيميائية المائعية الدقيقة هي إحدى فئات أجهزة التحليل الحيوي الدقيقة المستخدمة على نطاق واسع، وتطبيقاتما تتراوح بين أجهزة قياس السكر في الدم للاستخدام المنزلي وأجهزة تحليل الدم المتقدمة. هذه المستشعرات تمكّن من إجراء تحليلات فعّالة على المستوى المجهري في مجالات الأحياء والفيزياء والكيمياء. ومن الطرق المألوفة لتصنيع هذه الأجهزة استخدام طباعة الشاشة الحريرية، أو الطباعة النافثة للحبر، أو الطباعة الليثوغرافية الضوئية في غرف الأبحاث النظيفة. كل هذه الطرق لها أوقات تكرار بطيئة، كما أنّ مرافق غرف الأبحاث باهظة الثمن بشكل استثنائي، وتتّسم بمحدودية الوصول لها من قبل الباحثين في البلدان ذات الدخل المنخفض والمتوسط. في هذه الأطروحة، تم تطوير عملية تصنيع منخفضة التكلفة، وسهلة الوصول، وسريعة، للمستشعرات الحيوية الكهروكيميائية المائعية الدقيقة. يستفيد هذا العمل من إمكانية الوصول إلى أجهزة القطع الحرفية الإلكترونية المصممة للمستهلكين كأداة أساسية لتصميم وتشكيل أقطاب المستشعر ودوائر الموائع الدقيقة، بينما المواد الأولية مثل الأوراق الذهبية، والحبر الفضى الموصّل، والشريط اللاصق ذو الوجهين، ولاصق الفينيل، والأفلام الشفافة البلاستيكية، واللواصق القماشية، فإنها تستخدم كمواد هيكلية أساسية. وتتيح هذه العملية تصنيع أقطاب كهربائية ذهبية بأبعاد صغيرة تصل إلى 450 ميكرومتر، وفجوات مقدارها 110 ميكرومتر، وأقطاب فضية بأبعاد صغيرة تصل إلى 600 ميكرومتر، وقنوات مائعة صغيرة تصل إلى 300 ميكرومتر. وقد أجريت قياسات لتركيز بيروكسيد الهيدروجين ذات الحجم الصغير للتحقق من صحة أداء المستشعر البيولوجي، وحققت القياسات دقة في الكشف بلغت 0.713 مليمولار، وحساسية بمقدار 82.002 ميكروأمبير/مليمولار/سم²، وذلك باستخدام عينات بحجم 2 ميكرولتر. وتسمح هذه العملية السريعة بدورة (تصميم وبناء واختبار) تكرارية في أقل من ساعتين. هذه الطريقة قابلة للتطبيق في مختبرات الجامعة النموذجية وتكلفة تجهيزها أقل من 2100 رنجت ماليزي، مما يمكن من تذليل العقبات وتسهيل الوصول إلى مجال استخدام المستشعر الحيوي للباحثين الأكاديميين والصناعيين في الأماكن منخفضة الموارد.

ABSTRAK

Penderia elektrokimia mikrobendalir ialah salah satu kategori peranti bioanalisa mikro yang diguna secara meluas, dengan penggunaan yang merangkumi glukometer persendirian sehingga peranti analisa darah termaju. Alatan ini membolehkan analisa berskala mikro berkuasa tinggi untuk biologi, fizik dan kimia. Kaedah konvensional dalam pembuatan peranti-peranti ini bergantung kepada teknik percetakan skrin, teknik percetakan pancut dakwat, atau fotolitografi dalam bilik bersih (cleanroom). Semua teknik-teknik tersebut mempunyai kitaran ulang yang perlahan. Fasiliti bilik bersih terutamanya berkos tinggi dan mempunyai ketercapaian terhad bagi para penyelidik dari negara-negara berpendapatan rendah dan sederhana. Dalam tesis ini, proses pemprototaipan penderia elektrokimia mikrobendalir yang pantas, mudah capai, dan berkos rendah telah dihasilkan. Kajian ini mengeksplotasi kemudahperolehan penyurihpotong elektronik gred pengguna sebagai alat utama untuk pembentukan pola elektrod penderia dan litar mikrobendalir. Bahan-bahan komoditi seperti kerajang emas, dakwat perak konduktif, perekat dwimuka, pelekat vinil, kepingan plastik slaid lutsinar, dan perekat fabrik digunakan sebagai bahan struktur asas. Proses ini membolehkan fabrikasi elektrod emas dengan dimensi sekecil 450 µm dan sela selebar 110 µm, elektrod perak dengan lebar sekecil 600 µm, dan salur mikrobendalir sekecil 300 µm. Pengesahan prestasi biopenderia dijalankan melalui pengukuran kepekatan hydrogen peroksida berisipadu mikro berjaya mencapai had terendah pengesanan 0.713 mM dan kepekaan 82.002 μA mM⁻¹ cm⁻² dari sampel 2 μL. Proses pantas ini membolehkan kitaran rekabina-uji beriterasi dalam masa kurang dua jam. Kaedah ini terpakai di makmal universiti biasa dan berkos kurang RM2100 untuk dipasangsedia, sekaligus merendahkan sekatan penglibatan ke dalam bidang biopenderia bagi para penyelidik akademik dan industri dari sekitaran bersumber rendah.

