## ANTICANCER ACTIVITY OF IONIC LIQUID GRAVIOLA FRUIT (Annona muricata) EXTRACT ON MCF-7 AND HT29 CANCER CELL LINES USING IN VITRO AND IN VIVO METHODS

BY

## DADDIOUAISSA DJABIR

A thesis submitted in fulfilment of the requirement for the degree of Doctor of Philosophy (Engineering)

Kulliyyah of Engineering International Islamic University Malaysia

**APRIL 2020** 

#### **ABSTRACT**

Cancer, one of the major public health problems, is the leading cause of death worldwide. The available protocols of treatment include surgical intervention, radiation, and chemotherapy which cause numerous side effects on cancer patients. Many phytochemicals have anticancer properties comparable to conventional drugs. The major benefit of these compounds is the non-toxicity nature to the normal tissues. Annona muricate, commonly known as Graviola, is a member of the Annonaceae family. It has been used for ages in traditional medicine due to its biological activities including antioxidant, anti-inflammatory, antimicrobial and cytotoxicity to tumour cells. This research investigated the antiproliferative effect of the ionic liquid-Graviola fruit extract (IL-GFE) on in vitro breast MCF-7 and colon HT29 adenocarcinoma cell lines and their cytokinetic behaviour. It also identified the mechanism of IL-GFE inhibition by applying a flow cytometry technique and metabolomics study and assessed its toxicity on in vivo zebrafish developing embryos. The process of ionic liquidmicrowave assisted extraction (IL-MAE) method was optimised by Response Surface Methodology for three parameters, namely time, irradiation power and solid-liquid ratio. The optimum extraction conditions gave a yield of Graviola fruit extract up to 66.6 % and an average IC<sub>50</sub> of 4.75 μg/mL for MCF-7 and 10.56 μg/mL for HT29, while it was safe toward normal VERO cell lines. The crude IL-GFE was fractionated using the combination of thin-layer chromatography and column chromatography. Six fractions were semi-purified and subjected to phytochemical screening and antiproliferative assay in which, it revealed the presence of many phytoconstituents such as acetogenins, alkaloids, phenols, flavonoids, tannins and terpenoids. Moreover, fraction B exhibited the lowest IC<sub>50</sub> toward MCF-7 and HT29 cells at 12.6 and 13.56 μg/mL, respectively. However, the crude IL-GFE had a better IC<sub>50</sub> value. The crude IL-GFE with GC-TOFMS analysis revealed the presence of many phytochemicals with anticancer activity such as D-psicofuranose, pentakis ether, propyldecyl cyclopropane dodecacarbonyl, N-acetylimino dimethylsulfurane, tri-ruthenium pyranone, carbohydrazide and benzoic acid. The cytokinetic study showed that crude IL-GFE and Taxol inhibited the growth of MCF-7 and HT29 cells and proved their antiproliferative effect when they reduced the number of cell generations of MCF-7 from 3.71 to 1.67 and 2.18, respectively, and reduced the cell generations of HT29 cells from 3.93 to 2.96 and 2.01, respectively. Furthermore, the acute toxicity of IL-GFE was assessed on in vivo zebrafish model in which crude IL-GFE reduced the survival of zebrafish larvae at a relatively high dose of 250 μg/mL after 96 hpf treatment, while no significant changes on morphology of the treated zebrafish were recorded. The result of the flow cytometry also indicated that the crude IL-GFE arrested the cell cycle of MCF-7 and HT29 at G0/G1 phase and increased the apoptotic and necrotic cells in a timedependent manner compared with the control group. Finally, the metabolomics analysis of the treated MCF-7 and HT29 cells with crude IL-GFE treatment showed an alteration of many metabolic pathways in treated cancer cells. In conclusion, crude IL-GFE can be one of the promising anticancer agents due to its selective antiproliferation against breast and colon cancer cells and its safety for the healthy cells.

