EFFECTS OF TRIHONEY ON REPRODUCTIVE DYSFUNCTIONS IN HIGH CHOLESTEROL DIET-FED MALE RABBITS

BY

ZENAB B. HAMAD MOHAMED

A thesis submitted in fulfilment of the requirement for the degree of Doctor of Philosophy in Health Sciences

Kulliyyah of Allied Health Sciences International Islamic University Malaysia

SEPTEMBER 2020

ABSTRACT

Overconsumption of high-cholesterol diet induces hypercholesterolemia and disturbs cholesterol homeostasis in the body which adversely affects normal male reproductive functions. Use of honey has become of increasing interest due to the increase in the availability of evidence-based findings demonstrating the beneficial effects of honey in treating diverse diseases. The present study was undertaken to evaluate the potential protective effects of Trihoney (a mixture of Trigona, Mellifera and Tualang) against male reproductive dysfunctions in diet-induced hypercholesterolemic rabbits and compare its effects with atorvastatin. Forty-eight male New Zealand white rabbits at the age of 5 months were assigned into 6 groups. Two groups were fed commercial rabbit pellet and 0 and 0.6 g/kg/day of Trihoney respectively. The other four groups were fed 1% cholesterol diet and 0, 0.3, 0.6 g/kg/day of Trihoney, and 2 mg/kg/day of atorvastatin for 12 weeks. The study was planned in 5 distinct phases. The purpose of the first phase was to evaluate the effects of Trihoney on serum lipid profile and serum and testicular malondialdehyde (MDA) and antioxidant enzymes; superoxide dismutase (SOD) and glutathione peroxidase (GPx). Trihoney and atorvastatin reduced serum total cholesterol and LDL-c significantly. Trihoney was as effective as atorvastatin in the lipid lowering effect. Trihoney slightly reduced serum MDA but significantly enhanced serum SOD and GPx. It reduced testicular MDA and increased SOD significantly. Atorvastatin treatment significantly reduced serum and testicular MDA and enhanced serum and testicular SOD and GPx. In the second phase, the effect of Trihoney on serum inflammatory biomarkers was evaluated. Trihoney administration reduced serum levels of IL-6, TNF-α and IL-1β significantly. Atorvastatin reduced serum TNF-α and IL-1β significantly. In the third phase, the effects of Trihoney on serum and intra-testicular testosterone, serum FSH, serum LH, fasting insulin, fasting blood glucose and HOMA-IR were investigated. Trihoney particularly at the dose of 0.6 g/kg/day significantly improved serum and intratesticular testosterone and serum FSH; whereas, atorvastatin showed no improvement in these hormones. Both Trihoney and atorvastatin showed no effects on fasting serum insulin, fasting blood glucose and HOMA-IR. The fourth phase was aimed to evaluate the effects of Trihoney on sperm parameters. Trihoney particularly at the dose of 0.6 g/kg/day improved the percentages of sperm motility and sperm with normal morphology as well as reduced the percentages of immotile sperm and sperm with abnormal morphology. Trihoney improved sperm concentration but with no statistical significant. Atorvastatin group showed the worst outcome of sperm parameters. In the fifth phase, the effects of Trihoney on testicular and epididymal histopathological changes were evaluated. Trihoney ameliorated the testicular degenerative changes, improved spermatogenesis and maintained the normal histology of the epididymis with an increase in the number of sperm in its tubules. Atorvastatin treated group showed severe testicular tubular degenerative changes and epididymal atrophy with fibrosis. In conclusion, Trihoney showed its potential health benefits as an effective hypocholesterolemic, anti-inflammatory and antioxidant agent. It was shown to improve sperm parameters and male reproductive hormones, and attenuate testicular and epididymal histopathological alterations in high-cholesterol diet fed male rabbits. Hence, Trihoney plays a favourable role on several mechanisms involved in combating hypercholesterolemia-induced male reproductive dysfunctions.