APPROVAL PAGE

I certify that I have supervised and read this study and that in my opinion, it conforms to acceptable standards of scholarly presentation and is fully adequate, in scope and quality, as a thesis for the degree of Master of Science (Electronics Engineering).

Anis Nurashikin Nordin Supervisor Aliza 'Aini Md Ralib **Co-Supervisor**

Nabilah Ramli Co-Supervisor

Rosminazuin Ab Rahim Co-Supervisor

I certify that I have read this study and that in my opinion it conforms to acceptable standards of scholarly presentation and is fully adequate, in scope and quality, as a thesis for the degree of Master of Science (Electronics Engineering).

. .

Amelia Wong Azman Internal Examiner

Uda Hashim External Examiner

This thesis was submitted to the Department of Electrical and Computer Engineering and is accepted as a fulfilment of the requirement for the degree of Master of Science (Electronics Engineering).

> Mohamed Hadi Habaebi Head, Department of Electrical and Computer Engineering

This thesis was submitted to the Kulliyyah of Engineering and is accepted as a fulfilment of the requirement for the degree of Master of Science (Electronics Engineering).

Sany Izan Ihsan

Dean, Kulliyyah of Engineering

DECLARATION

I hereby declare that this thesis is the result of my own investigations, except where otherwise stated. I also declare that it has not been previously or concurrently submitted as a whole for any other degrees at IIUM or other institutions.

Mohd Afiq Bin Mohd Asri

INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

DECLARATION OF COPYRIGHT AND AFFIRMATION OF FAIR USE OF UNPUBLISHED RESEARCH

LOW-COST AND RAPID PROTOTYPING OF ELECTROCHEMICAL MICROFLUIDIC BIOSENSORS

I declare that the copyright holders of this thesis are jointly owned by the student and IIUM.

Copyright © 2021 Mohd Afiq Bin Mohd Asri and International Islamic University Malaysia. All rights reserved.

No part of this unpublished research may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder except as provided below

- 1. Any material contained in or derived from this unpublished research may be used by others in their writing with due acknowledgement.
- 2. IIUM or its library will have the right to make and transmit copies (print or electronic) for institutional and academic purposes.
- 3. The IIUM library will have the right to make, store in a retrieved system and supply copies of this unpublished research if requested by other universities and research libraries.

By signing this form, I acknowledged that I have read and understand the IIUM Intellectual Property Right and Commercialisation policy.

Affirmed by Mohd Afiq Bin Mohd Asri

Signature

.....15 / 3 / 2021..... Date

ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious and Most Merciful. All glory and praises are due to Him, through my times of ease and my times of difficulties.

Throughout my time at the International Islamic University Malaysia, I have been graced by the support from many people, some of which made this work possible, while many others made the journey bearable.

I particularly would like to thank my advisor, Prof. Dr. Anis Nurashikin Nordin, for providing me the opportunity for supervised research, and had granted me creative independence and provided the guidance and material resources to pursue a project of my own design. I am also grateful for the mentorship and the opportunities given to me to partake in the lab's day-to-day management, which helped me grow professionally.