## خلاصة البحث

يعد مرض السرطان من بين المشكلات الأساسية للصحة العامة وهو السبب الرئيسي للوفاة في العالم. تشمل البروتوكولات العلاجية المتاحة التدخل الجراحي والإشعاعي والعلاج الكيميائي الذي يسبب آثارًا جانبية عديدة لدى مرضى السرطان. العديد من المواد الكيميائية النباتية لها خصائص مضادة للسرطان منافسة للأدوية التقليدية. الفائدة الرئيسية لهذه المركبات تكمن في طبيعتها غير السمية للأنسجة السليمة. Annona muricata المعروفة باسم الجرافيولا، نبتة من عائلة الأنوناسي، تم استخدامها منذ القديم في الطب التقليدي بسبب أنشطتها البيولوجية بما في ذلك النشاط المضاد للأكسدة والالتهابات والميكروبات، إضافة الى السمية الخلوية للخلايا السرطانية. يهدف هذا البحث إلى دراسة التأثير المضاد للسرطان لمستخلص السائل الأيوني لفاكهة الجرافيولا على الخلايا السرطانية للثدي MCF-7 والخلايا السرطانية للقولون HT29 وسلوكهما الخلوي، إضافة الى تحديد آلية عمل مستخلص فاكهة الجرافيولا من خلال تطبيق تقنية التدفق الخلوي ودراسة الأيض وتقييم سميته في أجنة سمكة الزرد. أولا، تم تحسين طريقة الاستخلاص بالسائل الأيوني بمساعدة الميكروويف من خلال منهجية الاستجابة السطحية لثلاثة عوامل: الوقت (دقيقة)، قدرة التشعيع (واط) ونسبة المادة الصلبة بالنسبة للسائل (غرام/مل). أعطت ظروف الاستخلاص المثلي مستخلص الفاكهة بنسبة تصل إلى 66.6٪ ومتوسط تثبيط قدره 4.75 ميكروغرام/مل بالنسبة لخلايا سرطان الثدي و10.56 ميكروغرام/مل بالنسبة لخلايا سرطان القولون، في حين كانت آمنة تجاه الخلايا VERO السليمة. بعد ذلك، تم تنقية مستخلص فاكهة الجرافيولا الخام باستخدام كل من تقنية كروماتوجرافيا الطبقة الرقيقة وكروماتوجرافيا العمود. تم اصطفاء ستة أجزاء نشطة وتعريضها للفحص الكيميائي النباتي والنشاط المضاد للسرطان، حيث كشفت الدراسة عن وجود العديد من المركبات النباتية مثل الأسيتوجينات، القلويدات، الفينولات والفلافونويدات والتانينس والتيربينويدات. علاوة على ذلك، أظهر الجزء النشط باء أدبي ألكاري بالنسبة لخلايا سرطان الثدي والقولون عند 12.6 و13.56 ميكروغرام/مل، على التوالي، مقارنة بالأجزاء النشطة الأخرى. لكن يبقى مستخلص فاكهة الجرافيولا الخام هو الأكثر تثبيطا لخلايا سرطان الثدي والقولون مقارنة بالمركبات الفعالة الأخرى. بالإضافة الى ذلك، أظهر تحليل مستخلص فاكهة الجرافيولا الخام باستخدام اله GC-TOFMS وجود العديد من المواد الكيميائية النباتية ذات النشاط المضاد للسرطان. أظهرت دراسة الحركية الخلوية أن مستخلص فاكهة الجرافيولا الخام والتاكسول حالت دون نمو الخلايا السرطانية للثدي والقولون وأثبتت تأثيرها المضاد للسرطان عندما خفضت عدد الأجيال لخلايا سرطان الثدي من 3.71 إلى 1.67 و2.18 على التوالي. بينما، قللت عدد أجيال الخلايا السرطانية للقولون من 3.93 إلى 2.96 و 2.01، على التوالى. بعد ذلك، تم تقييم سمية مستخلص فاكهة الجرافيولا الخام على أجنة سمكة الزرد النامية، حيث تسبب المستخلص في موت يرقات سمكة الزرد بعد معالجتها بجرعة عالية نسبيًا قدرها 250 ميكروغرام/مل، بعد 96 ساعة من العلاج، في حين لم تحدث تغييرات كبيرة على شكل يرقات الزرد المعالجة. أشارت نتيجة قياس التدفق الخلوي إلى أن مستخلص فاكهة الجرافيولا الخام أوقف دورة الخلية للخلايا السرطانية الثديية والقولونية في مرحلة GO/G1 وزاد من الخلايا الميتة بسبب الموت الخلوي المبرمج والنخرية. أخيرًا، فيما يخص تحليل اللأيض للخلايا، فقد أظهر تغييرا واضحا في أيض الخلايا السرطانية المعالجة بالمستخلص. في الختام، يمكن أن يكون مستخلص فاكهة الجرافيولا الخام واحدًا من العوامل الواعدة المضادة للسرطان، نظرًا لنشاطه الانتقائي ضد الخلايا السرطانية للثدي والقولون وعدم تأثيره على الخلايا السليمة.

## APPROVAL PAGE

The thesis of Dao	ddiouaissa Djabir has been approved	by the following
	Azura Amid	_
	Supervisor	
	Nassereldeen Ahmed Kabbashi	-
	Co-Supervisor	
	Fazia Adyani Ahmad Fuad	_
	Co-Supervisor	
	Hamzah Mohd Salleh	-
	Internal Examiner	
	Noriham Abdullah	-
	External Examiner	
	Fadzilah Adibah Abdul Majid	-
	External Examiner	
		-
	Mohammad Naqib Ishan Jan Chairman	

## **DECLARATION**

I hereby declare that this thesis is the result of my own investigations, except where

otherwise stated. I also declare that it has not been p	reviously or concurrently submitted
as a whole for any other degrees at IIUM or other in	nstitutions.
Daddiouaissa Djabir	
Signature	Date

#### INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

## DECLARATION OF COPYRIGHT AND AFFIRMATION OF FAIR USE OF UNPUBLISHED RESEARCH

# ANTICANCER ACTIVITY OF IONIC LIQUID GRAVIOLA FRUIT (Annona muricata) EXTRACT ON CANCER CELL LINES USING IN VITRO AND IN VIVO METHODS

I declare that the copyright holders of this thesis are jointly owned by the student and IIUM.

Copyright © 2020 Daddiouaissa Djabir and International Islamic University Malaysia. All rights reserved.

No part of this unpublished research may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder except as provided below

- 1. Any material contained in or derived from this unpublished research may be used by others in their writing with due acknowledgement.
- 2. IIUM or its library will have the right to make and transmit copies (print or electronic) for institutional and academic purposes.
- The IIUM library will have the right to make, store in a retrieved system
  and supply copies of this unpublished research if requested by other
  universities and research libraries.

