DNA METHYLATION IN ESSENTIAL HYPERTENSION IN YOUNG ADULTS IN EAST COAST MALAYSIA

BY

WAN FATEIN NABEILA WAN OMAR

A thesis submitted in fulfilment of the requirement for the degree of Doctor of Philosophy (Medical Sciences)

Kulliyyah of Medicine International Islamic University Malaysia

JUNE 2020

ABSTRACT

Hypertension is emerging as the most prevalent risk factor of ischemic heart disease in young adults, but awareness is low in this age group. The prevalence of prehypertension in this population is also high, putting them at higher cardiovascular risk. The pathophysiology of essential hypertension has yet to be fully understood, and epigenetic modifications have been proposed to play some role. To date, very few epigenetic studies were done in young adults with prehypertension and hypertension. The aim of this study was to compare the level of DNA methylation in the promoter of implicated genes in young adults with normotensive blood pressure, prehypertension and hypertension. An observational cross-sectional study was conducted among 240 subjects age 18 to 45 years in Kuantan, Pahang, Malaysia. Eighty subjects were recruited for each blood pressure group; normotension, prehypertension, and hypertension as defined by the Ministry of Health Malaysia Clinical Practice Guidelines 4th edition. MethyLight analysis was performed to determine DNA methylation levels of IL-6, ADD1 and AGTR1 gene promoter in the blood. Differentially methylated genes in prehypertension and/or hypertension group were followed by gene expression study (n = 10 per group). There was no significant difference in IL-6 methylation between hypertensive and normotensive. IL-6 predicted prehypertension in males (p = 0.014), but not females. Hypertensive and prehypertensive males, and prehypertensive females, had lower ADD1 methylation than their respective normotensive counterparts. After adjusting for other covariates, ADD1 methylation predicted prehypertension and hypertension in males (p = 0.002)and p = 0.034 respectively). There was no significant difference in AGTR1 methylation between the three groups in both sexes. There was no significant association between IL-6 and ADD1 methylation level and gene expression level. DNA methylation of *IL-6* and *ADD1* are independent predictors of prehypertension and/or hypertension in males hence has potential as an adjunct biomarker for risk stratification or disease progression. This is the pioneering study of IL-6, ADD1 and AGTR1 methylation in prehypertensive and hypertensive young adults. Further study to delineate potential mechanisms linking DNA methylation to disease development is warranted.

خلاصة البحث

يظهر فرط ضغط الدم عامل الخطر الأكتر إنتشارا لمرضى القلب الاقفاري لدي البالغين, لكن الوعي منخفض لهده الفئة العمرية. كما أن انتشار فرط ضغظ الدم في هده الفئة العمرية من السكان مرتفع ايضا, مما يعرضهم لخطرأعلى لأمراض الأوعية الدموية والقلب. الفسيولوجيا المرضية لفرط ضغط الدم الأساسي لم يتم فهمه بالكامل, لّذلك تم إقتراح لتعديلات الجينيه لتلعب بعض الدور في هذا المرض. حتى الآن عدد قليل جدًا من الدرسات الجينيه لتى تم إجراؤها على البالغين ما قبل فرط ضغط الدم وفرط ضغط الدم. وكان الهدف من هذه الدراسة هومقارنة مستوي ميثيل الحمض النووي في محفز الجينات لدي البالغين ذوي الضغط الطبيعي, وماقبل فرط ضغط الدم, وفرط ضغط الدم. أحريت دراسة مقطعية رصدية لعدد 240 شخصا تتراوح أعمارهم ما بين 18 الي 45 سنة في مدينة كونتان بولاية باهانج, ماليزيا. تم تحديد ثمانين شخصا لكل المجموعات من ضغط الدم الطبيعي, و ما قبل فرط ضغط الدم, وفرط ضغط الدم. وذلك طبقا للنحو المحدد من قبل وزارة الصحة الماليزية للممارسة التوجهية الطبعة الرابعة. تم إجراء تحليل ميثيل لايت (MethyLight) لتحديد مستوي ميثيل الحمض النووي من مروج الجينات (ADD1) و(IL-6) و(AGTR1) في الدم. الاختلافات في الميثيل الجيني لمجموعة ما قبل فرط ضغط الدم وفرط الضغط تبعت بدراسة تفصيلية للتعبير الجيني لعدد 10 أشخاص لكل مجموعة. لم يكن هناك فرق كبيرفي ميثيل (IL-6) بين مجموعة ضغط الدم الطبيعي و فرط ضغط الدم IL-6. تنبأ بما قبل فرط ضغط الدم للذكور (p = 0.014) ولكن ليس للإناث. الذكور المصابين بفرط ضغط الدم و ماقبل فرط ضغط الدم والأناث المصابين بما قبل فرط ضغط الدم كان لديهم اقل ميثيل للحين (ADD1) من الاشخاص ذو الضغط الدم الطبيعي لنفس الجنس. بعد تعديل المتغير ات المشتركة , p = 0.002) قبل ارتفاع ضغط الدم وارتفاع ضغط الدم لدي الذكور (ADD1 قبل الأخرى, تنبأت مثيلة على التوالى). لم يكن هناك فرق كبير في ميثيل (AGTR 1) في الثلاثة مجموعات لكلا p=0.034الجنسين. كذلك لم يكن هناك إرتباط كبير في ميثيل الجين (IL-6) ومستوي ميثيل (ADD1) ومستوي التعبير الجيني. ميثيل الحمض النووي ل IL-6 و IL-6 مؤشر تنبؤ مستقل لما قبل فرط ضغط الدم وفرط ضغط الدم عند الذكور. ومن ثم قد تكون مؤشر حيوي مساعد لتحديد مخاطر المرض وتطوره.هذه دراسه الرائده AGTR 1 وADD1 وIL-6 لدي الشباب البالغين الذين بعانون من ارتقاع ضغط الدم وما قبل ارتفاع ضغط الدم. لذلك هناك مايبررالمزيد من الدرسات لتحديد الألية المحتملة التي تربط ميثيل الحمض النووي بنمو المرض.