Additionally, I would like to thank my co-supervisor, Asst. Prof. Dr. Nabilah Ramli, whom had generously augmented the financial and material resources for this project. I also thank Assoc. Prof. Dr. Raihan Othman, whom had allowed us free access to his laboratory, which was crucial to this work. I am also grateful to Alexandra Elbakyan for her development and advocacy of Sci-Hub, which has opened doors to many of us under-resourced researchers. My labmates who shared this journey have made my time in Anis Lab fulfilling: Fairuz, Dr. Anwar, Hakim, Farhan, Dr. Amalina, and Liyana, whom have helped me navigate through graduate school and engaged me intellectually and socially.

Most importantly, I would like to honor my family for their unconditional love and unyielding support – emotionally, financially, materially, and spiritually – throughout the highs and lows during this program and through personal tragedies that happened in between. I owe my gratitude to them for their prayers, their patience and for my upbringing. I am also most grateful to Norfatini for her friendship and emotional support throughout this journey.

Beyond the people I have honoured here, I acknowledge that far more have contributed and I am profoundly grateful to all.

TABLE OF CONTENTS

Abstract	i
Abstract (in Arabic)	.ii
Abstract (in Bahasa Malaysia)i	
Approval Pagei	iv
Declaration	vi
Copyrightv	ii
Acknowledgements	
Table of Contentsi	
List of Tablesx	
List of Figures	iii
List of Abbreviationsxv	
List of Symbols xvi	iii
, s	
CHAPTER 1: INTRODUCTION	.1
1.1. Introduction	
1.1.1. Electrochemical Microfluidic Biosensors	.1
1.1.2. Conventional and New Approaches to Microfabrication	.2
1.1.3 The Frugal Approach to Science Tools	.3
1.2. Problem Statement.	
1.3. Hypothesis	
1.4. Research Methodology	
1.5. Research Scope	
1.6. Research Objectives	
1.7. Thesis Organisation	
1	
CHAPTER 2: LITERATURE REVIEW	.9
2.1. Concept Fundamentals	
2.1.1. Biosensors	
2.1.2. Electrochemical Biosensors	
2.1.3. Microfluidic Biosensors	
2.1.4. Costs and Frugality in Microdevices Prototyping	
2.2. Approaches to Prototyping Microfluidic Devices	
2.2.1. Methods Using Photolithography and Industrial Machineries1	
2.2.2. Low-Cost and Desktop-Compatible Methods	
2.3. Approaches to Prototyping Printed Electronics	
2.3.1. Conventional Thin Film Fabrication and PCB Manufacturing2	
2.3.2. Conductive Ink-Based Printing	
2.3.3. Non-Conventional Techniques	
2.4. Integrated Electrochemical Microfluidic Biosensors	
2.4.1. Relevant Approaches for Characterisation of Electrochemical	10
Microfluidic Biosensors4	10
2.4.1.1. Baseline Parameter Characterisation	
2.4.1.2. Hydrogen Peroxide Sensing	
2.4.2. Examples of Desktop-Fabricated Biosensors with Integrated	10
Electronics and/or Microfluidics	13
	5