By signing this form, I acknowledged that I have read and understand the IIUM Intellectual Property Right and Commercialization policy.

Affirmed by Daddiouaissa Djabir	
Signature	 Date

#### **ACKNOWLEDGEMENTS**

In the Name of Allah, the Most Gracious, Most Merciful and Salam upon His Messenger, Prophet Mohammed (SAW).

All praise and thanks are due to my creator and sustainer, for guiding me to conceptualise, develop and accomplish this research work.

My sincere appreciation goes to my dear parents, who granted me the gift of their unwavering belief in my ability to accomplish this goal and my wife for her support and patience.

I would like to express the immense appreciation to my supervisors- Prof. Dr. Azura Amid, Prof. Dr. Nassereldeen Ahmed Kabbashi and Asst. Prof. Dr. Fazia Adyani Ahmad Fuad for their guidance, advice, patience, encouragement and valuable assistance that enabled me to accomplish this work smoothly and efficiently.

I wish to extend my warmest thanks to Dr. Shirwan for his time and guidance and all lab mates, Br. Oualid, Br. Ahmed, Br. Phirdaous, Br. Deni and my friends Hassen and Slimane as well as to Mr. Said Ammisaid for his proof reading and all those who have helped me go through this work.

Finally, I acknowledge the valuable inputs of the Department of Biotechnology Engineering and all lecturers and technical staff who helped me in accomplishing this study.

## **TABLE OF CONTENTS**

Abstract	ii
Abstract in Arabic	iii
Approval Page	iv
Declaration	V
Copyright	vi
Acknowledgements	vii
List of Tables	xii
List of Figures	xiv
List of Abbreviations	xviii
List of Symbols	xix
CHAPTER ONE: INTRODUCTION	1
1.1 Background of the Study	
1.2 Problem Statement and Significance of the Study	
1.3 Research Hypothesis	
1.4 Research Objectives	
1.5 Research Methodology	
1.6 Research Scope	
1.7 Thesis Organisation	
CHAPTER TWO: LITERATURE REVIEW	10
2.1 Introduction.	
2.2 GRAVIOLA (Annona Muricata L)	
2.2.1 Botanical Description and Distribution	
2.2.2 Ethnomedicinal Use	
2.2.3 Phytochemicals	
2.2.3.1 Annonaceous Acetogenins	
2.2.4 Anticancer Activity of Graviola Fruit	
2.3 Cancer Disease	
2.3.1 Classification of Cancer	
2.3.1.1 Carcinomas	
2.3.1.2 Sarcoma	
2.3.1.3 Leukaemia	
2.3.1.4 Lymphoma	
2.3.2 Cancer Statistics	
2.3.3 Cancer Staging Systems	
2.3.4 Breast Cancer	
2.3.5 Colon Cancer	
2.3.6 Conventional Cancer Treatments	
2.3.6.1 Radiation Therapy	
2.3.6.2 Surgical Intervention	
2.3.6.3 Chemotherapy	
2.3.7 Alternative Cancer Treatment from Plants	
2.4 Apoptosis and Necrosis	

2.5 Cell Cycle	30
2.6 Extraction of Anti-Cancer Compounds	31
2.6.1 Extraction Methods	32
2.6.1.1 Ionic Liquid Extraction	33
2.6.1.2 Ionic Liquid-Based Microwave-Assisted Extractions	34
2.7 Separation and Purification	
2.7.1 Thin Layer Chromatography	36
2.7.2 Column Chromatography	
2.8 <i>In Vitro</i> Mammalian Cell Culture Assay	
2.8.1 MCF-7 Breast Cancer Cell Line	
2.8.2 HT29 Colon Cancer Cell Line	
2.8.3 VERO Normal Kidney Cell Line	
2.9 Antiproliferative Assay (MTT Assay)	
2.10 Cytokinetics Study	
2.11 <i>In Vivo</i> Acute Toxicity Study on Zebrafish	
2.12 Metabolomics Study	
2.13 Research gap	
2.14 Summary	
2.14 Summary	40
CHAPTER THREE: MATERIALS AND METHODS	47
3.1 Introduction	
3.2 Materials and Equipment	
3.2.1 Raw Material	
3.2.2 Cell Lines and Zebrafish Embryos	
3.2.3 Chemicals and Reagents	
3.2.4 Consumable Items	
3.2.5 Equipment and Apparatus	
3.3 Methodology	
3.3.1 Preparation of Solutions and Reagents	
3.3.1.1 Preparation of Cell Culture Medium	
3.3.1.2 Preparation of Medium with Fetal Bovine Serum (FBS	/
3.3.1.3 Preparation of Phosphate Buffered Saline (PBS)	
3.3.1.4 Preparation MTT Assay Solution (5 mg/mL) in PBS	
3.3.1.5 Preparation of the Ionic Liquid Solutions	
3.3.2 Graviola Fruit Preparation	
3.3.3 Graviola Fruit Extraction	
3.3.4 Statistical Optimisation Experiments	
3.3.4.1 One-Factor-at-a-Time (OFAT) Experimental Design	
3.3.4.2 Optimisation of Extraction Conditions	
3.3.4.3 Validation of the Experimental Model	
3.3.5 Separation and Semi Purification of the Bioactive Fractions	
3.3.5.1 Thin Layer Chromatography (TLC)	
3.3.5.2 Open Column Chromatography	
3.3.6 Phytochemical Screening	
3.3.6.1 Detection of Acetogenins	
3.3.6.2 Detection of Alkaloids	
3.3.6.3 Detection of Phenols	
3.3.6.4 Detection of Flavonoids	
3.3.6.5 Detection of Saponins	61