APPROVAL PAGE

The thesis of Wan Fatein Nabeila Wan Omar has been approved by the following:

Aszrin Abdullah Supervisor
Norlelawati A. Talib Co-Supervisor
Jamalludin Ab. Rahman Co-Supervisor
Azarisman Shah Mohd. Shah Co-Supervisor
Solachuddin Jauhari Arief Ichwan Internal Examiner
Abdul Rashid Abdul Rahman External Examiner 1
Muhammad Farid Johan External Examiner 2
Mohd Zulfaezal Che Azemin Chairman

DECLARATION

I hereby declare that this thesis is the result of my own investigation, except where

otherwise	stated.	I also	declare	that	it	has	not	been	previously	or	concurre	ently
submitted	as a who	ole for a	any other	degre	ees	at II	ШM	or oth	er institutio	ns.		
Wan Fatei	n Nabeil	la Wan	Omar									
Signature.			• • •						Date			

INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA

DECLARATION OF COPYRIGHT AND AFFIRMATION OF FAIR USE OF UNPUBLISHED RESEARCH

DNA METHYLATION IN ESSENTIAL HYPERTENSION IN YOUNG ADULTS IN EAST COAST MALAYSIA

I declare that the copyright holder of this thesis is International Islamic University Malaysia.

Copyright © 2020 International Islamic University Malaysia. All rights reserved.

No part of this unpublished research may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without prior written permission of the copyright holder except as provided below

- 1. Any material contained in or derived from this unpublished research may be used by others in their writing with due acknowledgement.
- 2. IIUM or its library will have the right to make and transmit copies (print or electronic) for institutional and academic purposes.
- 3. The IIUM library will have the right to make, store in a retrieved system and supply copies of this unpublished research if requested by other universities and research libraries.

By signing this form, I acknowledged that I have read and understand the IIUM Intellectual Property Right and Commercialization policy.

Affirmed by Wan Fatein Nabeila Wan Omar	
Signature	Date

ACKNOWLEDGEMENTS

Alhamdulillah, with His grace, this thesis is completing, and the author's journey is (finally) progressing onto the next chapter. Many believe that PhD is the beginning, rather than the end. So hello world, I am back!

It has been set as an indisputable rule, that only one person can author a thesis. Well, in case there is a change in the rule—though *very* unlikely—and I can choose a co-author, I would have put Ammar bin Yusop next to my name. My husband may not contribute to the intellectual and academic content of this thesis, but his endless support and assistance in many aspects of my non-academic life could not be emphasised too much.

Some people did thank their children whom without their presence could have expedited the whole process by at least half, duration-wise. I would not deny that sentiment wholly, but I choose to thank my three beautiful minions; Khaulah, Wam and Hilya, for being so loving and tolerating, although it was hard to comprehend my problem statements that I vented out. Thank you Umi, Mak and Abah, for keeping me (and this thesis) in your prayers.

Of course, my heartiest appreciation would definitely go to Dr Aszrin, whom I leisurely address as Kak Rin (with her consent of course). She goes way beyond being an academic advisor, a project supervisor or a postgraduate coordinator. Often, she becomes a mother—offering advice, logistic assistance and beyond, especially for someone who is relatively new to Kuantan. Sometimes, she is more like a sister, comforting and protecting her baby sister. But most of the times, she is a sahabah, a dear accompany whom I can pour my thoughts, feelings and views, minus the fear of judgement. For all these and more to come, thank you Kak Rin.

Dr Norlela is the chairperson of the supervisory committee. I am always at awe when she speaks. She is highly-knowledgeable, intuitive, very experienced, yet remained so humble. In my humble attempt at being at least one-tenth as good as her, I am (kind of) addicted to meet and learn from her but am guilty realizing that too much too long of an appointment could cost substantial delay in her reviewing her patients' results. I really appreciate Dr Lela's boundless commitment for academic consultation and delivering timely feedback in between the stack of documents to sign and H&E slides to review.