خلاصة البحث

يؤدي الإستهلاك المفرط لغذاء عالى الكوليسترول إلى إرتفاع كوليسترول الدم وإختلال توازن الكوليسترول في الجسم مما يؤثر سلباً على الوظائف التناسلية للذكور.أصبح إستخدام العسل ذا أهمية متزايدة بسبب زيادة توافر الدلائل العلمية التي تُبيّن فوائد العسل. أُجريت هذه الدراسة لتقييم التأثير الوقائي المحتمل للعسل الثلاثي ضد ضعف القدرة الإنجابية للذكور والناتجة عن إرتفاع كوليستيرول الدم في الأرانب ومقارنته بالأتورفاستاتين. ثمانية وأربعون من ذكور الأرانب البيضاء النيوزيلاندية قُسمت إلى 6 مجموعات. غُذّيت مجموعتان بغذاء الأرانب التجاري مع 0 و0.6 جمر/كجم/يوم من العسل على التوالي بينما غُذّيت المجموعات الأربع الأخرى على غذاء عالي الكوليستيرول مع 0 و0.3 و0.6 جم/كجم/يوم من العسل و2 مجم/كجم/يوم من الأتورفاستاتين. قُسِّمت هذه الدراسة إلى خمس مراحل. هدفت المرحلة الأولى لدراسة تأثير العسل على مستوى الدهون ومؤشر الإجهاد التأكسدي والإنزيمات المضادة للأكسدة في مصل الدم والخصيتين. كان تأثير العسل مساو للأتورفاستاتين في خفض الكوليستيرول الكلى والكوليستيرول الضار. كانت الزيادة في الإنزيمات المضادة للأكسدة في مصل الدم أفضل في مجموعات العسل بينما أظهر الأتورفاستاتين أكثر تأثيراً في الخصيتين. في المرحلة الثانية دُرس تأثير العسل على المؤشرات الحيوية الإلتهابية في مصل الدم. خفّض كل من العسل والأتورفاستاتين من مستويات المؤشرات الحيوية الإلتهابية في مصل الدم. في المرحلة الثالثة، فُحص تأثير العسل على الهرمونات التناسلية الذكرية في مصل الدم والخصيتين، وعلى مؤشر مقاومة الإنسولين. حسَّن العسل خاصة بجرعة جم/كجم/يوم هرمون التستوستيرون وهرمون تحفيز الجريب. لم يُظهر الأتورفاستاتين أي تحسن في 0.6الهرمونات. لم يُؤثر العسل ولا الأتورفاستاتين على مؤشر مقاومة الإنسولين. في المرحلة الرابعة قُيّم تأثير العسل على الحيوانات المنوية. العسل الثلاثي خاصة بجرعة 0.6جم/كجم/يوم أثر إيجابياً على صفات الحيوانات المنوية بينما أحدث الأتورفاستاتين أسوأ النتائج. المرحلة الخامسة قيّمت تأثير العسل على التغيرات النسيجية في الخصيتين والبربخ. أحدث العسل الثلاثي تحسيناً في التغيرات التنكسية للخصية وفي تكوين الحيوانات المنوية والحفاظ على الأنسجة الطبيعية للبربخ. التغيرات النسيجية في الخصيتين والبربخ كانت أكثر شدة في مجموعة الأتورفاستاتين. بناءاً على ماسبق: أظهر العسل الثلاثي فوائده الصحية كخافض لكوليستيرول الدم ومعزز للإنزيمات المضادة للأكسدة، ومثبط للمؤشرات الحيوية الالتهابية، ومحسّن للحيوانات المنوية والهرمونات التناسلية الذكرية ومخفف من التغيرات النسيجية للخصيتين والبربخ.

APPROVAL PAGE

The thesis of Zenab B. Hamad Mohamed has been approved by the following:

Muhammad Bin Ibrahim
Supervisor
Che Anuar Che Mohamad
Co-Supervisor
Suzanah Binti Abdul Rahman
Internal Examiner
Mahaneem Mohamed
External Examiner
Zarina Bt Zainuddin
Chairman

DECLARATION

I hereby	declare	e tha	t this	thesis is	the	resi	ult of	my	own	investig	ations,	except	where
otherwis	e state	d. I	also	declare	that	it	has	not	been	previou	ısly or	concu	rrently
submitte	d as a v	vhole	e for a	ıny other	degr	ees	at III	JM	or oth	ner instit	utions.		

Zenab B. Hamad Mohamed	
Signature	Date

INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

DECLARATION OF COPYRIGHT AND AFFIRMATION OF FAIR USE OF UNPUBLISHED RESEARCH

EFFECTS OF TRIHONEY ON REPRODUCTIVE DYSFUNCTIONS IN HIGH CHOLESTEROL DIET-FED MALE RABBITS

I declare that the copyright holders of this thesis are jointly owned by the student and IIUM.

Copyright © 2020 Zenab B. Hamad Mohamed and International Islamic University Malaysia. All rights reserved.

No part of this unpublished research may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder except as provided below

- 1. Any material contained in or derived from this unpublished research may be used by others in their writing with due acknowledgement.
- 2. IIUM or its library will have the right to make and transmit copies (print or electronic) for institutional and academic purposes.
- 3. The IIUM library will have the right to make, store in a retrieved system and supply copies of this unpublished research if requested by other universities and research libraries.

By signing this form, I acknowledged that I have read and understand the IIUM Intellectual Property Right and Commercialization policy.

Affirmed by Zenab B. Hamad Mohamed	
Signature	Date

ACKNOWLEDGEMENTS

In the name of Allah, the Most Merciful Most Compassionate. May His peace and blessings be upon our beloved Prophet Mohammad (PBUH), his household, companions and those who follow them with sincerity till the day of reckoning.