3.3.6.6 Detection of Terpenoids	62
3.3.6.7 Detection of Tannins	62
3.3.7 Cell Culture	62
3.3.7.1 Thawing Frozen Cells	63
3.3.7.2 Cell Revival	
3.3.7.3 Cell Subculture	63
3.3.7.4 Cell Counting	64
3.3.7.5 Cryopreservation of Cells	65
3.3.8 Cell Viability (MTT Assay)	
3.3.8.1 Estimation of the Inhibitory Concentration at 50% (IC <sub>50</sub> )	
3.3.9 Cytokinetics Study	
3.3.10 Acute Toxicity of IL-GFE on Zebrafish Embryos/Larval	
3.3.10.1 Zebrafish Maintenance and Embryo Collection	
3.3.10.2 Zebrafish Embryos/Larvae Exposure	
3.3.11 Flow Cytometry for Cell Cycle Analysis and Detection of	
Apoptosis	69
3.3.11.1 Cell Cycle Phase Analysis	
3.3.11.2 Apoptosis Analysis	
3.3.12 Metabolomic Alterations in IL-GFE Treated Cancer Cells	
3.3.12.1 Sample Preparation and Quenching of Cell Metabolic	
3.3.12.2 Metabolite Extraction	
3.3.12.3 GC-TOFMS	
3.3.12.4 Multivariate Data Analysis	
3.3.13 Statistical Analysis	
3.4 Summary	
CHAPTER FOUR: RESULTS AND DISCUSSION	
4.1 Introduction	75
4.1 Introduction	75
<ul><li>4.1 Introduction</li></ul>	75 75 76
<ul> <li>4.1 Introduction</li></ul>	75 75 76 79
4.1 Introduction	75 76 79 81
4.1 Introduction	75 76 79 81 82
<ul> <li>4.1 Introduction</li></ul>	75 76 79 81 82
<ul> <li>4.1 Introduction</li></ul>	75 76 79 81 82 87
4.1 Introduction	75 76 79 81 82 87 94
4.1 Introduction	75 76 79 81 82 87 94 95
4.1 Introduction	75 76 79 81 82 87 94 95 95
4.1 Introduction 4.2 Extraction of Graviola Fruit Extract 4.3 Optimisation of ionic liquid solutions by OFAT 4.4 OptimiSation of IL-MAE Parameters by RSM 4.4.1 Analysis of Variance 4.4.1.1 Analysis of Variance (ANOVA) of the Extraction Yield. 4.4.1.2 Analysis of Variance (ANOVA) of the IC50 4.5 Partial Fractionation and Separation 4.5.1 Thin Layer Chromatography (TLC) 4.5.2 Column Chromatography 4.5.3 Phytochemical Screening 4.5.4 Discussion	757679818287949595
<ul> <li>4.1 Introduction</li></ul>	75767981828794959798
4.1 Introduction 4.2 Extraction of Graviola Fruit Extract 4.3 Optimisation of ionic liquid solutions by OFAT 4.4 OptimiSation of IL-MAE Parameters by RSM 4.4.1 Analysis of Variance 4.4.1.1 Analysis of Variance (ANOVA) of the Extraction Yield. 4.4.1.2 Analysis of Variance (ANOVA) of the IC <sub>50</sub> 4.5 Partial Fractionation and Separation 4.5.1 Thin Layer Chromatography (TLC) 4.5.2 Column Chromatography 4.5.3 Phytochemical Screening 4.5.4 Discussion 4.6 Qualitative phytoconstituent Identification of IL-GFE 4.6.1 GC-TOFMS Analysis	7576798182879495979899
<ul> <li>4.1 Introduction</li></ul>	757576798182879495979899102102
4.1 Introduction 4.2 Extraction of Graviola Fruit Extract 4.3 Optimisation of ionic liquid solutions by OFAT 4.4 OptimiSation of IL-MAE Parameters by RSM 4.4.1 Analysis of Variance 4.4.1.1 Analysis of Variance (ANOVA) of the Extraction Yield. 4.4.1.2 Analysis of Variance (ANOVA) of the IC <sub>50</sub> 4.5 Partial Fractionation and Separation 4.5.1 Thin Layer Chromatography (TLC) 4.5.2 Column Chromatography 4.5.3 Phytochemical Screening 4.5.4 Discussion 4.6 Qualitative phytoconstituent Identification of IL-GFE 4.6.1 GC-TOFMS Analysis 4.6.2 Discussion 4.7 Antiproliferative Activities	75767981879495979899102105
<ul> <li>4.1 Introduction</li> <li>4.2 Extraction of Graviola Fruit Extract</li> <li>4.3 Optimisation of ionic liquid solutions by OFAT</li> <li>4.4 OptimiSation of IL-MAE Parameters by RSM</li></ul>	75767981879495979899102105
<ul> <li>4.1 Introduction</li></ul>	757679818294959891102102106
4.1 Introduction 4.2 Extraction of Graviola Fruit Extract	7575767981829495979899102105106
4.1 Introduction	7575767981879495979899102106106
4.1 Introduction  4.2 Extraction of Graviola Fruit Extract	75757679818294959798102105106106