2.5. Advantages and Limitations of Frugal Prototyping	51
2.6. Project Design: Considerations and Benchmarks	
2.7. Chapter Summary	
CHAPTER 3: EXPERIMENTAL METHODS	56
3.1. Overview	56
3.2. Tools, Materials and Reagents	
3.2.1. Tools and Materials	
3.2.2. Reagents	
3.2.3. Optimising Tool Parameters	
3.3. Microfluidic Fabrication Process Development	
3.3.1. Selection of Materials for Lamination	
3.3.2. Microfluidic Circuit Fabrication	
3.3.3. Microfluidic Characterisation	
3.3.3.1. Burst Pressure Test	
3.3.3.2. Reynolds Number Analysis	
3.3.3.3. Test for Laminarity	
3.4. Electrode Fabrication Process Development	
3.4.1. Fabrication of Gold Leaf Electrodes	
3.4.1.1. Selection of Adhesives for Gold Leaf Mounting	
3.4.1.2. Reproducing Vinyl Stencilling Technique as Baselir	
5.1.1.2. Reproducing vinyi Stehenining Teeninque us Buseni	
3.4.1.3. Improved Fabrication Process Using Gold Leaf	
Xurography	68
3.4.1.4. Characterisation of Geometric Features of Gold Lea	
Devices	
3.4.2. Silver Reference Electrode Using Circuit Scribe Silver	/ 1
Conductive Pen	72
3.5. Fabrication of Electrochemical Microfluidic Device	
3.6. Validation of Baseline Electrochemical Properties	
3.7. Quantitative Measurement of Chemical and Biological Analytes	
3.7.1. Determination of Calibration Curve of Hydrogen Peroxide	70
Concentration	78
3.7.2. Determination of Calibration Curve of Glucose Concentration	
3.8. Cost and Rapidity Analysis	
3.9. Chapter Summary	
5.9. Chapter Summary	01
CHAPTER 4: RESULTS AND DISCUSSION	87
4.1. Development of Novel Process of Prototyping Electrochemical	02
Microfluidic Biosensor using Consumer Tools and Commodity Material	ູຈາ
4.1.1. Design of Electrochemical Microfluidic Biosensor	
-	
4.1.2. Summary of Fabrication Parameters and CAD Design Rules.	
4.1.3. Selection of Adhesive for PET–Gold Leaf Bonding	
4.1.3. Rationale for Selection of Tools and Materials	
4.2. Geometric Characterisation of Electrodes	
4.2.1. Gold Leaf Electrodes	
4.2.1.1. Resolution of Gold Leaf Electrodes	
4.2.1.2. Surface Profilometry Analysis	
4.3.2. Silver Electrodes	
4.3. Characterisation of Heat'N'Bond-Laminated Microfluidic Device	101

4.3.1. Burst Pressure Test	101
4.3.2. Reynolds Number Analysis	103
4.3.3. Laminarity of Flow	103
4.4. Electrochemical Characterisation of Integrated Microfluidic Biosens	sor by
Cyclic Voltammetry	104
4.4.1. Electrochemistry Using Integrated Sensors	104
4.4.2. Comparison to Commercial Sensors	107
4.5. Quantitative Measurement of Analyte Concentration by Amperomet	ry110
4.5.1. Determination of Peak Potential	110
4.5.2. Constructing Calibration Curve for Hydrogen Peroxide	112
4.5.3. Constructing Calibration Curve for Glucose	114
4.6. Cost and Rapidity Analysis	116
4.6.1. Bill of Materials and Cost Analysis	116
4.6.2. Rapidity of Prototyping Analysis	119
4.6.3. Process Yield	121
4.7. Advantages and Limitations	121
4.8. Concluding Remarks	124
CHAPTER 5: CONCLUSIONS AND RECOMMENDATIONS	125
5.1. Conclusions	
5.2. Significance to the Field	126
5.3. Future Work	
REFERENCES	129
PROJECT ACKNOWLEDGEMENTS	145
LIST OF PUBLICATIONS	146
APPENDIX A: CLEANROOM SERVICE FEES IN MALAYSIA	147
APPENDIX B: SURFACE PROFILER ANALYSIS	149

LIST OF TABLES

Table 2.1	Summary of low-cost techniques for the fabrication of microfluidic devices.	27
Table 2.2	Summary of low-cost techniques for the fabrication of printed electronics.	38
Table 2.3	Summary of relevant prior works.	49
Table 3.1.	Summary of Silhouette Cameo® settings for specific combination of materials.	61
Table 3.2.	Workflow summary for the improved gold electrode fabrication developed in this work.	70
Table 4.1.	Resolution limits for different combinations of materials.	86
Table 4.2.	List of the tested adhesives and its outcomes on adhesion on PET substrate through a vinyl stencil and its processability within the manufacturing process framework.	89
Table 4.3.	Measured width of gaps between finger of interdigitated electrodes from 0.5 mm IDTs (n=5) and 1.0 mm IDTs (n=3).	98
Table 4.4.	Corresponding flow rates at which leakage events occurred for each measured device.	102
Table 4.5.	Apparent fluid front velocity measured from video analysis of passive dye flow in microchannels, and corresponding Reynolds number.	103
Table 4.6.	Measured peak separation for scan rates from each replicate (n=3) from ferrocyanide redox cycling on the electrochemical microfluidic biosensor device.	106
Table 4.7.	Table comparing important parameters from the sensor in this work and DropSens SPGE.	109
Table 4.8.	Bill of materials and breakdown of upfront cost for setting up system.	116
Table 4.9.	Cost estimate per unit for an electrochemical microfluidic biosensor in this work.	117
Table 4.10	. Estimated time taken for the steps involved in the fabrication process.	119