Foremost, I would like to offer this endeavour to Allah Almighty for the wisdom he bestowed upon me, the strength, peace of my mind and good health in order to finish this research as a requirement for the degree of Doctor of Philosophy in Health Sciences-Nutrition Sciences.

I would like to dedicate this work to my parents; to my father who I adore and has left me forever, my father thank you so much for everything you have done to grow me up and upbringing me sound breeding to reach this level. My father you will stay in my heart forever and I will not stop praying for you. May Allah gather me with you in His Paradise. To my mother who I adore, thank you so much for your patience and abundant emotions that you have carried to me. May Allah protect you and give you the health and the strength.

I am sincerely grateful to my supervisor, Assoc. Prof. Dr. Muhammad Bin Ibrahim for his invaluable guidance, sincerity, encouragement and continuous support. My special thanks and appreciation to all my co-supervisors, Asst. Prof. Dr. Azantee Yazmie Abdul Wahab, Asst. Prof. Dr. Azliana binti Abd Fuaat and Asst. Prof. Dr. Che Anuar Che Mohamad. May your morale, wisdom, knowledge and piety continue to wax and wax forever.

I would like to express my deepest gratitude towards my family for the motivation and support. My beloved and supportive husband, Hamad Abdulsalam Alfarisi who is always by my side when times I needed him most and helped me, and my adorable kids, Hothaifa and Mohammed who served as my inspiration to pursue this research and their endurance with me for lots of the hard days during my study journey. May Allah preserve you all for me and Islam. I would like to extend the heartfelt gratitude to all of my siblings for their continuous support and motivation.

I would also like to express my sincere gratitude to International Islamic University Malaysia for funding part of this research (IRAGS18-043-0044). My utmost appreciations to the dean and staff of CPS for granting me IIUM postgraduate president assistantship (Semester 1, 2018 & 2019).

Lastly, special thanks and appreciations go to the entire staff of Kulliyyah of Allied Health Science, Central Research and Animal Facility (CREAM, IIUM), Basic Medical Sciences at Kulliyyah of Pharmacy and Histopathology and Laboratory Medicine at IIUM Medical Centre for providing the facilities and the suitable working environment during my lab work.

TABLE OF CONTENTS

Abstractii
Abstract in Arabiciii
Approval Pageiv
Declarationv
Acknowledgements vii
Table of Contentsviii
List of Tablesxv
List of Figuresxvii
List of Abbreviationsxix
List of Symbolsxxii
CHAPTER ONE: INTRODUCTION1
1.1 Background of the Study1
1.2 Statement of the Research Problem and Significance of the Study3
1.3 Research Objectives
1.3.1 General Objective6
1.3.2 Specific Objectives6
1.4 Research Questions
1.5 Research Hypothesis
1.5 Research Hypothesis
CHAPTER TWO: LITERATURE REVIEW9
2.1 Male Infertility
2.1.1 Overview and Epidemiology
2.1.2 Normal Physiology of Male Reproductive System
2.1.3 Causes of Male Infertility
2.1.3.1 Azoospermia
2.1.3.1.1 Obstructive Infertility
2.1.3.1.2 Non-obstructive Infertility
2.1.3.2 Coital Infertility
2.1.3.3 Oxidative Stress and Male Infertility
2.1.3.4 Inflammation and Male Infertility
2.1.3.5 Nutrition and Male Infertility
2.1.3.6 Hormonal Imbalance
2.1.4 Evaluation of Male Infertility
·
2.1.4.1 Semen Analysis
2.1.4.1.1 Specialized Clinical Tests on Semen and Sperm23
2.1.4.2 Endocrine Evaluation
2.1.4.3 Post Ejaculatory Urine Analysis
2.1.4.4 Ultrasonography
2.1.4.5 Testicular Biopsy
2.1.4.6 Microbiologic Assessment
2.1.4.7 Genetic Screening
2.1.5 Treatment of Male Infertility
2.1.5.1 Pharmacological Treatment of Male Infertility26
2.1.5.1.1 Hormonal Treatment