I wish to also write about Prof Jamal, Prof Azarisman, Kak Baiyah, Kak Shikin, Kak Huda, Dr Solah and every single soul who has showered me with unlimited assistance and unwavering support, but then it is gonna be too long. After all, I was advised that very few, *handpicked* people shall read the thesis; the student, supervisor(s) and examiners, that's about it. Thank you for reading (that is, *if* you do).

TABLE OF CONTENTS

Abstract	ii
Abstract in Arabic	
Approval Page	
Declaration	
Copyright	vi
Acknowledgements	
List of Tables	
List of Figures	xvi
List of Abbreviations	xix
CHAPTER ONE: INTRODUCTION	1
1.1 Overview of Study	
1.2 Statement of the Problem	
1.3 Aim of the Study	
1.4 Conceptual Framework	
1.5 Theoretical Framework	
1.6 Research Question	6
1.7 Research Objectives	6
1.7.1 General Objective	
1.7.2 Specific Objectives	
1.8 Research Hypothesis	
1.9 Significance of the Study	7
1.10 Chapter Summary	8
CHAPTER TWO: LITERATURE REVIEW	
2.1 Hypertension	
2.1.1 Overview of Prehypertension and Hypertension	
2.1.2 Definition of Hypertension	
2.1.3 Classification of Blood Pressure and Recognition of Prehypert Status	
2.2 Epidemiology	
2.2.1 Prehypertension and Hypertension in Adults	
2.2.2 Prehypertension and Hypertension in Young Adults	
2.3 Burden of Disease	
2.3.2 Mortality and Morbidity	
2.3.3 Economic Impact	
2.4 Factors Associated with Prehypertension and Hypertension in	
Adults	_
2.4.1 Other Cardiovascular Morbidities	
2.4.2 Dietary	
2.4.3 Early Life Factor	
2.4.4 Psychosocial Factor	
2.5 Physiology of Blood Pressure Regulations	
2.5.2 Determinants and Regulation of Blood Pressure	
2.5.3 Regulation of Blood Pressure	

2.6 Establishing Hypertension	36
2.6.2 Measuring Blood Pressure	36
2.6.2.2 Diagnosing Hypertension	38
2.7 Causes of Hypertension	38
2.8 Essential Hypertension	39
2.8.1 Definition of Essential Hypertension	39
2.8.2 Diagnosing Essential Hypertension	39
2.8.3 Pathophysiology of Essential Hypertension	40
2.8.3.2 Renin-Angiotensin-Aldosterone	41
2.8.3.3 Inflammation	43
2.8.3.4 Chronic Activation of the Sympathetic Nervous System	46
2.9 Epigenetics	51
2.9.2 Definition	51
2.9.3 Mechanisms of Action	52
2.9.3.2 Histone Modification	53
2.9.3.3 Non-coding Ribonucleic Acid (ncRNA)	54
2.10 DNA Methylation	
2.10.2 Definition	
2.10.3 Function of DNA Methylation	57
2.10.4 Variation in DNA Methylation Measurement	
2.10.5 DNA Methylation in Hypertension	
2.10.5.2 Global DNA Methylation	
2.10.5.3 DNA Methylation of Candidate Gene	
2.10.6 Methods of Measuring DNA Methylation	
2.10.6.2 MethyLight	
2.10.7 DNA Source in Genetic Study	
2.11 Review on Gene of Interest	
2.11.2 Interleukin–6	
2.11.3 α–Adducin (<i>ADD1</i>)	
2.11.4 Angiotensin II Receptor Type 1 (AGTR1)	
2.12 Chapter Summary	
CHAPTER THREE: METHODOLOGY	87
3.1 Study Flow	
3.2 Ethical Approval	
3.3 Study Design	
3.4 Subject Recruitment	
3.4.1 Sample Size Calculation	
3.4.2 Sampling Method	
3.5 Study Protocol	
3.5.2 Sociodemographic Data	
3.5.3 Anthropometric Data	
3.5.4 Haemodynamic Parameters	
3.5.5 Biochemical Profile	
3.6 Venous Blood Processing	
3.7 DNA Methylation study	
3.7.2 Deoxyribonucleic Acid (DNA) Extraction	
3.7.3 DNA Purity Determination	
3.7.4 DNA Integrity Determination	
3.7.4.2 Agarose Gel Preparation	
	5 5