4.8.2 Effect of Crude IL-GFE on HT29 Growth Kinetics	117
4.8.3 Discussion	120
4.9 In Vivo ZEBRAFISH Embryo/Larvae Toxicity Assay	123
4.9.1 Effects of Crude IL-GFE on the Survival Rate of Zebrafish	123
4.9.2 Effects of Crude IL-GFE on the Morphology of Zebrafish	124
4.9.3 Effects of Crude IL-GFE on Zebrafish Heart Rate	127
4.9.4 Discussion	128
4.10 Cell Cycle Distribution and Apoptosis	131
4.10.1 Discussion	
4.11 Metabolomic Analysis	135
4.11.1 Metabolomic Analysis of the Treated MCF-7 with IL-GFE	135
4.11.1.1 Pathway Analysis of Metabolomic Profiles	138
4.11.2 Metabolomic Analysis of the Treated HT29 with IL-GFE	
4.11.2.1 Pathway Analysis of Metabolomic Profiles	
4.11.3 Discussion	
4.12 Summary	
CHAPTER FIVE: CONCLUSION AND RECOMMENDATIONS	148
5.1 Conclusion	
5.2 Recommendations	
REFERENCES	153
APPENDIX A	
APPENDIX B	
APPENDIX C	181

## LIST OF TABLES

Table 2.1	Ethnomedicinal Use of the Graviola Fruit in the World.	13
Table 2.2	Description of the Breast Cancer Staging System	24
Table 2.3	Comparison of the Proposed IL-MAE Approach with Conventional Extraction Methods	33
Table 3.1	Experimental Design and Levels of Independent Process Variables	55
Table 3.2	Experimental Design for Optimisation of Extraction Parameters using RSM	56
Table 4.1	FCCCD Experimental Design and Result of the Responses	80
Table 4.2	ANOVA for Yield (%) Fitted Quadratic Model of Extraction Conditions	82
Table 4.3	The Analysis of the Model Fitting	84
Table 4.4	ANOVA for the IC <sub>50</sub> of IL-GFE on MCF-7 Cells Fitted Quadratic Model of Extraction Conditions	88
Table 4.5	ANOVA for the IC <sub>50</sub> of IL-GFE on HT29 Cells Fitted Quadratic Model of Extraction Conditions	88
Table 4.6	The Analysis of the Model Fitting	90
Table 4.7	Results from the Validation of the Model	94
Table 4.8	Number of spots and polarity index of different combinations of the solvent system	95
Table 4.9	Rf values with different solvents ratio for hexane: dichloromethane: methanol (MP6)	97
Table 4.10	Mobile phase ratios and fractions' combination based on the TLC profile	98
Table 4.11	Qualitative phytochemical components of the crude Graviola fruit extract and its semi-purified fractions (Fr A- F)	99

Table 4.12	Compounds present in the ionic liquid extract of Graviola fruit identified by CG-TOFMS	103
Table 4.13	$IC_{50}$ of IL-GFE and different fractions on MCF-7 and HT-29 cell lines.	109
Table 4.14	Number of generations (X), specific growth rate ( $\mu$ ) and doubling time (td) for untreated MCF-7 cells (control), MCF-7 cells treated with IL-GFE and Taxol	114
Table 4.15	Number of generations (X), specific growth rate ( $\mu$ ) and doubling time (td) for untreated HT29 cells (control), HT29 cells treated with IL-GFE and Taxol	118
Table 4.16	Teratogenicity effects of crude IL-GFE (7.81-500 $\mu$ g/mL) on zebrafish development at 72 hpf	125

## LIST OF FIGURES

Figure 1.1	Graviola fruit (Annona muricata L)	2
Figure 1.2	The flowchart describes the general methodology applied in this research	7
Figure 2.1	(A) Graviola ( <i>Annona muricata</i> L); (B) the appearance of the leaves; (C) flowers and (D) fruits (Moghadamtousi et al., 2015).	12
Figure 2.2	General structure of acetogenins (Bermejo et al., 2005)	15
Figure 2.3	Complex I of the mitochondrial respiratory chain. Source: https://medicinenewbie.blogspot.com/2009/08/	17
Figure 2.4	Cancer classifications (Silverstein et al., 2006)	19
Figure 2.5	Anatomy of a female breast (adopted from the American Cancer Society Web page).	23
Figure 2.6	Anatomy and Physiology of Large Intestine (Martini, 2001).	25
Figure 2.7	An overview of the cell cycle process.	31
Figure 2.8	Growth curve pattern of animal cells, in which $\mu$ is the specific growth rate (Castilho et al., 2008).	41
Figure 2.9	An overview protocol of metabolomics study of adherent mammalian cells (Sellick et al., 2011).	43
Figure 3.1	Flow diagram of the major experiments used in the research.	48
Figure 3.2	Thin Layer Chromatography	58
Figure 3.3	Column Chromatography: A) Silica gel packing, B) Sample dry loading, C) Fractions descending the column.	60
Figure 4.1	Effect of different ionic liquids on Graviola fruit extraction yield. Bars with different letters are significantly different (p<0.05).	77
Figure 4.2	3D Response surface plots show the extraction parameters effect of IL-MAE on the yield of Graviola fruit. (A) The effect of extraction time vs irradiation power on the yield; (B) The effect of	