LIST OF FIGURES

Figure 1.1.	Flow chart visualising the research methodology.	6
Figure 2.1.	Selected illustrations of low-cost fabrication processes for microfluidic devices.	26
Figure 2.2.	Selected illustrations of low-cost printed electronics devices, with focus on unconventional techniques.	36
Figure 2.3.	Illustrations depicting relevant prior works.	48
Figure 3.1.	Conceptual illustration of a frugal and rapid prototyping of electrochemical microfluidic biosensor.	57
Figure 3.2.	(a) A Silhouette Cameo® 3 desktop electronic plotter-cutter. (b) Gold leaf spatula made from popsicle stick.	59
Figure 3.3.	Summary of fabrication process of microfluidic circuits.	63
Figure 3.4.	Experimental setup for burst pressure test.	65
Figure 3.5.	Workflow summary for gold electrode fabrication based on Kao et al.'s technique.	68
Figure 3.6.	Conceptual diagram of surface profilometry analysis to determine gap size.	72
Figure 3.7.	Drawing silver circuits directly onto the substrate using the Silhouette Cameo® 3 and the Circuit Scribe silver conductive pen.	73
Figure 3.8.	Layout of individual device layers and their corresponding cut/plot sequence and settings in Silhouette Cameo®.	75
Figure 3.9.	 (a) Individual device layers prior to lamination and dicing. (b) Alignment of layers for assembly using header pins and masking tape. (c) Laminating the assembly using aluminum foil as carrier. (d) Dicing batch into individual devices using scissors. (e) Device after lamination and dicing. Inset: Malaysian 50 sen coin for scale. 	75
Figure 3.10.	Physical connections of the electrochemical microfluidic biosensor device to the Autolab PGSTAT128N potentiostat.	76
Figure 4.1.	Workflow of the rapid fabrication process for the biosensor.	83
Figure 4.2.	Design schematics of the electrochemical microfluidic biosensor device used for validation tests.	84
Figure 4.3.	(a) Illustration showing examples of design rules regarding minimum distance between vinyl and gold leaf cut lines (500 μ m), and minimum stencil opening of 2 mm. (b) Illustration	

	showing cut lines (light blue) added to reduce the size of peeled vinyl stencils. The stencil will be peeled along red and light blue lines. (c) Cross sectional illustration of the design rules in (a).	88
Figure 4.4.	Representative brightfield microscopy image of a gold leaf electrode consisting of a working electrode, a counter electrode, and inter-electrode gap. Scale bar is 500 μ m. No ruptures are observed in the middle region of the electrode. Wrinkled electrode are natural features of a gold leaf electrode, which increases its surface area.	93
Figure 4.5.	Comparison between thin rectangular electrodes (2 mm long) fabricated using vinyl stencilling (Mont-Marte gilding adhesive) versus tape-mounted, cut gold leaf (Teraoka 7070W).	94
Figure 4.6.	(a) Illustration of the possible formation of gold leaf rupture mid-electrode from the use of vinyl stencilling strategy to pattern the gold leaf electrodes. (b) Mounting gold leaf using ultrathin tape reduces the likelihood of gold leaf breaking in the middle of electrodes.	96
Figure 4.7.	Surface profilogram for gold leaf interdigitated electrodes with finger width 0.5 mm. (Inset) Image and sketch of IDT device used for the measurement. Red arrow indicate direction of profilometry.	97
Figure 4.8.	Surface profilogram for gold leaf interdigitated electrodes with finger width 1.0 mm. (Inset) Image and sketch of IDT device used for the measurement. Red arrow indicate direction of profilometry.	97
Figure 4.9.	(a) An electrical trace fabricated using Circuit Scribe conductive pen, overlaid with laser-printed lines. The electrical trace matches the linewidth of 550 μ m lines. (b) Measurement of minimum gap size required to be electrically isolating.	100
Figure 4.10.	Flow of dye in a microchannel during burst pressure test, through the viewing window of the burst pressure test rig.	102
Figure 4.11.	Image of dye flow merging in microchannel with pristine PET as its surface. RGB color profilogram across the width of the microchannel, showing that there is no mixing between the separated dye flow due to its laminar properties.	104
Figure 4.12.	(a) Representative cyclic voltammogram of 5 mM ferrocyanide solution using electrochemical microfluidic biosensor, at various scan rates, recorded over 5 cycles. Potential is applied against Ag pseudoreference electrode. (b) Plot of peak current density versus square root of scan rate. Error bars represent one standard deviation of the replicates ($n = 3$).	106
Figure 4.13.	(a) CV profile comparison of gold leaf electrochemical microfluidic biosensors versus commercial DropSens SPGE in	