	2.1.5.1.2 Dopamine Agonists	27
	2.1.5.1.3 Aromatase Inhibitors	27
	2.1.5.1.4 Sympathomimetic Agents	27
	2.1.5.1.5 Selective Oestrogen Receptor Modulators	28
	2.1.5.2 Management of Oxidative Stress-Related Male	
	Infertility	28
	2.1.5.2.1 Lifestyle Modification	
	2.1.5.2.2 Antibiotics and Anti-Inflammatory Treatment of	
	Infection/Inflammation	29
	2.1.5.2.3 Vitamins and Antioxidants	
	2.1.5.3 Natural Products in Management of Male Infertility	
	2.1.5.4 Sperm Retrieval and Assisted Reproductive Technology	
	2.1.6 Hyperlipidaemia/ Hypercholesterolemia and Male Infertility	
	2.1.6.1 Overview of Hyperlipidaemia	
	2.1.6.2 Overview of Hypercholesterolemia	
	2.1.6.3 Effects of Hyperlipidaemia on Semen Quality, Testes	
	and Epididymides	36
	2.1.6.4 Hyperlipidaemia and Erectile Dysfunction	
	2.1.6.5 Effects of Hypercholesterolemia on Semen Quality,	
	Testes and Epididymides	39
	2.1.6.6 Hypercholesterolemia and Capacitation	
	2.1.6.7 Effects of Hyperlipidaemia/ Hypercholesterolemia on	10
	Male Reproductive Hormones	42
	2.1.6.8 High Energy Diet/ Hypercholesterolemia-Induced	12
	Oxidative Stress and Male Infertility	43
	2.1.6.9 Hypercholesterolemia and Liver X Receptors	
	2.1.6.10 Atorvastatin	
	2.1.7 Obesity and Male Infertility	
	2.1.7.1 Overview of Obesity	
	2.1.7.2 Effects of Obesity on Semen Quality and Sperm	
	Parameters	52
	2.1.7.3 Effects of Obesity on Male Reproductive Hormones	
	2.1.7.4 Obesity and Increased Scrotal Temperature	
	2.1.7.5 Obesity and Deoxyribonucleic Acid Fragmentation	
	2.1.7.6 Insulin Resistance and Male Infertility	
	2.1.7.7 Obesity Induced-Sleep Apnoea and Male Infertility	
	2.1.7.8 Obesity and Erectile Dysfunction	
	2.1.7.9 Management of Obesity-Induced Male Infertility	
	2.1.8 Animal Models of Male Reproductive Disorders and	00
	Infertility	60
	2.1.8.1 Rabbits as a Model of Hypercholesterolemia	
	2.1.8.2 Rabbits as a Model of Infertility	
2	Honey	
۷.۷	2.2.1 Definition and Composition of Honey	
	2.2.2 Stingless Bees versus Honey Bees	
	2.2.2 Striigless Bees Versus Holley Bees	
	2.2.4 Tualang Honey	
	2.2.5 Apis Mellifera Honey	
	2.2.5 Apis Meintera Honey	
	2.2.0 Timoney	09

2.2.7 Honey in Islamic Medicine	71
2.2.8 Honey as a Food	71
2.2.9 Antioxidant Activities of Honey	72
2.2.10 Anti-Inflammatory Properties of Honey	73
2.2.11 Honey and Infertility	
2.2.12 Medicinal Importance of Honey	
2.2.12.1 Antimicrobial Activities of Honey	
2.2.12.2 Wound Healing Properties of Honey	
2.2.12.3 Honey and Gastrointestinal Tract Diseases	80
2.2.12.4 Honey and Cough in Children	
2.2.12.5 Honey and Cardiovascular Diseases' Risk Factors	
2.2.12.6 Hepatoprotective Effects of Honey	
2.2.12.7 Renoprotective Effects of Honey	
2.2.12.8 Antineoplastic and Antiproliferative Effects of Honey	
2.2.12.9 Honey and Eye Diseases	
2.2.12.10 Neuroprotective Potential of Honey	
2.2.12.11 Honey and Bone	
CHAPTER THREE: METHODOLOGY	88
3.1 Materials	88
3.2 Sample Size Calculation	88
3.3 Ethical Approval	
3.4 Preparation of 1% Cholesterol Diet	
3.5 Honey Dosage and Adminstration	
3.6 Atorvastatin Dosage and Adminstration	
3.7 Animal Grouping	
3.8 Blood Collection and Serum Separation	96
3.9 Animal Sacrificing and Organ Harvesting	96
3.10 Animal Handling Procedure and Experimental Design	
3.11 Preparation of Testicular Tissue Homogenate	
3.12 Protein Assay in Testicular Homogenate	101
3.13 Statistical Analysis	101
CHAPTER FOUR: EFFECTS OF TRIHONEY ON SERUM LIPID PROFILE, SERUM AND TESTICULAR MALONDIALDEHYDE AND ANTIOXIDANT ENZYMES IN HIGH CHOLESTEROL DIET-FED	
MALE RABBITS	
4.1 Introduction	
4.2 Methodology	
4.2.1 Animal Weight and Daily Food Intake	
4.2.2 Blood Collection and Serum Separation	
4.2.3 Measurement of Lipid Profile Parameters	
4.2.4 Testicular Homogenate Preparation	104
4.2.5 Protocols of Elisa Analysis for Serum and Testicular	105
Homogenate4.2.5.1 Serum Malondialdehyde	
•	
4.2.5.2 Testicular Malondialdehyde	103
Homogenate	106
110.000.020.0000	