	irradiation power vs solid-liquid ratio on the yield.	86
Figure 4.3	3D Response surface plots show the extraction parameters (IL-MAE) on the $IC_{50}$ of MCF-7 cell line. (A) The effect of extraction time vs irradiation power on the $IC_{50}$ of MCF-7; (B) The effect of extraction time vs solid-liquid ratio on the $IC_{50}$ of MCF-7; (C) The effect of irradiation power vs solid-liquid ratio on the $IC_{50}$ of MCF-7.	92
Figure 4.4	3D Response surface plots show the extraction parameters (IL-MAE) on the $IC_{50}$ of the HT29 cell line. (A) The effect of extraction time vs irradiation power on the $IC_{50}$ of HT29; (B) The effect of extraction time vs solid-liquid ratio on the $IC_{50}$ of HT29; (C) The effect of irradiation power vs solid-liquid ratio on the $IC_{50}$ of HT29.	
Figure 4.5	TLC Plates of Different Solvent Systems and the Number of Spots	96
Figure 4.6	The percentage of cell viability of breast adenocarcinoma MCF-7 cell lines vs concentrations of crude IL-GFE treatment	106
Figure 4.7	The percentage of cell viability of breast adenocarcinoma MCF-7 cell lines vs concentrations of Taxol treatment	107
Figure 4.8	The percentage of cell viability of colon adenocarcinoma HT29 cell lines vs different concentrations of IL-GFE treatment	107
Figure 4.9	The percentage of cell viability of colon adenocarcinoma HT29 cell lines vs different concentrations of Taxol treatment	108
Figure 4.10	The percentage of cell viability of normal VERO cell lines vs different concentrations of IL-GFE treatment	108
Figure 4.11	The percentage of MCF-7 cell viability vs different concentrations of IL-GFE Fractions.	110
Figure 4.12	The percentage of HT29 cell viability vs different concentrations of IL-GFE Fractions.	110
Figure 4.13	MCF-7 Growth profiles of Untreated (control) and Treated with IL-GFE (IC $_{50}$ =4.75 $\mu g/mL$ ) and Taxol (IC $_{50}$ =0.99 $\mu g/mL$ )	114
Figure 4.14	MCF-7 cells growth at A: exponential phase and B: death phase, for untreated and the treated MCF-7 with IL-GFE and Taxol.	115
Figure 4.15	Representative photographs of A; untreated MCF-7 cells, and B; MCF-7 cells treated with IL-GFE at a designated time point.	116

Figure 4.16	6 HT29 growth profiles of untreated (control) and treated cells with crude IL-GFE (IC <sub>50</sub> =10.56 $\mu$ g/mL) and Taxol (IC <sub>50</sub> =1.22 $\mu$ g/mL).	117
Figure 4.17	7 HT29 cells growth at A: exponential phase and B: death phase, for untreated HT29 and the treated HT29 with crude IL-GFE and Taxol	119
Figure 4.18	Representative photographs of A; untreated HT29 cells, and B; HT29 cells treated with crude IL-GFE at a designated time point.	120
Figure 4.19	Effect of crude IL-GFE (7.81-500 $\mu g/mL$ ) on the survival rate of zebrafish embryos from 0 to 120 hpf. The sample was compared to the untreated group	123
Figure 4.20	) Effect of crude IL-GFE (7.81-500 $\mu g/mL$ ) on zebrafish embryos mortality rate at 120 hours post-fertilization (hpf). The LC <sub>50</sub> value of crude IL-GFE towards zebrafish embryos was 173.45 $\mu g/mL$	124
Figure 4.2	Representative photographs of A: untreated zebrafish (control) and B: the treated zebrafish with crude IL-GFE (500 $\mu g/mL$ ) at a designated time point. Photographs were captured using an inverted microscope at 100X magnification	126
Figure 4.22	2 Effect of crude IL-GFE (7.81-500 μg/mL) on zebrafish embryos hatching rate at 0 to 120 hours post-fertilization (hpf)	127
Figure 4.23	B Heartbeat rates of zebrafish larvae treated with crude IL-GFE (7.81-500 μg/mL) at 96 hpf. Bars with different letters are significantly different (p<0.05).	128
Figure 4.24	Cell cycle distribution of the MCF-7 cells treated with IL-GFE at corresponding IC <sub>50</sub> in a time-dependent manner	132
Figure 4.25	5 Cell cycle phase distribution of the treated HT29 cells with IL-GFE at the corresponding IC <sub>50</sub> in a time-dependent manner	132
Figure 4.26	The percentage of apoptotic MCF-7 cells treated with IL-GFE at the corresponding IC <sub>50</sub> for 24, 48 and 72 hrs.	133
Figure 4.27	Agglomerative hierarchical clustering (AHC) dendrogram of intracellular and extracellular metabolites from the control and the treated MCF-7 cells with IL-GFE.	136
Figure 4.28	Principal component analysis (PCA) of intracellular and extracellular metabolites from the control and the treated MCF-7 cells with IL-GFE.	137

Figure 4.29	Plot of loadings formed by the first two principal components from the PCA of intracellular and extracellular metabolites of the control and the treated MCF-7 cells with IL-GFE.	138
Figure 4.30	Agglomerative hierarchical clustering (AHC) dendrogram of intracellular and extracellular metabolites from the control and the treated HT29 cells with IL-GFE.	140
Figure 4.31	Principal component analysis (PCA) of intracellular and extracellular metabolites from the control and the treated HT29 cells with IL-GFE	
Figure 4.32	Plot of loadings formed by the first two principal components from the PCA of intracellular and extracellular metabolites of the control and the treated HT29 cells with IL-GFE.	142