3.7.4.3 DNA Sample Preparation and Gel Electrophoresis	
3.7.4.4 DNA Band Inspection	
3.7.5 DNA Concentration Determination and DNA Sample Dilution	. 101
3.7.6 Bisulphite Conversion of DNA	. 102
3.7.6.2 Reagent Preparation	. 103
3.7.6.3 Bisulphite Conversion of DNA Samples	. 104
3.7.6.4 Bisulphite–DNA Quantification and Dilution	. 105
3.7.7 Gene Selection and Primer Design	
3.7.8 MethyLight Optimization	. 106
3.7.8.1 Primer Reconstitution	. 106
3.7.8.2 Optimum Annealing Temperature Determination	. 107
3.7.8.3 Primer Sensitivity and Specificity	. 108
3.7.8.4 Serial Percentage of Methylated DNA	. 110
3.7.9 MethyLight Reaction	. 111
3.7.9.2 Data Analysis	. 111
3.8 Gene Expression Study	. 112
3.8.2 Selection of Samples	
3.8.3 Ribonucleic Acid Extraction	
3.8.4 Determination of RNA Concentration and Purity	. 116
3.8.5 Determination of RNA Integrity	. 116
3.8.5.2 Setting up QIAxcel Gel Cartridge and Buffer Tray	. 117
3.8.5.3 RNA Sample Preparation	. 117
3.8.5.4 Determination of RNA Quality	. 118
3.8.6 Complementary DNA Synthesis	. 118
3.8.7 Primer Design	
3.8.8 Gene Expression Assay Optimization	. 120
3.8.8.1 Annealing Temperature Optimization	. 120
3.8.8.2 Evaluating Amplification Efficiency of the qPCR Assay	
3.8.9 Gene Expression Assay by Quantitative Polymerase Chain Rea	ction
3.9 Statistical Analyses	. 122
CHAPTER FOUR: RESULTS AND FINDINGS	
4.1 Total Number of Patients Screened	
4.2 Sociodemographic Distribution of Subjects	
4.2.2 Sociodemographic Distribution of Subjects by Blood Pressure S	
	. 125
4.2.3 Sociodemographic Distribution of Subjects by Sex	
4.3 Haemodynamic Parameters of Subjects	
4.4 Biochemical Profiles of Subjects	
4.4.2 Biochemical Profiles of Subjects by Blood Pressure Status	
4.4.3 Biochemical Profiles of Subjects by Sex	
4.5 DNA Integrity Analysis	
4.6 DNA Methylation Level Analysis	
4.6.2 Optimisation of Reaction Conditions	
4.6.2.1 Selection of Annealing Temperature (T _a)	. 135
4.6.2.2 Determination of Assay Efficiency	. 135
4.6.2.2 Determination of Assay Efficiency	135 136
4.6.2.2 Determination of Assay Efficiency	. 135 . 136 . 137

4.6.5 Methylation Level of Angiotensin II Receptor Type 1 (AGTR)) 139
4.7 Associations between DNA Methylation and Hemodynamic Paramet	ers 141
4.7.1 Association between IL-6 Methylation and Hemod	ynamic
Parameters	141
4.7.2 Association between AGTR1 Methylation and Haemod	ynamic
Parameters	•
4.7.3 Association between ADD1 Methylation and Haemod	
Parameters	-
4.8 Association between DNA Methylation and Other Related Covariate	141
4.8.1 Association between <i>IL-6</i> Methylation and hsCRP	
4.9 Multivariate Analysis of DNA Methylation with Blood Pressure	
4.9.2 Interleukin-6 (<i>IL-6</i>)	
4.9.3 α-adducin (<i>ADD1</i>)	
4.9.4 Angiotensin II Receptor Type 1 (AGTR1)	
4.9.5 Summary of Predictors of Prehypertension and Hypertension	
Young Adults	
4.10 Gene Expression Study	
4.10.2 RNA Quality Control	
4.10.3 Optimisation of Reaction Conditions	
4.10.3.1 Selection of Annealing Temperature (T _a)	
4.10.3.2 Determination of Efficiency of Assay	
4.10.4 Quantitative Polymerase Chain Reaction for Gene Expression	
4.10.4.1 Interleukin-6 (IL-6) mRNA Gene Expression Analysis	
4.10.4.2 α-adducin (ADD1) mRNA Gene Expression Analysis	
CHAPTER FIVE: DISCUSSION	169
5.1 Overview of the Study	
5.2 Sociodemographic Characteristics of Subjects	
5.3 Haemodynamic Parameters of Subjects	
5.4 Biochemical Profiles of Subjects	
5.5 Use of Peripheral Blood as Source of DNA in Hypertension study	
5.6 MethyLight as Quantitative Measurement of DNA Methylation	
5.6.2 Proposing a Revised Unit for MethyLight	
5.7 Justification of Inclusion of Covariates in Multivariate Analysis	
5.8 DNA Methylation Variation across Blood Pressure	
5.8.1 Relationship between Interleukin-6 Methylation, Gene Exp	
and Blood Pressure	
5.8.2 Relationship between α-adducin Methylation, Gene Expressi	on and
Blood Pressure	
	vlation
5.8.3 Relationship between Angiotensin II Receptor Type 1 Meth	
5.8.3 Relationship between Angiotensin II Receptor Type 1 Meth and Blood Pressure	197
5.8.3 Relationship between Angiotensin II Receptor Type 1 Meth and Blood Pressure	197 200
5.8.3 Relationship between Angiotensin II Receptor Type 1 Meth and Blood Pressure	197 200 200
5.8.3 Relationship between Angiotensin II Receptor Type 1 Methand Blood Pressure	197 200 200 201
5.8.3 Relationship between Angiotensin II Receptor Type 1 Meth and Blood Pressure	197 200 200 201
5.8.3 Relationship between Angiotensin II Receptor Type 1 Meth and Blood Pressure	197 200 201 201 201
5.8.3 Relationship between Angiotensin II Receptor Type 1 Meth and Blood Pressure	197 200 201 201 201 202