4.2.6.1 Superoxide Dismutase Activity Assay	106
4.2.6.2 Glutathione Peroxidase Assay	107
4.2.6.2.1 NADPH Standard Curve	108
4.2.6.2.2 Positive Control and Reagent Blank	108
4.2.6.2.3 Reaction Mix	
4.2.6.2.4 Calculation of Results	109
4.3 Results	109
4.3.1 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Animals' Weights, Weight Gain and Daily Food Intake	109
4.3.2 Baseline Lipid Profile	
4.3.3 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Lipid Profile After 12 Weeks	112
4.3.4 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Serum Malondialdehyde, Superoxide Dismutase and	
Glutathione Peroxidase	114
4.3.5 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Testicular Malondialdehyde, Superoxide Dismutase and	
Glutathione Peroxidase	116
4.4 Discussion	
4.5 Conclusion	
1.0 Concission	120
CHAPTER FIVE: EFFECTS OF TRIHONEY ON SERUM	
INFLAMMATORY BIOMARKERS IN HYPERCHOLESTEROLEMIC	
RABBITS	
5.1 Introduction.	
5.2 Methodology	
5.2.1 Blood Collection and Serum Separation	
5.2.2 Protocols of Elisa Analysis of Serum Pro-Inflammatory	150
Biomarkers	131
5.2.2.1 Interleukin-6 and Tumour Necrosis Factor-Alpha	
5.2.2.2 Interleukin-1β	
5.3 Results	
5.3.1 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Serum Levels of Pro-Inflammatory Cytokines	
5.4 Discussion	
5.5 Conclusion	
3.3 Coliciusion	139
CHAPTER SIX: EFFECTS OF TRIHONEY ON MALE	ı
CHAPTER SIX: EFFECTS OF TRIHONEY ON MALE REPRODUCTIVE HORMONES AND INSULIN RESISTANCE IN	
HYPERCHOLESTEROLEMIC RABBITS	
6.2 Methodology	
6.2.1 Blood Collection and Serum Separation	141
6.2.2 Measurement of Fasting Blood Glucose, Fasting Serum	1 / 1
Insulin and HOMA-IR	
6.2.3 Testicular Homogenate Preparation	
6.2.4 Protocols of Elisa Analysis	142
6.2.4.1 Serum Luteinizing Hormone and Follicle Stimulating	4 40
Hormone	142

6.2.4.2 Serum and Testicular Testosterone	143
6.3 Results	
6.3.1 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Serum and Intra-Testicular Testosterone	144
6.3.2 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Serum Follicle Stimulating Hormone and Luteinizing	
Hormone	145
6.3.3 Correlations of Serum Hormones with Serum Total	
Cholesterol and Low Density Lipoprotein Cholesterol	147
6.3.4 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Fasting Blood Glucose, Fasting Serum Insulin and HOMA-IR	148
6.4 Discussion	
6.5 Conclusion	155
CHAPTER SEVEN: EFFECTS OF TRIHONEY ON SPERM	I
PARAMETERS IN HYPERCHOLESTEROLEMIC MALE RABBITS	156
7.1 Introduction	156
7.2 Methodology	157
7.2.1 Caudal Epididymal Sperm Analysis	157
7.2.1.1 Analysis of Sperm Motility	159
7.2.1.2 Analysis of Sperm Vitality and Morphology	160
7.2.1.3 Analysis of Sperm Concentration	161
7.3 Results	162
7.3.1 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Sperm Motility	162
7.3.2 Correlation of Sperm Motility with Serum Total Cholesterol	
and Low-Density Lipoprotein Cholesterol	164
7.3.3 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Sperm Vitality	165
7.3.4 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Sperm Morphology	166
7.3.5 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Sperm Concentration	167
7.3.6 Correlations of Sperm Parameters with Serum Total	
Cholesterol and Low-Density Lipoprotein Cholesterol	170
7.3.7 Correlations Between Serum Hormones and Sperm	
Parameters	
7.4 Discussion	
7.5 Conclusion	181
CHAPTER EIGHT: EFFECTS OF TRIHONEY ON TESTICULAR AND	
EPIDIDYMAL HISTOPATHOLOGICAL ALTERATIONS IN	•
HYPERCHOLESTEROLEMIC MALE RABBITS	
8.1 Introduction	
8.2 Methodology	
8.2.1 Relative Organs Weight	
8.2.2 Tissue Histology	
8.2.2.1 Tissue Fixation	
8.2.2.2 Tissue Grossing	185