xiv

5 mM potassium ferrocyanide in 0.1 M KCl solution at 100 mV/s. The last 3 consecutive cycles are plotted side-by-side. The current output is normalised to geometric surface area of working electrode.

- Figure 4.14. (a) Cyclic voltammogram (n=1) of PBS and 1 mM hydrogen peroxide in PBS across a wide potential range. Both curves have similar profiles at V > 0, however shows different profiles at V < 0.
- Figure 4.15. Signal-to-background ratio of one-way (linear sweep) voltammetry from 0 V to -1.0 V extracted from the CV profile. The S/B ratio has a maximum of 2.56, at V = -0.479 V vs. Ag QRE.
- Figure 4.16. (a) Representative amperogram for measurement of H2O2 solutions at -0.45 V with concentrations 313 μ M, 625 μ M, 1.25 mM, 2.5 mM, 5 mM, 7.5 μ M and 10 mM, and blank. (b) Calibration curve for H2O2 measurement, current value is recorded at 60 s. Error bars represent one standard deviation of the replicates (n = 3).
- Figure 4.17. (a) Representative amperogram for measurement of glucose solutions at -0.45 V with concentrations 625 μ M, 1.25 mM, 2.5 mM, 5 mM, 7.5 μ M and 10 mM, and blank. (b) Calibration curve for glucose measurement, current value is recorded at 60 s. Error bars represent one standard deviation of the replicates (n = 3).

115

113

108

111

111

LIST OF ABBREVIATIONS

μTAS	Micro-total analysis system
$[FeCN_6]^{3-/4-}$	Ferri/ferrocyanide redox couple ion
μPAD	Micro-paper analytical devices
24K	24 karats
2DPN	Two dimensional paperfluidic networks
3D	Three dimensional
AgCl	Silver chloride
AgNP	Silver nanoparticles
ASSURED	Affordable, Sensitive, Specific, User-friendly, Rapid and robust, Equipment-free, and Deliverable to end users
CAD	Computer Aided Design
CE	Counter Electrode
CMOS	Complementary metal-oxide semiconductor
CNC	Computer numerical control
COC	Cyclic olefin copolymer
CV	Cyclic voltammetry
DI	Deionised water
DIW	Direct ink writing
ELISA	Enzyme-linked immunosorbent assay
ESCARGOT	Embedded SCAffold RemovinG Open Technology
FDM	Fused deposition modelling
GOx	Glucose oxidase
H_2O_2	Hydrogen peroxide
HNB	Heat 'N' Bond iron-on adhesive
IDT	Interdigitated electrodes
IJP	Inkjet printing

IPA	Isopropyl alcohol
LDI	Laser desorption ionisation
LFA	Lateral flow assay
LMIC	Low- and middle-income countries
LOC	Lab on a chip
LOD	Limit of detection
MEMS	Microelectromechanical systems
PAAm	Polyacrylamide
PBS	Phosphate buffered saline
PCB	Printed circuit board
PCL	Print-cut-laminate
PCL	Polycaprolactone
PDMS	Polydimethylsiloxane
PET	Polyethylene terephthalate
PLA	Polylactic acid
PMMA	Polymethyl methacrylate
POC	Point of care
POCT	Point of care testing
PSA	Pressure sensitive adhesive
PVA	Polyvinyl acetate
PVD	Physical vapor deposition
R&D	Research and development
RACER	Research Acculturation of Early Career Researchers
RE	Reference electrode
RIGS	Research Incentive Grant Scheme
SAM	Self-assembling monolayer
VIA	Vertical interconnect access
WE	Working electrode