CHAPTER SIX: CONCLUSION	205
6.1 Conclusion	205
6.2 Future Studies	206
REFERENCES	208
APPENDIX I: ETHICAL APPROVAL	239
APPENDIX II: PATIENT INFORMATION SHEET	244
APPENDIX III: CASE RECORD FORM	247
APPENDIX IV: OPTIMISATION OF METHYLIGHT ASSAYS	256
APPENDIX V: OPTIMISATION OF GENE EXPRESSION ASSAYS	259
APPENDIX VI: PRIMER SEQUENCE IN METHYLIGHT ASSAYS	261
APPENDIX VII: COMPARISON OF BASELINE CHARACTERISTICS	oF
STUDY SUBJECTS WITH PREVIOUS STUDIES	263
APPENDIX VIII: LIST OF PUBLICATIONS	264
APPENDIX IX: LIST OF WORKS PRESENTED	267
APPENDIX X: LIST OF AWARDS	273

LIST OF TABLES

Table 2.1	Changes in blood pressure classification according to Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure.	12
Table 2.2	Blood pressure classification for adults according to Malaysia Clinical Practice Guideline.	13
Table 2.3	Blood pressure classification according to ACC/AHA Hypertension Guidelines 2017	14
Table 2.4	Prevalence of hypertension in general adult population in Asia countries	16
Table 2.5	Prevalence of unknown and known hypertension in Malaysia by age	17
Table 2.6	Prevalence of hypertension and prehypertension in young adults	21
Table 2.7	Secondary causes of hypertension	38
Table 2.8	Global DNA methylation in hypertensive subjects	60
Table 2.9	DNA methylation study of candidate gene of essential hypertension in animal studies	62
Table 2.10	DNA methylation study of candidate genes in hypertension, or blood pressure variation in human	63
Table 2.11	The different methods of measuring DNA methylation	68
Table 3.1	The inclusion and exclusion criteria of subjects recruited in the study	90
Table 3.2	Blood pressure status as classified by Clinical Practice Guidelines (CPG) Management of Hypertension 4 th edition	90
Table 3.3	Collection of venous blood	95
Table 3.4	Components of Gentra® Puregene® Blood Kit (Qiagen, USA).	97
Table 3.5	Components of QuantiFluor® ONE dsDNA System	101
Table 3.6	Components of EZ-96 DNA Methylation-Gold TM Kit (Zymo Research, USA)	103

Table 3.7	Protocol for CT–Conversion Reaction	104
Table 3.8	Sequence of primer-probe used in MethyLight study	106
Table 3.9	Components of master mix for MethyLight optimisation	107
Table 3.10	Protocol for annealing temperature optimization	108
Table 3.11	Preparation of serial dilution of bisulphite-treated DNA for assay efficiency	108
Table 3.12	Components of master mix for MethyLight assay efficiency	109
Table 3.13	Protocol for MethyLight assay	110
Table 3.14	Preparation of serial percentage of bisulphite-treated DNA	110
Table 3.15	Components of MethyLight reaction	111
Table 3.16	Components of RiboPure™ RNA Purification Kit, Blood (Life Technologies Corporation, USA)	113
Table 3.17	Detail of pre-designed QuantiTect Primer Assays (Qiagen, USA) used in gene expression study	119
Table 3.18	Components of master mix for gene expression optimisation	120
Table 3.19	Annealing temperature optimization and melt curve protocol	120
Table 3.20	Preparation of cDNA sample for efficiency assay	121
Table 3.21	Assay efficiency determination protocol	121
Table 4.1	Sociodemographic distribution of the subjects	125
Table 4.2	Sociodemographic distribution of the subjects by blood pressure status	126
Table 4.3	Sociodemographic distribution of the subjects by sex	128
Table 4.4	Haemodynamic parameters of the subjects by blood pressure status	129
Table 4.5	Hemodynamic parameters of the subjects by sex	129
Table 4.6	Biochemical profile of the subjects	130
Table 4.7	Biochemical profile of the subjects by blood pressure status	131
Table 4.8	Biochemical profiles of subjects by sex	133