8.2.2.3 Tissue Processing	185
8.2.2.4 Tissue Embedding	
8.2.2.5 Trimming and Sectioning	
8.2.2.6 Haematoxylin and Eosin Staining	
8.2.2.6.1 Deparaffinisation	
8.2.2.6.2 Hydration	
8.2.2.6.3 Haematoxylin and Eosin Stain	
8.2.2.6.4 Dehydration	
8.2.2.6.5 Mounting	
8.2.2.6.6 Slide Inspection and Picture Caption	
8.2.2.7 Masson's Trichrome Staining	
8.2.2.7.1 Deparaffinisation	
8.2.2.7.2 Hydration	
8.2.2.7.3 Masson's Trichrome Stain	
8.2.2.7.4 Dehydration	
8.2.2.7.5 Mounting	
8.2.2.7.6 Slide Inspection and Picture Caption	190
8.2.2.8 Histological Measurements	190
8.2.2.9 Spermatogenesis Evaluation	190
8.3 Results	
8.3.1 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Testicular and Epididymal Weights	191
8.3.2 Correlations of Testicular and Epididymal Weights with	
Serum Total Cholesterol and Low-Density Lipoprotein	
Cholesterol	194
8.3.3 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Testicular and Epididymal Gross Morphology	196
8.3.4 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	170
Testicular Histology	197
8.3.4.1 Whole Testicular Sections from all Groups	
8.3.4.2 Control Group	199
8.3.4.3 High-cholesterol Diet Group	
8.3.4.4 Atorvastatin Treated Group	
8.3.4.5 Trihoney (0.3 g/kg/day) Received Group	
8.3.4.6 Trihoney (0.6 g/kg/day) Received Group	209
8.3.4.7 Effects of 1% Cholesterol Diet, Trihoney and	
Atorvastatin on the Diameter of Seminiferous Tubules,	211
Thickness of Tunica Albuginea and Johnsen's Score	211
8.3.5 Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on	
Epididymal Histology	
8.3.5.1 Control Group	
8.3.5.2 High Cholesterol Diet Group	
8.3.5.3 Atorvastatin Treated Group	
8.3.5.4 Trihoney (0.3 g/kg/day) Received Group	
8.3.5.5 Trihoney (0.6 g/kg/day) Received Group	225
8.3.5.6 Effects of 1% Cholesterol Diet, Trihoney and	
Atorvastatin on Epithelial Height and Diameter of Ductal	
Lumens of Caput and Cauda of the Epididymis	228
8.4 Discussion.	231

8.5 Con	clusion			239
			CONCLUSION	
			•••••	
9.1 Gen	eral Conclusion	on		240
9.2 Rec	ommendation			247
			••••••	
			••••••	
			INARS	
APPENDIX E	:: ACADEMI	C SCHOLARSH	IPS	305

LIST OF TABLES

Table No.		Page No
2.1	Classification and Major Causes of Male Infertility	15
2.2	The Lower Reference Limit of Semen Parameters Based on WHO Laboratory Manual for the Examination and Processing of Human Semen	23
2.3	Composition of Trihoney	70
3.1	Composition of Coconut Oil	90
3.2	Animal Grouping of The Current Experiment	94
3.3	Nutritional Composition of Rabbit Pellet (Percentage/25kg)	95
4.1	Preparation of Samples and Blank for SOD Activity Measurement	107
4.2	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Animals' Weights, Weight Gain and Daily Food Intake	111
4.3	Baseline Lipid Profile	112
4.4	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Lipid Profiles After 12 Weeks	114
4.5	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Serum Malondialdehyde, Superoxide Dismutase and Glutathione Peroxidase	116
4.6	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Testicular Malondialdehyde, Superoxide Dismutase and Glutathione Peroxidase	118
5.1	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Serum Pro-Inflammatory Cytokines	134
6.1	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Serum and Intra-Testicular Testosterone	145
6.2	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Serum Follicle Stimulating Hormone and Luteinizing Hormone	147

6.3	Cholesterol and Low-Density Lipoprotein Cholesterol	
6.4	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Fasting Blood Glucose, Fasting Insulin and HOMA-IR	149
7.1	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on the Percentages of Sperm Motility	164
7.2	Correlation Coefficients Between Serum Lipid Profile and Sperm Motility	165
7.3	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Percentages of Sperm Vitality, Sperm Morphology and Sperm Concentration	168
7.4	Correlation Coefficients Between Serum Lipid Profiles and Sperm Parameters	171
7.5	Correlations Between Serum Hormones and Sperm Parameters	172
8.1	Modified Johnsen's Score	191
8.2	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Testicular and Epididymal Weights	193
8.3	Correlations of Serum Total Cholesterol and Low-Density Lipoprotein Cholesterol with Testicular and Epididymal Weights	195
8.4	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Seminiferous Tubules Diameter, Tunica Albuginea thickness and Johnsen's Score	212
8.5	Effects of 1% Cholesterol Diet, Trihoney and Atorvastatin on Epithelial Height and Diameter of Ductal Lumen of Caput and Cauda of the Epididymis	230