### LIST OF ABBREVIATIONS

ACS American Cancer Society

ACGs Acetogenins

AHC Agglomeration Hierarchical Clustering

ANOVA Analysis of Variance Annexin-V-FITC Apoptosis Test FITC kit

ATCC American Type Culture Collection

ATP Adenosine Triphosphate
A-549 Human Lung Carcinoma Cells

BC Breast Cancer

BCAAs Branched-Chain Amino Acids

Bcl2 B Cell Lymphoma 2
BSC Biological Safety Cabinet

BSTFA N,O bis(trimethylsilyl)-trifluoroacetamide [C4MIM]BF4 1-butyl-3-methylimidazolium tetrafluoroborate

[C4MIM]Cl 1-butyl-3-methylimidazolium chloride

[C4MIM]PF6 1-butyl-3-methylimidazolium hexafluorophosphate

CC Column chromatography
CV Coefficient of Variation

DB 3 Durian Belanda 3
DCM Dichloromethane
DF Dilution Factor

DMEM Dulbecco's Modified Eagle's Medium

DMSO Dimethyl Sulfoxide
DNA Deoxyribonucleic Acid
DOE Design of Experiments
et al. et alia – and others
etc. and other types

EtOH Ethanol

EU European Union FBS Fetal Bovine Serum

FCCCD Face-Centered Central Composite Design

FDA Food and Drug Administration

GC-TOFMS Gas Chromatography- Time-of-Flight Mass Spectrometry

HepG2 Human Hepatoma Cell Line
HCT-116 Human Colon Cancer Cell Line
HL-60 Human Leukemia Cell Line
HPF Hours Post Fertilisation
HRE Heat Reflux Extraction

Hrs Hours

HT29 Human Colon Adenocarcinoma Cell Line IACUC Institutional Animal Care and Use Committee

IC<sub>50</sub> Half Maximal Inhibitory Concentration IL-GFE Ionic Liquid-Graviola Fruit Extract

IL-MAE Ionic Liquid-based Microwave-Assisted Extraction ISO International Organization for Standardization

KAED Kulliyyah of Architecture and Environmental Design

KMO Kaiser-Meyer-Olkin

LC<sub>50</sub> 50 % Lethality Concentration LDHA Lactic Acid Dehydrogenase-A LLE Liquid-Liquid Extraction

LOF Lack of Fit

MCF-7 Human Breast Adenocarcinoma Cell Line MCF-10A Nontumorigenic Human Breast Epithelial

MeOH Methanol

MMP Matrix Metalloproteinases
MRI Magnetic Resonance Imaging

MTT 4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide

MTS 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-

(4-sulfophenyl)-2H-tetrazolium)

NADH Nicotinamide Adenine Dinucleotide

NCI National Cancer Institute NEAA Nonessential Amino Acids

NHEKN Neonatal Normal Human Epithelial Keratinocytes NIST National Institute of Standards and Technology

NMR Nuclear Magnetic Resonance NMSC Non-Melanoma Skin Cancer

OD Optical Density, unit of Absorbance

OFAT One-Factor-At-a-Time

PACA-2 Human Pancreatic Cancer Lines
PBS Phosphate Buffered Saline
PCA Principal Component Analysis

PC-3 Human Prostate Cancer

PI Polarity Index
PS Phosphatidylserine
p53 Tumor Protein 53
Rf Retardation Factor
rpm Rotation Per Minute

RNase A/PI Ribonuclease A / Propidium Iodide RSM Response Surface Methodology

R.Time Retention Time
SD Standard Deviation
SE Standard Error

SPE Solid-Phase Extraction
TCA Tricarboxylic Acid Cycle
TLC Thin Layer Chromatography
TNM Tumor Node Metastasis

UAE Ultrasound Assisted Extraction

VERO Normal Kidney Cell – African Green Monkey

WST 1;2-(4-Iodophenyl)-3-(4-nitrophenyl)-5-(2,4-disulfophenyl)-

2H-tetrazolium, monosodium salt)

XTT 2,3-bis-(2-methoxy-4-nitro-5-sulfophenyl)-2H-tetrazolium-5-

carboxanilide

## LIST OF SYMBOLS

°C Degree Celsius

% Percentage ± Plus-Minus

< Less than

g Gram

G0 Gap 0 Phase G1 Gap 1 Phase

G2/M Gap 2 / Mitotic Phase

S Synthesis Phase

mg/L Milligram per Milliliter

mol/L Mole per Liter

R<sup>2</sup> Coefficient of Determination

td Doubling Time

 $\mu g/mL$  Microgram per Millilitre

μ Specific Growth Rate

X Number of Generations

#### **CHAPTER ONE**

#### INTRODUCTION

#### 1.1 BACKGROUND OF THE STUDY

Natural plant extracts have been used for a considerable length of time by many cultures and civilisations for the treatment of various health problems. Over 80% of the worldwide population nowadays relies on natural plant extracts through conventional therapies. Many studies have focused on the prevention, progression, and treatment of cancer using natural products. However, there is still room for improvement. Currently, the use of synthetic chemotherapeutic drugs fails to be considered as effective therapeutic agents, and this is because of the greater part of severe toxic side effects to the normal cells. According to Ioannis et al. (2015), an alternative therapeutic approach that uses natural products showed more advantages with fewer side effects.