Table 4.9	The quantification cycle (C_q) of genes of interest and Alu using the temperature gradient protocol	135
Table 4.10	C_q values of each gene of interest and Alu in serial methylation percentage assay using mixed bisulphite-treated standard 100% methylated and non-methylated human DNA.	136
Table 4.11	DNA methylation of Interleukin-6 (IL-6)	137
Table 4.12	DNA methylation of α -adducin (ADD1)	138
Table 4.13	DNA methylation of angiotensin II receptor type 1 (AGTR1)	140
Table 4.14	Multivariate analysis of effect of DNA methylation of <i>IL-6</i> on blood pressure	144
Table 4.15	Multivariate analysis of effect of DNA methylation of <i>IL-6</i> on blood pressure by sex	146
Table 4.16	Multivariate analysis of effect of DNA methylation of IL -6 converted to antilog $C_q[IL$ -6/ Alu] on blood pressure by sex	148
Table 4.17	Multivariate analysis of effect of DNA methylation of <i>ADD1</i> on blood pressure	150
Table 4.18	Multivariate analysis of effect of DNA methylation of <i>ADD1</i> on blood pressure by sex	152
Table 4.19	Multivariate analysis of effect of DNA methylation of $ADD1$ converted to antilog $C_q[ADD1/Alu]$ on blood pressure by sex	154
Table 4.20	Multivariate analysis of effect of DNA methylation of AGTR1 on blood pressure	156
Table 4.21	Multivariate analysis of effect of DNA methylation of <i>AGTR1</i> on blood pressure by sex	158
Table 4.22	Multivariate analysis of effect of DNA methylation of $AGTR1$ converted to antilog $C_q[AGTR1/Alu]$ on blood pressure	160
Table 4.23	Significant factors of prehypertension and hypertension in general, male and female subjects adjusted to specific gene methylation and other confounders	161
Table 4.24	The quantification cycle (C_q) of genes of interest and Alu using the temperature gradient protocol	164

LIST OF FIGURES

Figure 1.1	The conceptual framework of the study.	4
Figure 1.2	The relationship between DNA methylation to progression of hypertension from normotension through prehypertension.	5
Figure 2.1	Factors affecting blood pressure. RAAS = renin-angiotensin-aldosterone system.	36
Figure 2.2	The Renin-angiotensin-aldosterone system serves as a long-term regulator of blood volume and blood pressure.	42
Figure 2.3	Triangulation of neurogenic hypertension.	50
Figure 2.4	Illustration of DNA methylation process which changes cytosine to 5-methyl-cytosine (5mC).	55
Figure 2.5	Illustration of DNA methylation phenomenon in the genome.	56
Figure 2.6	Illustration of theoretical effect of DNA methylation on gene expression.	57
Figure 2.7	Available methods of quantifying or qualifying DNA methylation.	65
Figure 2.8	Illustration of changes in final product of DNA following sodium bisulphite conversion and amplification.	66
Figure 2.9	Cytogenetic location of interleukin–6 (<i>IL</i> –6) gene	74
Figure 2.10	The bisulphite-treated methylated sequence of <i>IL-6</i> promoter region.	76
Figure 2.11	Adducin monomer	77
Figure 2.12	Cytogenetic location of α –Adducin (<i>ADD1</i>) gene	78
Figure 2.13	Illustration of effect of mutated Adducin in reducing sodium-potassium adenosine triphosphatase (Na ⁺ /K ⁺ -ATPase) endocytosis.	80
Figure 2.14	The bisulphite-treated methylated sequence of <i>ADD1</i> promoter region.	81
Figure 2.15	Cytogenetic location of Angiotensin II receptor type 1 (AGTR1) gene	82

Figure 2.16	Overview of the bisulphite-treated methylated sequence of <i>AGTR1</i> promoter region	84
Figure 2.17	The conceptual framework of the study.	86
Figure 3.1	Flow of the present study.	87
Figure 3.2	Sample size calculation.	89
Figure 3.3	Schematic representation of sampling method and sampling sites.	91
Figure 3.4	Summary of study protocol.	92
Figure 3.5	Collection and stabilisation of blood sample for mRNA gene expression study.	96
Figure 3.6	Overview of DNA methylation study.	97
Figure 3.7	A simplified illustration of steps involved in genomic DNA purification.	98
Figure 3.8	Illustration of summary of protocol of bisulphite conversion of DNA samples.	102
Figure 3.9	Preparation of DNA samples of serially diluted concentrations for determination of primer sensitivity and specificity.	108
Figure 3.10	Overview of gene expression study.	112
Figure 3.11	Overview of sample selection in gene expression study for one gene.	113
Figure 3.12	Summary of ribonucleic acid purification.	114
Figure 4.1	Comparison of age, body mass index, waist circumference among subjects, between three blood pressure status.	127
Figure 4.2	Comparison of mean biochemical profile between three blood pressure status.	132
Figure 4.3	Image of agarose gel electrophoresis of purified DNA	134
Figure 4.4	Mean methylation level of Interleukin-6 (<i>IL</i> -6) in three blood pressure groups by sex.	138
Figure 4.5	Mean methylation level of α -adducin ($ADD1$) in three blood pressure groups by sex.	139
Figure 4.6	Mean methylation level of Angiotensin II receptor type 1 (AGTR1) in three blood pressure groups by sex	140