LIST OF FIGURES

Figure No.		Page No.
2.1	Anatomy of Testis Depicting the Site of Spermatogenesis.	12
2.2	Different Origins of Sperm Oxidative Stress	16
2.3	Management of Obesity-Induced Male Infertility	
2.4	Overview of Male Reproductive System of Rabbits	63
2.5	Honey Comb (A) of Sting Bee Honey And Pot (B) of Stingless Bee Honey	67
2.6	Trihoney	70
2.7	Medicinal Importance of Honey	87
3.1	Preparation of 1% Cholesterol Diet	91
3.2	Animal Handling Procedure and Experimental Design	98
3.3	Preparation of Testicular Tissue Homogenate	100
7.1	The Procedure of Sperm Analysis	159
7.2	Eosin-Nigrosin Stain of Sperm Morphology and Vitality	169
8.1	Testicular and Epididymal Gross Morphology	197
8.2	Pictomicrograph of Testicular Whole Sections	198
8.3	Pictomicrograph of Testicular Sections from Control Group	200
8.4	Pictomicrograph of Testicular Sections from Commercial Pellet with Trihoney Group	201
8.5	Pictomicrograph of Testicular Sections from High- Cholesterol Diet Group	203
8.6	Pictomicrograph of Testicular Sections from High- Cholesterol Diet Group	204
8.7	Pictomicrograph of Testicular Sections from Atorvastatin Treated Group	206
8.8	Pictomicrograph of Testicular Sections from Trihoney (0.3 g/kg/day) Received Group	208

8.9	(0.6 g/kg/day) Received Group	210
8.10	Pictomicrograph of Sections of Caput Epididymis from Control Group	213
8.11	Pictomicrograph of Sections of Cauda Epididymis from Control Group	214
8.12	Pictomicrograph of Sections of Caput Epididymis from Commercial Pellet with Trihoney Group	215
8.13	Pictomicrograph of Sections of Cauda Epididymis from Commercial Pellet with Trihoney Group	216
8.14	Pictomicrograph of Sections of Caput Epididymis from High Cholesterol Diet Group	218
8.15	Pictomicrograph of Sections of Cauda Epididymis from High Cholesterol Diet Group	219
8.16	Pictomicrograph of Sections of Caput Epididymis from Atorvastatin Treated Group	221
8.17	Pictomicrograph of Sections of Cauda Epididymis from Atorvastatin Treated Group	222
8.18	Pictomicrograph of Sections of Caput Epididymis from Trihoney (0.3 g/kg/day) Received Group	224
8.19	Pictomicrograph of Sections of Cauda Epididymis from Trihoney (0.3 g/kg/day) Received Group	225
8.20	Pictomicrograph of Sections of Caput Epididymis from Trihoney (0.6 g/kg/day) Received Group	226
8.21	Pictomicrograph of Sections of Cauda Epididymis from Trihoney (0.6 g/kg/day) Received Group	227

LIST OF ABBREVIATIONS

ABCA1 ATP-binding transporters A1
ART Assisted reproductive technology

BMI Body mass index

ATP Adenosine triphosphate

cAMP Cyclic adenosine monophosphate

CA-MRSA Community-associated methicillin-resistant *Staphylococcus aureus*

CoQ10 Coenzyme Q (10)

COX2 Cyclooxygenase enzyme CRP C-Reactive protein

DFI DNA fragmentation index

dH₂O Deionized waster
DNA Deoxyribonucleic acid
Fas Fatty acid synthase

FSH Follicle -stimulating hormone

g Gram

GnRH Gonadotropin releasing hormone

GPx Glutathione peroxidase

GSH Glutathione

GR Glutathione reductase
GSSG Oxidized glutathione
HCD High cholesterol diet

hCG Human chorionic gonadotropin HDL-c High density lipoprotein cholesterol

H&E Haematoxylin and Eosin

HED High-energy diet

hMG Human menopausal gonadotropin

HOMA-IR Homeostatic model assessment of insulin resistance

HPT-axis Hypothalamic pituitary testicular axis

HRP Horseradish peroxidase HTF Human tubal fluid

ICSI Intracytoplasmic sperm injection

IIUM International Islamic University Malaysia

IL Interleukin

IVF In vitro fertilization

kg Kilogram

LAC L-acetyl-carnitine

LC L-carnitine

LDH Lactate dehydrogenase LH Luteinizing hormone LXRs Liver X receptors

M Mean

MDA Malondialdehyde

mg Milligram

MM6 Monocytic cell line and precursor of macrophages

mL Millilitre

 $\begin{array}{ccc} \mu L & Microliter \\ mM & Millimole \\ \mu M & Micromole \\ \mu m & Micrometre \end{array}$

mmol/L Millimole per litre MT Masson's Trichrome

NADP+ Nicotinamide adenine dinucleotide phosphate

NADPH Nicotinamide adenine dinucleotide phosphate hydrogen

NF-kB Nuclear translocation of nuclear factor kappa B

ng/mL Nano gram per millilitre

NO Nitric oxide

NOI Non-obstructive Infertility

NSAID Nonsteroidal anti-inflammatory drugs

OI Obstructive Infertility
PBS Phosphate buffer saline
PBUH Peace Be Upon Him
PC Protein carbonyl
PG Prostaglandin