Numerous herbs and species are used in the world such as ginger, onion, garlic, cardamon, coriander, and turmeric to treat many diseases. Among them, Annona muricata shows enormous medicinal properties. A. muricata Linn is a lowland fruit tree under the Annonaceae family. A. muricata is likewise commonly known as Graviola or Soursop or Guanabana. It is called soursop because of the sour and sweet flavour of its extensive fruit (Patel & Patel, 2016). The Graviola is native to the tropical zones, South America and Africa but is currently widely cultivated in the tropical areas around the world, including Southeast Asia and Southern Florida, from the ocean level to the altitudes of around 1150 meters. Graviola is a thin, small, and cold-intolerant tree, which achieves heights of 4 to 6 meters. Its edible fruits are large, heart-shaped, dark green in

colour, with a diameter which varies between 5-20 cm and an average weight of 0.4 to 1.0 kg (Figure 1.1) (Daddiouaissa & Amid, 2018).



Figure 1.1 Graviola fruit (*Annona muricata* L)

Phytochemical analysis of the plant reveals the presence of alkaloids, phenols, terpenoids, flavonoids and it has an enormous potential anticancerous compound coined as acetogenins which assume to play a vital role towards numerous types of cancer. Acetogenins are potent inhibitors of NADH oxidase (nicotinamide adenine dinucleotide phosphate-oxidase) of the membrane mitochondrial of cancer cells. The fruit is of economic value and consequently cultivated and used broadly as expendable food (Patel & Patel, 2016).

Cancer is a group of diseases that are still considered as one of the leading causes of morbidity and mortality in the world. This disease can be defined as an abnormal growth of cells with the potential to spread or invade to the other tissues of the body. It is caused by mutations in gene expression leading to disequilibrium of cell proliferation and cell death (Ruddon, 2007).

Breast adenocarcinoma is one of the most cancer incidents among women, with an estimated 268,600 new cases, and 41,760 breast cancer (BC) deaths estimated to occur in the United States' women in 2019 (Siegel, Miller, & Jemal, 2019). Therefore,

BC is a challenge among research communities around the world; this is because of the BC incidence rate keep increasing by 0.4 % annually worldwide according to the final report conducted by Jemal and co-workers (2017). On another hand, 101,420 new cases of colon adenocarcinoma were estimated to occur for both sexes and 51,020 estimated deaths in the United States in 2019. Current protocols of treatment include radiation therapy, surgical intervention, and chemotherapy which induce numerous side effects including nausea, fatigue, vomiting, weak of the immune system and hair loss (Griffin et al., 1996). Thus, the search for alternative treatment is necessary.

#### 1.2 PROBLEM STATEMENT AND SIGNIFICANCE OF THE STUDY

Cancer is among the most common causes of mortality in the world, with an estimation of 18.1 million new cancer cases and 9.6 million cancer deaths to occur in 2018 worldwide, projected to rise by at least 70% by 2030 (Antoni et al., 2016; Bray et al., 2018). Among of command cancer treatments are surgery, chemotherapy and radiotherapy. However, more people are suffering every day from the effect of chemotherapy which causes various kinds of undesirable side effects such as hair loss, weakness of the immune system, loss of appetite, hormonal fluctuation, anxiety, depression and some even die not because of cancer but because not able to cope with the side effects. In addition to that, these drugs are costly and not affordable for everyone especially for the patients from low-income families. The development of new alternative anticancer drugs from plants remains one of the most challenging areas of research. Graviola has various pharmacological properties namely anti-cancerous properties, hepatoprotective, antioxidant, antidiabetic, insecticidal and pesticidal properties and anti-microbial activity resulted from its phytoconstituents. Most Graviola bio-compounds were extracted through solvent extraction methods which present many

disadvantages including a large amount of extractant waste, use of hazardous and flammable organic solvents, potential toxicity emissions during extractions. To our knowledge, there are no phytochemicals extracted by ionic liquid microwave-assisted extraction method (IL-MAE) from Graviola fruit. IL-MAE method may be a better choice in producing safe compounds which are solvent-free for human consumption.

Thus, this research aimed to investigate the therapeutic potential of the crude IL-GFE and its isolated active fractions as an anticancer agent. It is strongly believed that the discovery of anticancer properties from Graviola fruit could lead to the development of a new generation of anticancer drugs that possess both chemopreventive and chemotherapeutic properties which are safer and more influential without weakening the patients' health.

#### 1.3 RESEARCH HYPOTHESIS

Natural products can be consumed as alternative medicine that may not have any side effects on individuals. Nowadays, much interest in maintaining health care and efforts are made to accomplish this through the extraction of plants' compounds and evolution in medicinal use. Today, traditional knowledge and practices have contributed to our modern medicine which is attested by more than 40% of commonly prescribed medications throughout the world. Most of them found their origins directly or indirectly in plants or animals (Ahmad & Ismail, 2003).

Different parts of Graviola have been reported and used traditionally to treat many types of diseases. Graviola fruits contain many useful phytochemicals therefore, phytochemicals from Graviola fruit may be the suitable candidate to be used as a chemopreventive medicine in treating cancer.