Figure 4.7	Correlation between HsCRP and <i>IL-6</i> methylation analysed using Spearman's rho correlation test.	142
Figure 4.8	Flow chart of gene expression study following differentially methylated genes.	162
Figure 4.9	The superimposed electropherogram view a representative RNA sample	163
Figure 4.10	Gene expression of Interleukin-6 (IL-6).	165
Figure 4.11	Gene expression of Interleukin-6 (<i>IL-6</i>) high and low-methylated samples.	166
Figure 4.12	Gene expression of α -Adducin (ADD1).	167
Figure 4.13	Gene expression of α -Adducin (<i>ADD1</i>) high and low-methylated samples.	168
Figure 5.1	<i>IL-6</i> PCR product analysed in current and previous studies.	190
Figure 5.2	ADD1 PCR product analysed in current and previous studies.	194
Figure 5.3	AGTR1 PCR product analysed in current and previous studies.	198

LIST OF ABBREVIATIONS

A Adenine

ACC American College of Cardiology ACE Angiotensin converting enzyme gene

ACTBBeta-actin geneADDI α -Adducin geneADHAntidiuretic hormoneADRBAdrenergic receptor gene

AGTR1 Angiotensin II Type 1 Receptor gene

AHA American Heart Association

Alu Sequence of Alu gene

Ang II Angiotensin II

Anti-HPT Anti-hypertensive medications

ANOVA Analysis of variance

ATIaR Angiotensin II Type 1 Receptor gene

B Coefficient

baPWV brachial—ankle pulse wave velocity

BMI Body mass index

C cytosine

CARDIA Coronary Artery Risk Development in Young Adults Study

cDNA Complementary DNA

CH₃ Methyl group
CI Confidence interval
CO Cardiac output

CPG Clinical Practice Guidelines

CpG Cytosine-phospodiesterase bond-guanidine

C_q Quantitation cycle
 DBP Diastolic blood pressure
 DNA Deoxyribonucleic acid
 DNMT DNA methyl transferase
 df Degree of freedom

ECV Effective circulating volume EH Essential hypertension FBG Fasting blood glucose

G Guanidine

GAD Generalised anxiety disorder

GAPDH Glyceraldehyde-3-phosphate dehydrogenase gene

GCK Glucokinase gene

GRACE Global Registry for Acute Coronary Effect
HbA1c Glycosylated haemoglobin, type A1c
HDLC High density lipoprotein cholesterol
Hpt Newly-diagnosed hypertensive subjects

HR Heart rate

hsCRP High sensitivity C-reactive protein

HSD11β2 11-β-hydroxysteroid dehydrogenase 2 gene

ICAM-1 Intercellular adhesion molecule-1

IFNInterferon gene $IL-1\beta$ Interleukin- 1β IL-6Interleukin-6 geneIQRInterquartile range

IREC Institutional Research Ethics Committee

JNC Joint National Committee on Prevention, Detection, Evaluation,

and Treatment of High Blood Pressure

LINE Long interspersed nuclear elements gene LDLC Low density lipoprotein cholesterol

MAP Mean arterial pressure

MCP-1 Monocyte chemotactic protein-1

MDA Malondialdehyde

MDD Major depressive disorder
MLPD Maternal low protein diet
MMF Mycophenolate mofetil

MREC Medical Research Ethics Committee

mRNA Messenger ribonucleic acid MSNA Muscle sympathetic nerve activity

MTHFD Methylenetetrahydrofolate dehydrogenase gene

n Number Na⁺ Sodium

NCVD-ACS National Cardiovascular Disease-Acute Coronary Syndrome

Registry

NFKB nuclear factor kappa B

NHMS National Health and Morbidity Survey

NKCC Sodium–potassium–chloride co–transporter gene

NMRR National Medical Research Registry

Nt Normotensive subjects

Obs/Exp CpG Observed to expected CpG ratio

OR Odd ratio

p Significant level

PBL Peripheral blood leukocytes PCR Polymerase chain reaction PMR Percentage methylation ratio

POMC Proopiomelanocortin

PP Pulse pressure PR Pulse rate

Pre Prehypertensive subjects
R Resistance in blood flow
r Radius of blood vessel
r Correlation coefficient