LIST OF SYMBOLS

μ	Dynamic viscosity, N m ⁻² s
$\mu_{ m blank}$	Mean value of blank measurement, A
A	Cross-sectional area, m ²
С	Concentration, M / mol cm ⁻³
D	Diffusion coefficient, cm ² s ⁻¹
$E_{1/2}$	Half-cell potential, mV
E _{p-p}	Peak-to-peak separation, mV
i_p	Peak current, A
I_p*	Specific polar moment of inertia
J	Peak current density, A mm ⁻¹
L	Characteristic length, m
l	Length, m
n	Number of replicates
Q	Flow rate, m ³ s ⁻¹
R	Electrical resistance, Ω
r^2	Coefficient of determination
Re	Reynolds number
и	Fluid front velocity, m s ⁻¹
ν	Kinematic viscosity, m ² s ⁻¹
ν	Scan rate, V s ⁻¹
V	Applied potential, V
W	Width, m
ΔE_p	Peak separation, mV
ΔP	Pressure drop, N m ⁻²
θ	Angle, °
$\sigma_{ m blank}$	Standard deviation of blank measurement, A

CHAPTER 1

INTRODUCTION

1.1. INTRODUCTION

Micro-total analysis systems (μ TAS), more commonly known as "lab-on-a-chip" (LOC), is a powerful emerging technology used for analytical applications in biology, chemistry and physics. These devices exploit the special physical properties in nature at the microscale, such as laminar flow and diffusion-dominated kinetics to engineer features such as low resource consumption, rapidness, and high precision when doing analytical techniques. The use of LOC enables multiple applications across the research laboratory, including biological/chemical analysis, chemical synthesis, high-throughput screening, precise liquid manipulation and creation of new tools to pursue novel scientific questions. Additionally, they expand the laboratory capabilities into the clinic and in the field outside of the laboratory (Kovarik et al., 2013).

1.1.1. Electrochemical Microfluidic Biosensors

One such category of LOC devices are electrochemical microfluidic biosensors, which is of high interest in the fields of point-of-care diagnostics, clinical chemistry, environmental monitoring, and precision cellular and molecular analysis. Electrochemistry-based biosensors holds several advantages over their optical-based and electromechanical-based counterparts, but most particularly for its relatively inexpensive instrumentation (compared to optical-based biosensing) and reduced unit cost at scale (compared to electromechanical-based biosensing) (Rackus et al., 2015). The most commonly used and most commercially successful example of electrochemical microfluidic biosensor is the glucometer (Turner, 2013).

1.1.2. Conventional and New Approaches to Microfabrication

The majority of LOC are made using cleanroom-associated technologies, which was first developed for fabrication of semiconductor devices such as diodes and transistors, and later adopted by the micro-electromechanical systems (MEMS) in the 1980s that produces accelerometers, miniature pressure and temperature sensors and GPS integrated devices (Reyes et al., 2002). Among cleanroom-associated equipment are photolithographic mask aligners and thin film deposition machines, such as metal sputtering chambers and plasma-enhanced chemical vapor deposition. Cleanrooms are not necessarily easy to access, especially to researchers in low-and-middle-income countries (LMIC), and the facilities and equipment involved are expensive (Pan & Wang, 2011; Walsh et al., 2017). Additionally, with biosensors, often there are regionspecific modifications on the device involving bio-recognition capture molecules, such as immobilised proteins, antibodies, and nucleic acid hybridisation probes. These molecules are often functionalised onto the device or sensors using microarray spotter, a costly robotic instrumentation meant for customising nucleic acid microarrays (Park et al., 2008). Each of these instruments may cost anywhere between RM 250,000 to RM 3 million, and even membership access to the few available cleanroom facilities may cost from RM 5000 to RM 10,000 per annum in Malaysia, non-inclusive of equipment per use basis fees. These associated costs and access barriers hinder prototyping through iterative design process, which inadvertently delays product delivery and discourages LOC development and applications in LMICs.

To overcome these financial and access barriers, several independent works have been developed to build electrochemical sensors and microfluidic devices, including gold leaf lamination (Thompson, Birch, Nelson, et al., 2016), inkjet printing (Kawahara et al., 2014), heat-sensitive adhesive lamination (Birch et al., 2017), and xurography (Bartholomeusz et al., 2005; Martínez-López et al., 2016; Yuen & Goral, 2010) – all to a certain degree of limitations in geometric resolution, material versatility, and complexity.