Pg/mL Pictogram per millilitre

PKA Protein kinase A
PM Progressive motility
RC Reagent control

r-hFSH Recombinant human FSH

rHuIL-6 Recombinant human interleukin-6

R/N Reference number RNA Ribonucleic acid

RNS Reactive nitrogen species
ROS Reactive oxygen species
rpm Revolution per minute

RSM Response Surface Methodology

SC Segar smoke

scd Stearoyl Co-A desaturases

SD Standard deviation SDH Sorbitol dehydrogenase

SERMs Selective oestrogen receptor modulators

SMC Smooth muscle cells

S/N Serial number

SOD Superoxide dismutase

srebp1c Sterol response element binding protein-1c

STZ Streptozotcin

TBARS Thiobarbituric acid reactive substances

TC Total cholesterol
TG Triglycerides
TM Total motility

TMB Tetramethylbenzidine

TNF-α Tumour necrosis factor- alpha

U/L Activity unit per litre

US\$ Dollars

VLDL Very low density lipoprotein

World Health Organization Xanthine oxidase WHO

XOD

LIST OF SYMBOLS

-	Hyphen-minus
+	Plus sign
=	Equal sign
%	Percent sign
&	Ampersand
(Left parenthesis
)	Right parenthesis
,	Comma
	Full stop
/	Solidus
:	Colon
;	Semicolon
[Left square bracket
]	Right square bracket
<	Less-than sign
>	Greater-than sign
±	Plus-minus sign
0	Degree sign

CHAPTER ONE

INTRODUCTION

1.1 BACKGROUND OF THE STUDY

Infertility is defined as the failure to conceive in sexually active, noncontracepting couples for a period of one year or more (Yilmaz et al., 2017). It is a common problem affecting 15% of couples of childbearing age, with detectable male factor in 30-50% of all infertile couples (Eisenberg et al., 2014; Oyeyipo et al., 2015). In 20% of couples, male factor is the only causative actiology for infertility (Attaman et al., 2012). The prevalence of male infertility is on the rise globally and is of public concern owing to its socioeconomic burden (Michael et al., 2015). Due to environmental contamination and life style changes, infertility rate is going to increase in the future (Pushpendra & Jain, 2015). Lifestyle-related external factors including eating disorders can negatively affect spermatogenesis both at central and gonadal levels (Al Kushi et al., 2016). Poor dietary habits with high-fat or high-cholesterol intake are the main cause towards the development of hyperlipidaemia and hypercholesterolemia which are increasing in young people in both developed and developing nation (Aurelia Ouvrier et al., 2011; Onwe et al., 2015). Dyslipidaemia is a major risk factor for the development of cardiovascular complications. Its deleterious effects extend to affect the reproductive functions (Aurelia Ouvrier et al., 2011). The negative impact of hypercholesterolemia on male reproductive system and fertility has been reported in animal (Saez Lancellotti et al., 2010) and human (Schisterman et al., 2014). Hypercholesterolemia affects testicular structure and function, spermatogenesis, semen quality and ejaculatory function through disruption of hypothalamic-pituitary-testicular (HPT) axis, impairment of steroid hormone

biosynthesis, impairment of Sertoli and Leydig cells secretory functions, induction of oxidative stress and disruption of various testicular genes (Pushpendra & Jain, 2015). Furthermore, hypercholesterolemia affects structure and function of the epididymides (Aurelia Ouvrier et al., 2011).

Complementary and alternative medicine is widely used and rapidly growing in developing and developed countries. It is used by 80% of African population. In China, traditional medicine constitutes 40% of health care system delivered. In Malaysia, US\$500 million is spent annually for this kind of care. Complementary and alternative medicine is used by 70% and 42% of population in Canada and United States respectively. The wide use of traditional medicine is attributed in developing countries to its affordability and accessibility, in Asia due to historical and cultural believes; whereas, in developed countries the main cause of increasing use of complementary and alternative medicine is the concern about the side effects of conventional medicine (WHO, 2002).

Honey is an important and unique natural product (Ramanauskiene et al., 2012). It has been used since ancient times as a therapeutic agent (Pyrzynska & Biesaga, 2009). Recently, the attention has been increased towards the use of honey for prevention and treatment of numerous diseases as well as for improving and maintaining the overall wellbeing (Inoue et al., 2005; Pyrzynska & Biesaga, 2009; Nweze et al., 2016). The medicinal importance of honey has been demonstrated in several previous studies. It has been reported to have antioxidant activity (Alvarez-Suarez et al., 2010), anti-inflammatory activity (Borsato et al., 2014) and Antihyperlipidaemic effect (Yaghoobi et al., 2008; Adnan, Sadiq & Jehangir, 2011). Traditionally, honey has been used in different cultures for enhancement of male fertility (Abdul-Ghani et al., 2008; Mohamed et al., 2012). It showed its ability to