RAAS Renin-angiotensin-aldosterone system

RM Malaysian Ringgit SBP Systolic blood pressure

SCNN Epithelial sodium channel gene

sd Standard deviation SE Standard error

SHMT1 Serine hydroxymethyltransferase gene

SHR Spontaneous hypertensive rats.
SNP Single nucleotide polymorphism

SULFSulfatase geneSVStroke volumeTThymine

TC Total cholesterol

TC/HDL Total cholesterol to high density lipoprotein ratio

TLR Toll–like receptor

TNF- α Tumor necrosis factor α TPR Total peripheral resistance

U Uracil

USD United States Dollar

VCAM-1 Vascular cell adhesion molecule-1

WHO World Health Organisation

5mC 5-methylcytosine

CHAPTER ONE

INTRODUCTION

1.1 OVERVIEW OF STUDY

Cardiovascular diseases dictates the highest mortality and morbidity for non-communicable disease worldwide, accounting for approximately 17 millions death per annum, or one third of total death, and is projected to further rise in 2030 (World Health Organization, 2013). The main pathophysiology of cardiovascular disease is the development of atherosclerosis, which is associated with several risk factors including hypertension, obesity, smoking, dyslipidaemia, diabetes mellitus and family history of cardiovascular disease. Of all deaths due to cardiovascular disease, hypertension alone is responsible for 45 to 51 % of ischemic heart disease and stroke death (World Health Organization, 2013).

Hypertension was also the most prevalent cardiovascular disease risk factors among the acute coronary syndrome (ACS) patients in Malaysia according to the latest National Cardiovascular Disease–Acute Coronary Syndrome Registry (NCVD–ACS 2011–2013) (Wan Ahmad & Sim, 2015). This is contrasting the global data reported by the Global Registry for Acute Coronary Effect (GRACE) study in which smoking precedes the other cardiovascular disease risk factors (Global Registry for Acute Coronary Effect, 2007). It is also important to note that the mean age of ACS patients in Malaysia is approximately 6.5 years younger than other countries included in the GRACE study.

In 95 % of hypertension cases, there is no exact cause identified and therefore is termed as essential hypertension (Carretero & Oparil, 2000). National Morbidity

and Health Survey in 2011 indicated that as much as two-third of adults age 18 years and above have raised blood pressure, with almost half of the adult population have prehypertension—the pre-disease transition state between normotension to hypertension (Naidu et al., 2019). Furthermore, the awareness of hypertension among Malaysians is low, especially in younger age group of age 18 to 54 years (Institute for Public Health (IPH), 2015a). Additionally, prehypertension largely affects young adults and is associated with higher cardiovascular risk, especially in young adults (Egan & Stevens-Fabry, 2015; Elliott & Black, 2007). An earlier study in United States—the Framingham Heart Study—reported that up to 37 % of prehypertensive cases below 65 years, and up to 50 % above 65 years convert to hypertension in 4 years (Vasan et al., 2001). Meanwhile, a more recent local study indicated that the prehypertension—hypertension conversion rate was 69% in 10 years (Ching et al., 2012).

The exact cause of essential hypertension remains unknown although evidences have suggested that both genetic and environmental factors have roles in its pathophysiology (Carretero & Oparil, 2000; Kunes & Zicha, 2009). Nevertheless, several mechanisms were proposed to be involved, for example, inflammatory, abnormal sodium handling and the renin angiotensin aldosterone system (Montecucco et al., 2011; Orlov et al., 2014; Solak et al., 2016). Most studies into the pathophysiology of essential hypertension focused on genetic polymorphism, gene expression, and protein expression in these implicated pathways. Genetic–environmental interaction that underlies essential hypertension may be explained by the epigenetics phenomenon, in which alteration in the gene expression regulation occurs in response to environmental stimuli without changing the nucleotide sequence (Millis, 2011; Raftopoulos et al., 2015). One of the most understood epigenetic

mechanisms is deoxyribonucleotide acid (DNA) methylation. It is hypothesised that DNA methylation at the promoter region of a gene could alter the gene expression at the transcription level (D. H. K. Lim & Maher, 2010). Since modification of DNA methylation is implicated in many complex diseases from cardiovascular, metabolic, cancer and mental health diseases, it is proposed that DNA methylation could also affect the pathways involved in blood pressure regulation (Baccarelli, Wright, et al., 2010; D. H. K. Lim & Maher, 2010).

1.2 STATEMENT OF THE PROBLEM

DNA methylation serves the bridge linking between environmental factors and genetics onto phenotypes. Changes in DNA methylation has been identified to be involved in the pathophysiology of essential hypertension in adults; however its role in the pathogenesis of prehypertension and essential hypertension in young adults is not known. Furthermore, although some genes of interest have been studied, these pathways are yet to be fully explored. Based on the current literature, there were several unexplored areas that need to be addressed; 1) There is very limited literature on epigenetic studies in hypertensive young adults. 2) Available DNA methylation studies did not compare across three blood pressure status, i.e., normotension, prehypertension and hypertension, despite prehypertension as an established predisease position. 3) The different approaches and methods utilised to qualify or quantify DNA methylation results in varying, incomparable outcomes. 4) Furthermore, the studies were not extended to gene expression study; hence, the effect of differential DNA methylation on gene expression is largely unknown.