1.1.3 The Frugal Approach to Science Tools

The concept of frugal science and innovation has recently gained momentum. The approach of 'constraint-based science', often starting by asking questions about costs, accessibility and inclusivity, leads to an innovator's ability to reframe problems and solutions (Ahuja, 2014; Reardon, 2013). Some of the prominent frugal innovations in science tools include the 50 cents paper microscope i.e. the Foldscope (Cybulski et al., 2014), a 125,000 rpm paper centrifuge i.e. Paperfuge (Bhamla et al., 2017), a gas lighter-based electroporator i.e. ElectroPen (Byagathvalli et al., 2020), a nebuliser powered by bicycle pump (Dzwonczyk et al., 2015), and a solar-powered medical oxygen concentrator (Hawkes et al., 2018). These tools have already revolutionised chemical and life sciences, as well as being used in real world applications.

Within the field of microdevices and biosensors, a common approach is to develop "tools to create tools", usually kits containing modular parts for non-specialists to build custom microfluidic circuits and diagnostic kits. A few well known example of this is Ampli, which is based on laser-cut lateral flow assay modules (Phillips et al., 2018), and micromachined Lego bricks (Owens & Hart, 2018). For microfabrication in

general, several 'cleanroom-to-makerspaces' approaches have been introduced, as reviewed by Walsh et al (Walsh et al., 2017).

This research seeks to explore a combination of these various techniques to compensate each techniques' limitation, to develop a novel, composite process to fabricate electrochemical microfluidic devices.

1.2. PROBLEM STATEMENT

Lab-on-a-chip (LOC) are emerging technologies that has been enabling powerful microscale analyses in biology, physics and chemistry. This technology has given rise to point-of-care (POC) diagnostics, single cell-associated physiological studies, and rapid, low-consumption chemical/bio-reactor systems. Most LOC components such as sensors and microfluidic circuits rely on traditional microfabrication methods associated with cleanrooms, most notably photolithography and sputtering techniques; and/or robotic handling methods such as microarray spotting. Cleanrooms are not necessarily easily accessible, especially to researchers in low-and-middle-income (LMIC) countries, and the facilities and equipment involved are expensive for early stage prototyping. In Malaysia, the rental costs for cleanrooms may go up to RM7500 per annum, and fees for use of equipment may range between RM50 to RM300 per hour or process (see Appendix A).

Alternatives to cleanroom-based prototyping include industrial and researchgrade material inkjet printers such as the Dimatix DMP-2800 series, and screenprinting. The Dimatix printer, while overall cheaper than a cleanroom, may cost up to RM300,000 (conservative estimate), which is still cost limiting for majority of researchers. Furthermore, the Dimatix involve a complex process optimisation for each given type of ink and substrate (A. A. Zainuddin et al., 2017). Meanwhile, screenprinting enables a low-cost mass manufacturing option for fabrication of sensors. However, during early prototyping phase where iterative design is often required, fabrication of masks for screen-printing is a time-consuming process which may take several hours (if fabricated in-house) to over a week (if made to order). Additionally, both methods require functional inks which are either custom formulated in-house or purchased from specialty manufacturers which are often expensive. Specific to electrochemical sensors, another alternative is present in commercial screen-printed electrodes, such as ones commercialised by DropSens. A limitation presented by these commercial sensors is that they are sold according to manufacturer specified designs, which are non-customisable and not necessarily integrable into microfluidic systems.

These associated costs and access barriers extend the turnaround time of each iteration during the device prototyping phase, which delays product completion and delivery. Given that the typical seed funding for academic research in countries such as Malaysia has a small quantum – the Research Incentive Grant Scheme (RIGS), for example, funds at RM20,000 over two years, with only RM5000 allocated for materials and supplies – exacerbating the need for frugal approaches to microfabrication for those intending to pursue such endeavor.

1.3. HYPOTHESIS

We hypothesise that a systematic combination of various frugal approaches will enable a reasonable alternative to the cleanroom-based techniques in miniaturised chemical and biological systems at a significantly lower cost, with more accessible set of instrumentation.